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(54) Title: BMP-12, BMP-13 AND TENDON-INDUCING COMPOSITIONS THEREOF

(57) Abstract

Bone morphogenetic proteins BMP-12 and BMP-13 have been cloned. Compositions of these proteins with tendon/ligament-like tissue inducing activity are disclosed. The compositions are useful in the treatment of tendinitis and tendon or ligament defects and in related tissue repair.

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TITLE OF THE INVENTION

BMP-12, BMP-13 AND TENDON-INDUCING COMPOSITIONS THEREOF

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RELATED APPLICATIONS

The present invention is a continuation-in-part of application serial number 08/217,780, filed March 25, 1994, 08/164,103, filed on December 7, 1993 and 08\333,576, filed on November 2, 1994.

FIELD OF THE INVENTION

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The present invention relates to a novel family of purified proteins, and compositions containing such proteins, which compositions are useful for the induction of tendon/ligament-like tissue formation, wound healing and ligament and other tissue repair. These proteins may also be used in compositions for augmenting the activity of bone morphogenetic proteins.

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BACKGROUND OF THE INVENTION

The search for the molecule or molecules responsible for formation of bone, cartilage, tendon and other tissues present in bone and other tissue extracts has led to the discovery of a novel set of molecules called the Bone Morphogenetic Proteins (BMPs). The structures of several proteins, designated BMP-1 through BMP-11, have previously been elucidated. The unique inductive activities of these proteins, along with their presence in bone, suggests that they are important regulators of bone repair processes, and may be involved in the normal maintenance of bone tissue. There is a need to identify additional proteins which play a role in forming other vital tissues. The present invention relates to the identification of a family of proteins, which have tendon/ligament-like tissue inducing activity, and which are useful in compositions for the induction of tendon/ligament-like tissue formation and repair.

SUMMARY OF THE INVENTION

In one embodiment, the present invention comprises DNA molecules encoding a tendon/ligament-like inducing protein which the inventors have named V1-1. This novel protein is now called BMP-12. The present invention also includes DNA molecules encoding BMP-12 related proteins.

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BMP-12 related proteins are a subset of the BMP/TGF-β/Vg-1 family of proteins, including BMP-12 and VL-1, which are defined as tendon/ligament-like tissue inducing proteins encoded by DNA sequences which are cloned and identified, e.g., using PCR, using BMP-12 specific primers, such as primers #6 and #7 described below, with reduced stringency conditions. It is preferred that the DNA sequences encoding BMP-12 related proteins share at least about 80% homology at the amino acid level from amino acids with amino acids #3 to #103 of SEQ ID NO:1.

The DNA molecules preferably have a DNA sequence encoding the BMP-12 protein, the sequence of which is provided in SEQ ID NO:1, or a BMP-12 related protein as further described herein. Both the BMP-12 protein and BMP-12 related proteins are characterized by the ability to induce the formation of tendon/ligament-like tissue in the assay described in the examples.

The DNA molecules of the invention preferably comprise a DNA sequence, as described in SEQUENCE ID NO:1; more preferably nucleotides #496 to #882, #571 to #882 or #577 to #882 of SEQ ID NO:1; or DNA sequences which hybridize to the above under stringent hybridization conditions and encode a protein which exhibits the ability to form tendon/ligament-like tissue. The DNA molecules of the invention may also comprise a DNA sequence as described in SEQ ID NO:25; more preferably nucleotides #604 or #658 to #964 of SEQ ID NO:25.

The DNA molecules of the invention also include DNA molecules comprising a DNA sequence encoding a BMP-12 related protein with the amino acid sequence shown in SEQ ID NO:2 or SEQ ID NO:26, as well as naturally occurring allelic sequences and equivalent degenerative codon sequences of SEQ ID NO:2 or SEQ ID NO:26. Preferably, the DNA sequence of the present invention encodes amino acids #-25 to # 104, #1 to # 104 or #3 to #103 of SEQ ID NO:2; or amino acids #1 to #120 or #19 to #120 of SEQ ID NO:26. The DNA sequence may comprise, in a 5' to 3' direction, nucleotides encoding a propeptide, and nucleotides encoding for amino acids #-25 to #104, #1 to #104 or #3 to #103 of SEQ ID NO:2; or amino acids #1 to #120 or #19 to #120 of SEQ ID NO:26. The propeptide useful in the above embodiment is preferably selected from the group consisting of native BMP-12 propeptide and a protein propeptide from a different member of the TGF-B

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superfamily or BMP family. The invention further comprises DNA sequences which hybridize to the above DNA sequences under stringent hybridization conditions and encode a BMP-12 related protein which exhibits the ability to induce formation of tendon/ligament-like tissue.

In other embodiments, the present invention comprises host cells and vectors which comprise a DNA molecule encoding the BMP-12 protein, or a BMP-12 related protein. The host cells and vectors may further comprise the coding sequence in operative association with an expression control sequence therefor.

In another embodiment, the present invention comprises a method for producing a purified BMP-12 related protein, said method comprising the steps of culturing a host cell transformed with the above DNA molecule or vector comprising a nucleotide sequence encoding a BMP-12 related protein; and (b) recovering and purifying said BMP-12 related protein from the culture medium. In a preferred embodiment, the method comprises (a) culturing a cell transformed with a DNA molecule comprising the nucleotide sequence from nucleotide #496, #571 or #577 to #879 or #882 as shown in SEQ ID NO:1; or the nucleotide sequence from #604 or #658 to #963 of SEQ ID NO:25; and

(b) recovering and purifying from said culture medium a protein comprising the amino acid sequence from amino acid #-25, #1 or #3 to amino acid #103 or #104 as shown in SEQ ID NO:2; or from amino acid #1 or #19 to amino acid #120 as shown in SEQ ID NO:26. The present invention also includes a purified protein produced by the above methods.

The present invention further comprises purified BMP-12 related protein characterized by the ability to induce the formation of tendon/ligament-like tissue. The BMP-12 related polypeptides preferably comprise an amino acid sequence as shown in SEQ ID NO:2. The polypeptide more preferably comprise amino acids #-25, #1 or #3 to #103 or #104 as set forth in SEQ ID NO:2; or amino acids #1 or #19 to #120 as set forth in SEQ ID NO:26. In a preferred embodiment, the purified polypeptide may be in the form of a dimer comprised of two subunits, each with the amino acid sequence of SEQ ID NO:2.

In another embodiment, the present invention comprises compositions comprising an effective amount of the above-described BMP-12 related proteins.

In the compositions, the protein may be admixed with a pharmaceutically acceptable vehicle.

The invention also includes methods for tendon/ligament-like tissue healing and tissue repair, for treating tendinitis, or other tendon or ligament defects, and for inducing tendon/ligament-like tissue formation in a patient in need of same, comprising administering to said patient an effective amount of the above composition.

Other embodiments include chimeric DNA molecules comprising a DNA sequence encoding a propeptide from a member of the TGF- β superfamily of proteins linked in correct reading frame to a DNA sequence encoding a BMP-12 related polypeptide. One suitable propeptide is the propeptide from BMP-2. The invention also includes heterodimeric protein molecules comprising one monomer having the amino acid sequence shown in SEQ ID NO:2, and one monomer having the amino acid sequence of another protein of the TGF- β subfamily.

Finally, the present invention comprises methods for inducing tendon/ligament-like tissue formation in a patient in need of same comprising administering to said patient an effective amount of a composition comprising a protein which exhibits the ability to induce formation of tendon/ligament-like tissue, said protein having an amino acid sequence shown in SEQ ID NO:2 or SEQ ID NO:4 or SEQ ID NO:26. The amino acid sequences are more preferably one of the following: (a) amino acids #-25, #1 or #3 to #103 or #104 of SEQ ID NO:2; (b) amino acids #1 or #19 to #119 or #120 of SEQ ID NO:4; (c) amino acids #1 or #19. to #119 or #120 of SEQ ID NO:26; (d) mutants and/or variants of (a), (b) or (c) which exhibit the ability to form tendon and/or ligament. In other embodiments of the above method, the protein is encoded by a DNA sequence of SEQ ID NO:1, SEO ID NO:3 or SEO ID NO:25, more preferably one of the following: (a) nucleotides #496, #571 or #577 to #879 or #882 of SEQ ID NO:1; (b) nucleotides #845 or #899 to #1201 or #1204 of SEQ ID NO:3; (c) nucleotides #605 or #659 to #961 or #964 of SEQ ID NO:25; and (d) sequences which hybridize to (a) or (b) under stringent hybridization conditions and encode a protein which exhibits the ability to form tendon/ligament-like tissue.

Description of the Sequences

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SEQ ID NO:1 is the nucleotide sequence encoding the human BMP-12.

SEQ ID NO:2 is the amino acid sequence comprising the mature human BMP-12 polypeptide.

SEQ ID NO:3 is the nucleotide sequence encoding the protein MP52.

SEQ ID NO:4 is the amino acid sequence comprising the mature MP52 polypeptide.

SEQ ID NO:5 is the nucleotide sequence of a specifically amplified portion of the human BMP-12 encoding sequence.

SEQ ID NO:6 is the amino acid sequence encoded by the nucleotide sequence of SEQ ID NO:5.

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SEQ ID NO:7 is the nucleotide sequence of a specifically amplified portion of the human VL-1 encoding sequence.

SEQ ID NO:8 is the amino acid sequence encoded by the nucleotide sequence of SEQ ID NO:7.

SEQ ID NO:9 is the nucleotide sequence of the plasmid pALV1-781, used for expression of BMP-12 in E. coli.

SEQ ID NO:10 is the nucleotide sequence of a fragment of the murine clone, mV1.

SEQ ID NO:11 is the amino acid sequence of a fragment of the murine protein encoded by mV1.

SEQ ID NO:12 is the nucleotide sequence of a fragment of the murine clone, mV2.

SEQ ID NO:13 is the amino acid sequence of a fragment of the murine protein encoded by mV2.

SEQ ID NO:14 is the nucleotide sequence of a fragment of the murine clone, mV9.

SEQ ID NO:15 is the amino acid sequence of a fragment of the murine protein encoded by mV9.

SEQ ID NO:16 is the amino acid sequence of a BMP/TGF- β /Vg-1 protein consensus sequence. The first Xaa represents either Gln or Asn; the second Xaa represents either Val or Ile.

SEQ ID NO:17 is the nucleotide sequence of oligonucleotide #1.

SEQ ID NO:18 is the amino acid sequence of a BMP/TGF- β /Vg-1 protein consensus sequence. The Xaa represents either Val or Leu.

SEQ ID NO:19 is the nucleotide sequence of oligonucleotide #2.

SEQ ID NO:20 is the nucleotide sequence of oligonucleotide #3.

SEQ ID NO:21 is the nucleotide sequence of oligonucleotide #4.

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SEO ID NO:22 is the nucleotide sequence of oligonucleotide #5

SEQ ID NO:23 is the nucleotide sequence of oligonucleotide #6.

SEO ID NO:24 is the nucleotide sequence of oligonucleotide #7.

SEQ ID NO:25 is the nucleotide sequence of the human VL-1 (BMP-13) encoding sequence.

SEQ ID NO:26 is the amino acid sequence encoded by the nucleotide sequence of SEQ ID NO:25.

SEQ ID NO:27 is the nucleotide sequence encoding a fusion of BMP-2 propertide and the mature coding sequence of BMP-12.

SEQ ID NO:28 is the amino acid sequence encoded by the nucleotide sequence of SEQ ID NO:27.

SEQ ID NO:29 is the nucleotide sequence encoding the murine mV1 protein. X01 is Val, Ala, Glu or Gly; X02 is Ser, Pro Thr or Ala; X03 is Ser or Arg; X04 is Leu, Pro, Gln or Arg; X05 is Cys or Trp; X06 is Val, Ala, Asp or Gly; X07 is Val, Ala, Glu or Gly; X08 is Gln, Lys or Glu.

SEQ ID NO:30 is the amino acid sequence encoded by the nucleotide sequence of SEQ ID NO:29. X01 through X08 are the same as in SEQ ID NO: 29.

SEQ ID NO:31 is the nucleotide sequence encoding the murine mV2 protein. X01 is Pro or Thr; X02 is Val.

SEQ ID NO:32 is the amino acid sequence encoded by the nucleotide sequence of SEQ ID NO:31. X01 and X02 are the same as in SEQ ID NO:31.

SEQ ID NO:33 is the nucleotide sequence encoding human BMP-12 protein.

SEQ ID NO:34 is the amino acid sequence encoded by the nucleotide sequence of SEQ ID NO:33.

30 SEQ ID NO:35 is the nucleotide sequence of oligonucleotide #8.

Brief Description of the Figures

Figure 1 is a comparison of the human BMP-12 and human MP52 sequences.

Detailed Description of the Invention

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The DNA sequences of the present invention are useful for producing proteins which induce the formation of tendon/ ligament-like tissue, as described further below. The DNA sequences of the present invention are further useful for isolating and cloning further DNA sequences encoding BMP-12 related proteins with similar activity. These BMP-12 related proteins may be homologues from other species, or may be related proteins within the same species.

Still, a further aspect of the invention are DNA sequences coding for expression of a tendon/ligament-like tissue inducing protein. Such sequences include the sequence of nucleotides in a 5' to 3' direction illustrated in SEQ ID NO:1 or SEQ ID NO:25, DNA sequences which, but for the degeneracy of the genetic code, are identical to the DNA sequence SEQ ID NO:1 or 25, and encode the protein of SEQ ID NO:2 or 26. Further included in the present invention are DNA sequences which hybridize under stringent conditions with the DNA sequence of SEQ ID NO:1 or 25 and encode a protein having the ability to induce the formation of tendon or ligament. Preferred DNA sequences include those which hybridize under stringent conditions as described in Maniatis et al, Molecular Cloning (A Laboratory Manual), Cold Spring Harbor Laboratory (1982), pages 387 to 389. Finally, allelic or other variations of the sequences of SEQ ID NO:1 or 25, whether such nucleotide changes result in changes in the peptide sequence or not, but where the peptide sequence still has tendon/ligament-like tissue inducing activity, are also included in the present invention.

The human BMP-12 DNA sequence (SEQ ID NO:1) and amino acid sequence (SEQ ID NO:2) are set forth in the Sequence Listings. Another protein that is useful for the compositions and methods of the present invention is VL-1. VL-1 is a BMP-12 related protein which was cloned using sequences from BMP-12. The inventors have now designated VL-1 as BMP-13. A partial DNA sequence of VL-1 (SEQ ID NO:7) and the encoded amino acid sequence (SEQ ID NO:8); as well as a DNA sequence encoding the mature VL-1 (SEQ ID NO:25) and the encoded amino acid sequence (SEQ ID NO:26) are set forth in the Sequence Listings. Although further descriptions are made with reference to the BMP-12 sequence of SEQ ID NO:1 and 2, it will be recognized that the invention includes similar modifications and

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improvements which may be made to other BMP-12 related sequences, such as the VL-1 sequence shown in SEQ ID NO:25 and 26.

The sequence of BMP-12 shown in SEQ ID NO. 1 includes the entire mature sequence and approximately 190 amino acids of the propeptide. The coding sequence of the mature human BMP-12 protein appears to begin at nucleotide #496 or #571 and continues through nucleotide #882 of SEQ ID NO:1. The first cysteine in the seven cysteine structure characteristic of TGF- β proteins begins at nucleotide #577. The last cysteine ends at #879. Thus, it is expected that DNA sequences encoding active BMP-12 species will comprise nucleotides #577 to #879 of SEQ ID NO:1.

It is expected that BMP-12, as expressed by mammalian cells such as CHO cells, exists as a heterogeneous population of active species of BMP-12 protein with varying N-termini. It is expected that all active species will contain the amino acid sequence beginning with the cysteine residue at amino acid #3 of SEQ ID NO:2 and continue through at least the cysteine residue at amino acid 103 or until the stop codon after amino acid 104. Other active species contain additional amino acid sequence in the N-terminal direction. As described further herein, the N-termini of active species produced by mammalian cells are expected to begin after the occurrence of a consensus cleavage site, encoding a peptide sequence Arg-X-X-Arg. Thus, it is expected that DNA sequences encoding active BMP-12 proteins will have a nucleotide sequence comprising the nucleotide sequence beginning at any of nucleotides #196, 199, 208, 217, 361, 388, 493, 496 or 571 to nucleotide #879 or 882 of SEQ ID NO:1.

The N-terminus of one active species of human BMP-12 has been experimentally determined by expression in E. coli to be as follows: [M]SRXSRKPLHVDF, wherein X designates an amino acid residue with no clear signal, which is consistent with a cysteine residue at that location. Thus, it appears that the N-terminus of this species of BMP-12 is at amino acid #1 of SEQ ID NO:1, and a DNA sequence encoding said species of BMP-12 would start at nucleotide #571 of SEQ ID NO:1. The apparent molecular weight of this species of human BMP-12 dimer was determined by SDS-PAGE to be approximately 20-22 kd on a

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Novex 16% tricine gel. The human BMP-12 protein exists as a clear, colorless solution in 0.1% trifluoroacetic acid.

As described earlier, BMP-12 related proteins are a subset of the BMP/TGF-β/Vg-1 family of proteins, including BMP-12 and VL-1, which can be defined as tendon/ligament-like tissue inducing proteins encoded by DNA sequences which can be cloned and identified, e.g., using PCR, using BMP-12 specific primers, such as primers #6 and #7 described below, with reduced stringency conditions. It is preferred that DNA sequences of the present invention share at least about 80% homology at the amino acid level from amino acids with the DNA encoding amino acids #3 to #103 of SEQ ID NO:1. For the purposes of the present invention, the term BMP-12 related proteins does not include the human MP52 protein. Using the sequence information of SEQ ID NO:1 and SEQ ID NO:3, and the comparison provided in Figure 1, it is within the skill of the art to design primers to the BMP-12 sequence which will allow for the cloning of genes encoding BMP-12 related proteins.

One example of the BMP-12-related proteins of the present invention is VL-1, presently referred to as BMP-13. The sequence of the full mature BMP-13 sequence and at least a part of the propeptide of BMP-13 is given in SEQ ID NO:25. Like BMP-12, it is expected that BMP-13, as expressed by mammalian cells such as CHO cells, exists as a heterogeneous population of active species of BMP-13 protein with varying N-termini. It is expected that all active species will contain the amino acid sequence beginning with the cysteine residue at amino acid #19 of SEQ ID NO:26 and continue through at least the cysteine residue at amino acid 119 or until the stop codon after amino acid 120. Other active species contain additional amino acid sequence in the N-terminal direction. As described further herein, the N-termini of active species produced by mammalian cells are expected to begin after the occurrence of a consensus cleavage site, encoding a peptide sequence Arg-X-X-Arg. Thus, it is expected that DNA sequences encoding active BMP-13 proteins will have a nucleotide sequence comprising the nucleotide sequence beginning at any of nucleotides #410, 458, 602, 605 or 659, to nucleotide #961 or 964 of SEQ ID NO:25.

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In order to produce the purified tendon/ligament-like tissue inducing proteins useful for the present invention, a method is employed comprising culturing a host cell transformed with a DNA sequence comprising a suitable coding sequence, particularly the DNA coding sequence from nucleotide #496, #571 or #577 to #879 or #882 of SEQ ID NO:1; and recovering and purifying from the culture medium a protein which contains the amino acid sequence or a substantially homologous sequence as represented by amino acids #-25, #1 or #3 to #103 or #104 of SEQ ID NO:2. In another embodiment, the method employed comprises culturing a host cell transformed with a DNA sequence comprising a suitable coding sequence, particularly the DNA coding sequence from nucleotide #605 or #659 to #961 or #964 of SEQ ID NO:25; and recovering and purifying from the culture medium a protein which contains the amino acid sequence or a substantially homologous sequence as represented by amino acids #1 or #19 to #119 or #120 of SEQ ID NO:26.

The human MP52 DNA is described in WO93/16099, the disclosure of which is incorporated herein by reference. However, this document does not disclose the ability of the protein to form tendon/ligament-like tissue, or its use in compositions for induction of tendon/ligament-like tissue. Human MP52 was originally isolated using RNA from human embryo tissue. The human MP52 nucleotide sequence (SEQ ID NO:3) and the encoded amino acid sequences (SEQ ID NO:4) are set forth in the Sequence Listings herein. The MP52 protein appears to begin at nucleotide #845 of SEQ ID NO:3 and continues through nucleotide #1204 of SEQ ID NO:3. The first cysteine of the seven cysteine structure characteristic of TGF-β proteins begins at nucleotide #899. The last cysteine ends at #1201. Other active species of MP52 protein may have additional nucleotides at the N-terminal direction from nucleotide #845 of SEQ ID NO:3.

Purified human MP52 proteins of the present invention may be produced by culturing a host cell transformed with a DNA sequence comprising the DNA coding sequence of SEQ ID NO:3 from nucleotide #845 to #1204, and recovering and purifying from the culture medium a protein which contains the amino acid sequence or a substantially homologous sequence as represented by amino acids #1 to #120 of SEQ ID NO:4. It is also expected that the amino acid sequence from amino acids

> #17 or #19 to #119 or #120 of SEQ ID NO:4 will retain activity. Thus, the DNA sequence from nucleotides #845, #893 or #899 to #1201 or #1204 are expected to encode active proteins.

> For expression of the protein in mammalian host cells, the host cell is transformed with a coding sequence encoding a propeptide suitable for the secretion of proteins by the host cell is linked in proper reading frame to the coding sequence for the mature protein. For example, see United States Patent 5,168,050, the disclosure of which is hereby incorporated by reference, in which a DNA encoding a precursor portion of a mammalian protein other than BMP-2 is fused to the DNA encoding a mature BMP-2 protein. Thus, the present invention includes chimeric DNA molecules comprising a DNA sequence encoding a propertide from a member of the TGF-\(\theta \) superfamily of proteins, is linked in correct reading frame to a DNA sequence encoding a tendon/ligament-like tissue inducing polypeptide. The term "chimeric" is used to signify that the propeptide originates from a different polypeptide than the encoded mature polypeptide. Of course, the host cell may be transformed with a DNA sequence coding sequence encoding the native propeptide linked in correct reading frame to a coding sequence encoding the mature protein shown in SEQ ID NO:2, SEQ ID NO:4, or SEQ ID NO:26. The full sequence of the native propeptide may be determined through methods known in the art using the sequences disclosed in SEQ ID NO:1, SEQ ID NO:3, or SEQ ID NO:25 to design a suitable probe for identifying and isolating the entire clone.

> The present invention also encompasses the novel DNA sequences, free of association with DNA sequences encoding other proteinaceous materials, and coding for expression of tendon/ligament-like tissue inducing proteins. sequences include those depicted in SEQ ID NO:1 in a 5' to 3' direction and those sequences which hybridize thereto under stringent hybridization conditions [for example, 0.1X SSC, 0.1% SDS at 65°C; see, T. Maniatis et al, Molecular Cloning (A Laboratory Manual), Cold Spring Harbor Laboratory (1982), pages 387 to 389] and encode a protein having tendon/ligament-like tissue inducing activity.

> Similarly, DNA sequences which code for proteins coded for by the sequences of SEO ID NO:1 or SEO ID NO:25, or proteins which comprise the amino acid sequence of SEQ ID NO:2 or SEQ ID NO:26, but which differ in codon

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sequence due to the degeneracies of the genetic code or allelic variations (naturally-occurring base changes in the species population which may or may not result in an amino acid change) also encode the tendon/ligament-like tissue inducing proteins described herein. Variations in the DNA sequences of SEQ ID NO:1 or SEQ ID NO:25 which are caused by point mutations or by induced modifications (including insertion, deletion, and substitution) to enhance the activity, half-life or production of the polypeptides encoded are also encompassed in the invention.

Another aspect of the present invention provides a novel method for producing tendon/ligament-like tissue inducing proteins. The method of the present invention involves culturing a suitable cell line, which has been transformed with a DNA sequence encoding a protein of the invention, under the control of known regulatory sequences. The transformed host cells are cultured and the proteins recovered and purified from the culture medium. The purified proteins are substantially free from other proteins with which they are co-produced as well as from other contaminants.

Suitable cells or cell lines may be mammalian cells, such as Chinese hamster ovary cells (CHO). As described above, expression of protein in mammalian cells requires an appropriate propeptide to assure secretion of the protein. The selection of suitable mammalian host cells and methods for transformation, culture, amplification, screening, product production and purification are known in the art. See, e.g., Gething and Sambrook, Nature, 293:620-625 (1981), or alternatively, Kaufman et al, Mol. Cell. Biol., 5(7):1750-1759 (1985) or Howley et al, U.S. Patent 4,419,446. Another suitable mammalian cell line, which is described in the accompanying examples, is the monkey COS-1 cell line. The mammalian cell CV-1 may also be suitable.

Bacterial cells may also be suitable hosts. For example, the various strains of \underline{E} . $\underline{\operatorname{coli}}$ (e.g., HB101, MC1061) are well-known as host cells in the field of biotechnology. Various strains of \underline{B} . $\underline{\operatorname{subtilis}}$, $\underline{\operatorname{Pseudomonas}}$, other bacilli and the like may also be employed in this method. For expression of the protein in bacterial cells. DNA encoding a propeptide is not necessary.

Bacterial expression of mammalian proteins, including members of the TGF- β family is known to produce the proteins in a non-glycosylated form, and in the form

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of insoluble pellets, known as inclusion bodies. Techniques have been described in the art for solubilizing these inclusion bodies, denaturing the protein using a chaotropic agent, and refolding the protein sufficiently correctly to allow for their production in a soluble form. For example, see EP 0433225, the disclosure of which is hereby incorporated by reference.

Alternatively, methods have been devised which circumvent inclusion body formation, such as expression of gene fusion proteins, wherein the desired protein is expressed as a fusion protein with a fusion partner. The fusion protein is later subjected to cleavage to produce the desired protein. One example of such a gene fusion expression system for <u>E. coli</u> is based on use of the <u>E. coli</u> thioredoxin gene as a fusion partner, LaVallie et al., <u>Bio/Technology</u>, 11:187-193 (1993), the disclosure of which is hereby incorporated by reference.

Many strains of yeast cells known to those skilled in the art may also be available as host cells for expression of the polypeptides of the present invention. Additionally, where desired, insect cells may be utilized as host cells in the method of the present invention. See, e.g. Miller et al, <u>Genetic Engineering</u>, <u>8</u>:277-298 (Plenum Press 1986) and references cited therein.

Another aspect of the present invention provides vectors for use in the method of expression of these tendon/ligament-like tissue inducing proteins. Preferably the vectors contain the full novel DNA sequences described above which encode the novel factors of the invention. Additionally, the vectors contain appropriate expression control sequences permitting expression of the protein sequences. Alternatively, vectors incorporating modified sequences as described above are also embodiments of the present invention. Additionally, the sequence of SEQ ID NO:1 or SEQ ID NO:3 or SEQ ID NO:25 could be manipulated to express a mature protein by deleting propeptide sequences and replacing them with sequences encoding the complete propeptides of BMP proteins or members of the TGF- β superfamily. Thus, the present invention includes chimeric DNA molecules encoding a propeptide from a member of the TGF- β superfamily linked in correct reading frame to a DNA sequence encoding a protein having the amino acid sequence of SEQ ID NO:2 or SEQ ID NO:4 or SEQ ID NO:26. The vectors may be employed in the method of transforming cell lines and contain selected regulatory sequences in operative

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association with the DNA coding sequences of the invention which are capable of directing the replication and expression thereof in selected host cells. Regulatory sequences for such vectors are known to those skilled in the art and may be selected depending upon the host cells. Such selection is routine and does not form part of the present invention.

A protein of the present invention, which induces tendon/ligament-like tissue or other tissue formation in circumstances where such tissue is not normally formed, has application in the healing of tendon or ligament tears, deformities and other tendon or ligament defects in humans and other animals. Such a preparation employing a tendon/ligament-like tissue inducing protein may have prophylactic use in preventing damage to tendon or ligament tissue, as well as use in the improved fixation of tendon or ligament to bone or other tissues, and in repairing defects to tendon or ligament tissue. De novo tendon/ligament-like tissue formation induced by a composition of the present invention contributes to the repair of congenital, trauma induced, or other tendon or ligament defects of other origin, and is also useful in cosmetic plastic surgery for attachment or repair of tendons or ligaments. The compositions of the invention may also be useful in the treatment of tendinitis, carpal tunnel syndrome and other tendon or ligament defects. The compositions of the present invention can also be used in other indications wherein it is desirable to heal or regenerate tendon and/or ligament tissue. Such indications include, without limitation, regeneration or repair of injuries to the periodontal ligament, such as occurs in tendonitis, and regeneration or repair of the tendon-to-bone attachment. The compositions of the present invention may provide an environment to attract tendon- or ligament-forming cells, stimulate growth of tendon- or ligament-forming cells or induce differentiation of progenitors of tendon- or ligament-forming cells.

The BMP-12 related proteins may be recovered from the culture medium and purified by isolating them from other proteinaceous materials from which they are co-produced and from other contaminants present. The proteins of the present invention are capable of inducing the formation of tendon/ligament-like tissue. These proteins may be further characterized by the ability to demonstrate tendon/ligament-like tissue formation activity in the rat ectopic implant assay

described below. It is contemplated that these proteins may have ability to induce the formation of other types of tissue, such as ligaments, as well.

The tendon/ligament-like tissue inducing proteins provided herein also include factors encoded by the sequences similar to those of SEQ ID NO:1 or SEQ ID NO:25, but into which modifications are naturally provided (e.g. allelic variations in the nucleotide sequence which may result in amino acid changes in the polypeptide) or deliberately engineered. For example, synthetic polypeptides may wholly or partially duplicate continuous sequences of the amino acid residues of SEQ ID NO:2. These sequences, by virtue of sharing primary, secondary, or tertiary structural and conformational characteristics with tendon/ligament-like tissue growth factor polypeptides of SEQ ID NO:2 may possess tendon/ligament-like or other tissue growth factor biological properties in common therewith. Thus, they may be employed as biologically active substitutes for naturally-occurring tendon/ligament-like tissue inducing polypeptides in therapeutic compositions and processes.

Other specific mutations of the sequences of tendon/ligament-like tissue inducing proteins described herein involve modifications of glycosylation sites. These modifications may involve O-linked or N-linked glycosylation sites. For instance, the absence of glycosylation or only partial glycosylation results from amino acid substitution or deletion at asparagine-linked glycosylation recognition sites. The asparagine-linked glycosylation recognition sites comprise tripeptide sequences which are specifically recognized by appropriate cellular glycosylation enzymes. These tripeptide sequences may be asparagine-X-threonine, asparagine-X-serine or asparagine-X-cysteine, where X is usually any amino acid except proline. A variety of amino acid substitutions or deletions at one or both of the first or third amino acid positions of a glycosylation recognition site (and/or amino acid deletion at the second position) results in non-glycosylation at the modified tripeptide sequence. Additionally, bacterial expression of protein will also result in production of a non-glycosylated protein, even if the glycosylation sites are left unmodified.

The compositions of the present invention comprise a purified BMP-12 related protein which may be produced by culturing a cell transformed with the DNA sequence of SEQ ID NO:1 or SEQ ID NO:25 and recovering and purifying protein having the amino acid sequence of SEQ ID NO:2 or SEQ ID NO:26 from the culture

medium. The purified expressed protein is substantially free from other proteinaceous materials with which it is co-produced, as well as from other contaminants. The recovered purified protein is contemplated to exhibit tendon/ligament-like tissue formation activity, and other tissue growth activity, such as ligament regeneration. The proteins of the invention may be further characterized by the ability to demonstrate tendon/ligament-like tissue formation activity in the rat assay described below.

The compositions for inducing tendon/ligament-like tissue formation of the present invention may comprise an effective amount of a tendon/ligament-like tissue inducing protein, wherein said protein comprises the amino acid sequence of SEQ ID NO:2, preferably amino acids #-25, #1 or #3 to #103 or #104 of SEQ ID NO:2; or amino acids #1 or #19 to #120 of SEQ ID NO:26; as well as mutants and/or variants of SEQ ID NO:2 or SEQ ID NO:26, which exhibit the ability to form tendon and/or ligament like tissue.

Compositions of the present invention may further comprise additional proteins, such as additional members of the TGF- β superfamily of proteins, such as activins. Another aspect of the invention provides pharmaceutical compositions containing a therapeutically effective amount of a tendon/ligament-inducing protein, such as BMP-12 or VL-1, in a pharmaceutically acceptable vehicle or carrier. These compositions may be used to induce the formation of tendon/ligament-like tissue or other tissue. It is contemplated that such compositions may also be used for tendon and ligament repair, wound healing and other tissue repair, such as skin repair. It is further contemplated that proteins of the invention may increase neuronal survival and therefore be useful in transplantation and treatment of conditions exhibiting a decrease in neuronal survival. Compositions of the invention may further include at least one other therapeutically useful agent, such as the BMP proteins BMP-1, BMP-2, BMP-3, BMP-4, BMP-5, BMP-6 and BMP-7, disclosed for instance in United States Patents 5,108,922; 5,013,649; 5,116,738; 5,106,748; 5,187,076; and 5,141,905; BMP-8, disclosed in PCT publication WO91/18098; BMP-9, disclosed in PCT publication WO93/00432; and BMP-10 or BMP-11, disclosed in co-pending patent applications, serial number 08/061,695 and 08/061,464, filed on May 12,

> 1993. The disclosure of the above documents are hereby incorporated by reference herein.

> The compositions of the invention may comprise, in addition to a tendon/ligament-inducing protein such as BMP-12 or VL-1 (BMP-13), other therapeutically useful agents including MP52, epidermal growth factor (EGF), fibroblast growth factor (FGF), platelet derived growth factor (PDGF), transforming growth factors (TGF- α and TGF- β), and fibroblast growth factor-4 (FGF-4), parathyroid hormone (PTH), leukemia inhibitory factor (LIF/HILDA/DIA), insulinlike growth factors (IGF-I and IGF-II). Portions of these agents may also be used in compositions of the present invention. For example, a composition comprising both BMP-2 and BMP-12 implanted together gives rise to both bone and tendon/ligament-like tissue. Such a composition may be useful for treating defects of the embryonic joint where tendon, ligaments, and bone form simultaneously at contiguous anatomical locations, and may be useful for regenerating tissue at the site of tendon attachment to bone. It is contemplated that the compositions of the invention may also be used in wound healing, such as skin healing and related tissue repair. The types of wounds include, but are not limited to burns, incisions and ulcers. (See, e.g. PCT Publication WO84/01106 for discussion of wound healing and related tissue repair).

> It is expected that the proteins of the invention may act in concert with or perhaps synergistically with other related proteins and growth factors. Further therapeutic methods and compositions of the invention therefore comprise a therapeutic amount of at least one protein of the invention with a therapeutic amount of at least one of the BMP proteins described above. Such compositions may comprise separate molecules of the BMP proteins or heteromolecules comprised of different BMP moieties. For example, a method and composition of the invention may comprise a disulfide linked dimer comprising a BMP-12 related protein subunit and a subunit from one of the "BMP" proteins described above. Thus, the present invention includes compositions comprising a purified BMP-12 related polypeptide which is a heterodimer wherein one subunit comprises the amino acid sequence from amino acid #1 to amino acid #104 of SEQ ID NO:2, and one subunit comprises an amino acid sequence for a bone morphogenetic protein selected from the group

consisting of BMP-1, BMP-2, BMP-3, BMP-4, BMP-5, BMP-6, BMP-7, BMP-8, BMP-9, BMP-10 and BMP-11. A further embodiment may comprise a heterodimer of disulfide bonded tendon/ligament-like tissue inducing moieties such as BMP-12, VL-1 (BMP-13) or MP52. For example the heterodimer may comprise one subunit comprising an amino acid sequence from #1 to # 104 of SEQ ID NO:2 and the other subunit may comprise an amino acid sequence from #1 to #120 of SEQ ID NO:4 or #1 to #120 of SEQ ID NO:26. Further, compositions of the present invention may be combined with other agents beneficial to the treatment of the defect, wound, or tissue in question.

The preparation and formulation of such physiologically acceptable protein compositions, having due regard to pH, isotonicity, stability and the like, is within the skill of the art. The therapeutic compositions are also presently valuable for veterinary applications due to the lack of species specificity in $TGF-\beta$ proteins. Particularly domestic animals and thoroughbred horses in addition to humans are desired patients for such treatment with the compositions of the present invention.

The therapeutic method includes administering the composition topically, systemically, or locally as an implant or device. When administered, the therapeutic composition for use in this invention is, of course, in a pyrogen-free, physiologically acceptable form. Further, the composition may desirably be encapsulated or injected in a viscous form for delivery to the site of tissue damage. Topical administration may be suitable for wound healing and tissue repair. Therapeutically useful agents other than the proteins which may also optionally be included in the composition as described above, may alternatively or additionally, be administered simultaneously or sequentially with the composition in the methods of the invention.

The compositions may also include an appropriate matrix and/or sequestering agent as a carrier. For instance, the matrix may support the composition or provide a surface for tendon/ligament-like tissue formation and/or other tissue formation. The matrix may provide slow release of the protein and/or the appropriate environment for presentation thereof. The sequestering agent may be a substance which aids in ease of administration through injection or other means, or may slow the migration of protein from the site of application.

The choice of a carrier material is based on biocompatibility, biodegradability, mechanical properties, cosmetic appearance and interface properties. The particular application of the compositions will define the appropriate formulation. Potential matrices for the compositions may be biodegradable and chemically defined. Further matrices are comprised of pure proteins or extracellular matrix components. Other potential matrices are nonbiodegradable and chemically defined. Preferred matrices include collagen-based materials, such as Helistat sponge (Integra LifeSciences, Plainsboro, N.J.), or collagen in an injectable form, as well as sequestering agents, which may also be biodegradable, and which may include alkylcellulosic materials.

Another preferred class of carrier are porous particulate polymer matrices, including polymers of poly(lactic acid), poly(glycolic acid) and copolymers of lactic acid and glycolic acid. These matrices may also include a sequestering agent. Suitable polymer matrices are described, for example, in WO 93/00050, the disclosure of which is incorporated herein by reference.

A preferred family of sequestering agents is cellulosic materials such as alkylcelluloses (including hydroxyalkylcelluloses), including methylcellulose, ethylcellulose, hydroxyethylcellulose, hydroxypropylcellulose, hydroxypropylmethylcellulose, and carboxymethylcellulose, the most preferred being cationic salts of carboxymethylcellulose (CMC). Other preferred sequestering agents include hyaluronic acid, sodium alginate, poly(ethylene glycol), polyoxyethylene oxide, carboxyvinyl polymer and poly(vinyl alcohol). The amount of sequestering agent useful herein is 0.5-20 wt%, preferably 1-10 wt% based on total formulation weight, which represents the amount necessary to prevent desorbtion of the protein from the polymer matrix and to provide appropriate handling of the composition, yet not so much that the progenitor cells are prevented from infiltrating the matrix, thereby providing the protein the opportunity to assist the activity of the progenitor cells.

Additional optional components useful in the practice of the subject application include, e.g. cryogenic protectors such as mannitol, sucrose, lactose, glucose, or glycine (to protect the protein from degradation during lyophilization), antimicrobial preservatives such as methyl and propyl parabens and benzyl alcohol:

antioxidants such as EDTA, citrate and BHT (butylated hydroxytoluene); and surfactants such as poly(sorbates) and poly(oxyethylenes); etc.

As described above, the compositions of the invention may be employed in methods for treating a number of tendon defects, such as the regeneration of tendon/ligament-like tissue in areas of tendon or ligament damage, to assist in repair of tears of tendon tissue, ligaments, and various other types of tissue defects or wounds. These methods, according to the invention, entail administering to a patient needing such tendon/ligament-like tissue or other tissue repair, a composition comprising an effective amount of a tendon/ligament-like tissue inducing protein, such as described in SEQ ID NO:2, SEQ ID NO:4 and/or SEQ ID NO:26. These methods may also entail the administration of a tendon/ligament-like tissue inducing protein in conjunction with at least one of the BMP proteins described above.

In another embodiment, the methods may entail administration of a heterodimeric protein in which one of the monomers is a tendon/ligament-like tissue inducing polypeptide, such as BMP-12, VL-1 (BMP-13) or MP52, and the second monomer is a member of the TGF- β superfamily of growth factors. In addition, these methods may also include the administration of a tendon/ligament-like tissue inducing protein with other growth factors including EGF, FGF, TGF- α , TGF- β , and IGF.

Thus, a further aspect of the invention is a therapeutic method and composition for repairing tendon/ligament-like tissue, for repairing tendon or ligament as well as treating tendinitis and other conditions related to tendon or ligament defects. Such compositions comprise a therapeutically effective amount of one or more tendon/ligament-like tissue inducing proteins, such as BMP-12, a BMP-12 related protein, or MP52, in admixture with a pharmaceutically acceptable vehicle, carrier or matrix.

The dosage regimen will be determined by the attending physician considering various factors which modify the action of the composition, e.g., amount of tendon or ligament tissue desired to be formed, the site of tendon or ligament damage, the condition of the damaged tendon or ligament, the size of a wound, type of damaged tissue, the patient's age, sex, and diet, the severity of any infection, time of administration and other clinical factors. The dosage may vary with the type of

matrix used in the reconstitution and the types of additional proteins in the composition. The addition of other known growth factors, such as IGF-I (insulin like growth factor I), to the final composition, may also affect the dosage.

Progress can be monitored by periodic assessment of tendon/ligament-like tissue formation, or tendon or ligament growth and/or repair. The progress can be monitored by methods known in the art, for example, X-rays, arthroscopy, histomorphometric determinations and tetracycline labeling.

The following examples illustrate practice of the present invention in recovering and characterizing human tendon/ligament-like tissue inducing protein and employing them to recover the other tendon/ligament-like tissue inducing proteins, obtaining the human proteins, expressing the proteins via recombinant techniques, and demonstration of the ability of the compositions of the present invention to form tendon/ligament-like tissue in an in vivo model. Although the examples demonstrate the invention with respect to BMP-12, with minor modifications within the skill of the art, the same results are believed to be attainable with MP52 and VL-1.

EXAMPLE 1

Isolation of DNA

DNA sequences encoding BMP-12 and BMP-12 related proteins may be isolated by various techniques known to those skilled in the art. As described below, oligonucleotide primers may be designed on the basis of amino acid sequences present in other BMP proteins, Vg-1 related proteins and other proteins of the TGF- β superfamily. Regions containing amino acid sequences which are highly conserved within the BMP family of proteins and within other members of the TGF- β superfamily of proteins can be identified and consensus amino acid sequences of these highly conserved regions can be constructed based on the similarity of the corresponding regions of individual BMP/TGF- β /Vg-1 proteins. An example of such a consensus amino acid sequence is indicated below.

Consensus amino acid sequence (1):

Trp-Gln/Asn-Asp-Trp-Ile-Val/Ile-Ala (SEQ ID NO:16)

Where X/Y indicates that either amino acid residue may appear at that position.

The following oligonucleotide is designed on the basis of the above identified consensus amino acid sequence (1):

#1: CGGATCCTGGVANGAYTGGATHRTNGC (SEQ ID NO:17)

This oligonucleotide sequence is synthesized on an automated DNA synthesizer. The standard nucleotide symbols in the above identified oligonucleotide primer are as follows: A,adenosine; C,cytosine; G,guanine; T,thymine; N,adenosine or cytosine or guanine or thymine; R,adenosine or cytosine or thymine; H,adenosine or cytosine or thymine; V,adenosine or cytosine or guanine; D,adenosine or guanine or thymine.

The first seven nucleotides of oligonucleotide #1 (underlined) contain the recognition sequence for the restriction endonuclease BamHI in order to facilitate the manipulation of a specifically amplified DNA sequence encoding the BMP-12 protein and are thus not derived from the consensus amino acid sequence (1) presented above.

A second consensus amino acid sequence is derived from another highly conserved region of BMP/TGF- β /Vg-1 proteins as described below:

His-Ala-Ile-Val/Leu-Gln-Thr (SEQ ID NO:18)

The following oligonucleotide is designed on the basis of the above identified consensus amino acid sequence (2):

#2: TTTCTAGAARNGTYTGNACDATNGCRTG (SEQ ID NO:19)

This oligonucleotide sequence is synthesized on an automated DNA synthesizer. The same nucleotide symbols are used as described above.

The first seven nucleotides of oligonucleotide #1 (underlined) contain the recognition sequence for the restriction endonuclease XbaI in order to facilitate the manipulation of a specifically amplified DNA sequence encoding the BMP-12 protein and are thus not derived from the consensus amino acid sequence (2) presented above.

It is contemplated that the BMP-12 protein of the invention and other BMP/TGF-β/Vg-1 related proteins may contain amino acid sequences similar to the consensus amino acid sequences described above and that the location of those sequences within a BMP-12 protein or other novel related proteins would correspond to the relative locations in the proteins from which they were derived. It is further

contemplated that this positional information derived from the structure of other BMP/TGF- β /Vg-1 proteins and the oligonucleotide sequences #1 and #2 which have been derived from consensus amino acid sequences (1) and (2), respectively, could be utilized to specifically amplify DNA sequences encoding the corresponding amino acids of a BMP-12 protein or other BMP/TGF- β /Vg-1 related proteins.

Based on the knowledge of the gene structures of BMP/TGF-β/Vg-1 proteins it is further contemplated that human genomic DNA can be used as a template to perform specific amplification reactions which would result in the identification of BMP-12 BMP/TGF-β/Vg-1 (BMP-12 related protein) encoding sequences. Such specific amplification reactions of a human genomic DNA template could be initiated with the use of oligonucleotide primers #1 and #2 described earlier. Oligonucleotides #1 and #2 identified above are utilized as primers to allow the specific amplification of a specific nucleotide sequence from human genomic DNA. The amplification reaction is performed as follows:

Human genomic DNA (source: peripheral blood lymphocytes), provided by Ken Jacobs of Genetics Institute, is sheared by repeated passage through a 25 gauge needle, denatured at 100°C for 5 minutes and then chilled on ice before adding to a reaction mixture containing 200 μM each deoxynucleotide triphosphates (dATP, dGTP, dCTP and dTTP), 10 mM Tris-HCl pH 8.3, 50 mM KCl, 1.5 mM MgCl₂, 0.001% gelatin, 1.25 units Taq DNA polymerase, 100 pM oligonucleotide #1 and 100 pM oligonucleotide #2. This reaction mixture is incubated at 94°C for two minutes and then subjected to thermal cycling in the following manner: 1 minute at 94°C, 1 minute at 40°C, 1 minute at 72°C for three cycles; then 1 minute at 94°C, 1 minute at 55°C, 1 minute at 72°C for thirty-seven cycles, followed by a 10 minute incubation at 72°C.

The DNA which is specifically amplified by this reaction is ethanol precipitated, digested with the restriction endonucleases BamHI and XbaI and subjected to agarose gel electrophoresis. A region of the gel, corresponding to the predicted size of the BMP-12 or other BMP/TGF-β/Vg-1 encoding DNA fragment, is excised and the specifically amplified DNA fragments contained therein are electroeluted and subcloned into the plasmid vector pGEM-3 between the XbaI and BamHI sites of the polylinker. DNA sequence analysis of one of the resulting BMP-

12 related subclones indicates the specifically amplified DNA sequence product contained therein encodes a portion of the BMP-12 protein of the invention.

The DNA sequence (SEQ ID NO:5) and derived amino acid sequence (SEQ ID NO:6) of this specifically amplified DNA fragment of BMP-12 are shown in the SEQUENCE Listings.

Nucleotides #1-#26 of SEQ ID NO:5 comprise a portion of oligonucleotide #1 and nucleotides #103 - #128 comprise a portion of the reverse compliment of oligonucleotide #2 utilized to perform the specific amplification reaction. Due to the function of oligonucleotides #1 and #2 in initiating the amplification reaction, they may not correspond exactly to the actual sequence encoding a BMP-12 protein and are therefore not translated in the corresponding amino acid derivation (SEQ ID NO:6).

DNA sequence analysis of another subclone indicates that the specifically amplified DNA product contained therein encodes a portion of another BMP/TGF- β /Vg-1 (BMP-12 related) protein of the invention named VL-1.

The DNA sequence (SEQ ID NO:7) and derived amino acid sequence (SEQ ID NO:8) of this specifically amplified DNA fragment are shown in the Sequence Listings.

Nucleotides #1 - #26 of SEQ ID NO:7 comprise a portion of oligonucleotide #1 and nucleotides #103 - #128 comprise a portion of the reverse compliment of oligonucleotide #2 utilized to perform the specific amplification reaction. Due to the function of oligonucleotides #1 and #2 in initiating the amplification reaction, they may not correspond exactly to the actual sequence encoding a VL-1 protein of the invention and are therefore not translated in the corresponding amino acid derivation (SEQ ID NO:8).

The following oligonucleotide probe is designed on the basis of the specifically amplified BMP-12 human DNA sequence set forth above (SEQ ID NO:5) and synthesized on an automated DNA synthesizer:

#3: CCACTGCGAGGCCTTTGCGACTTCCCTTTGCGTTCGCAC (SEQ ID NO:20)

This oligonucleotide probe is radioactively labeled with ^{32}P and employed to screen a human genomic library constructed in the vector λFIX (Stratagene catalog

#944201). 500,000 recombinants of the human genomic library are plated at a density of approximately 10,000 recombinants per plate on 50 plates. Duplicate nitrocellulose replicas of the recombinant bacteriophage plaques and hybridized to oligonucleotide probe #3 in standard hybridization buffer (SHB = 5X SSC, 0.1% SDS, 5X Denhardt's, 100 µg/ml salmon sperm DNA) at 65°C overnight. The following day the radioactively labelled oligonucleotide containing hybridization solution is removed an the filters are washed with 0.2X SSC, 0.1% SDS at 65°C. A single positively hybridizing recombinant is identified and plaque purified. This plaque purified recombinant bacteriophage clone which hybridizes to the BMP-12 oligonucleotide probe #3 is designated \(\lambda\)HuG-48. A bacteriophage plate stock is made and bacteriophage DNA is isolated from the λHuG-48 human genomic clone. The bacteriophage λHuG-48 has been deposited with the American Type Culture Collection, 12301 Parklawn Drive, Rockville, MD "ATCC" under the accession #75625 on December 7, 1993. This deposit meets the requirements of the Budapest Treaty of the International Recognition of the Deposit of Microorganisms for the Purpose of Patent Procedure and Regulations thereunder. The oligonucleotide hybridizing region of this recombinant, λHuG-48, is localized to a 3.2 kb BamHI fragment. This fragment is subcloned into a plasmid vector (pGEM-3) and DNA sequence analysis is performed. This plasmid subclone is designated PCR1-1#2 and has been deposited with the American Type Culture Collection, 12301 Parklawn Drive, Rockville, MD "ATCC" under the accession #69517 on December 7, 1993. This deposit meets the requirements of the Budapest Treaty of the International Recognition of the Deposit of Microorganisms for the Purpose of Patent Procedure and Regulations thereunder. The partial DNA sequence (SEQ ID NO:1) and derived amino acid sequence (SEQ ID NO:2) of the 3.2 kb DNA insert of the plasmid subclone PCR1-1#2, derived from clone λHuG-48, are shown in the Sequence Listings.

It should be noted that nucleotides #639 - #714 of SEQ ID NO:1 correspond to nucleotides #27 - #102 of the specifically amplified BMP-12 encoding DNA fragment set forth in SEQ ID NO:5 thus confirming that the human genomic bacteriophage clone λHuG-48 and derivative subclone PCR1-1#2 encode at least a portion of the BMP-12 protein of the invention. The nucleotide sequence of a

portion of the 3.2 kb BamHI insert of the plasmid PCR1-1#2 contains an open reading frame of at least 882 base pairs, as defined by nucleotides #1-#882 of SEQ ID NO:1. This open reading frame encodes at least 294 amino acids of the human BMP-12 protein of the invention. The encoded 294 amino acid human BMP-12 protein includes the full mature human BMP-12 protein (amino acids #1-#104 of SEQ ID NO:2), as well as the C-terminal portion of the propeptide region of the primary translation product (amino acid #-190 to #-1 of SEQ ID NO:2).

Additional DNA sequence of the 3.2 kb BamHI insert of the plasmid PCR1-1#2 set forth in SEQ ID NO:33 demonstrates the presence of an 1164 bp open reading frame, as defined by nucleotides #138 through #1301 of SEQ ID NO:33. [NOTE that all the sequence disclosed in SEQ ID NO:1 is contained within SEQ ID NO:33]. As this sequence is derived from a genomic clone it is difficult to determine the boundary between the 5' extent of coding sequence and the 3' limit of intervening sequence (intron/non-coding sequence).

Based on the knowledge of other BMP proteins and other proteins within the TGF- β family, it is predicted that the precursor polypeptide would be cleaved at the multibasic sequence Arg-Arg-Gly-Arg in agreement with a proposed consensus proteolytic processing sequence of Arg-X-X-Arg. Cleavage of the BMP-12 precursor polypeptide is expected to generate a 104 amino acid mature peptide beginning with the amino acid Ser at position #1 of SEQ ID NO:2. The processing of BMP-12 into the mature form is expected to involve dimerization and removal of the N-terminal region in a manner analogous to the processing of the related protein TGF- β [Gentry et al., Molec & Cell. Biol., 8:4162 (1988); Derynck et al. Nature, 316:701 (1985)].

It is contemplated therefore that the mature active species of BMP-12 comprises a homodimer of two polypeptide subunits, each subunit comprising amino acids #1 to #104 of SEQ ID NO:2 with a predicted molecular weight of approximately 12,000 daltons. Further active species are contemplated comprising at least amino acids #3 to #103 of SEQ ID NO:2, thereby including the first and last conserved cysteine residue. As with other members of the TGF-β/BMP family of proteins, the carboxy-terminal portion of the BMP-12 protein exhibits greater sequence conservation than the more amino-terminal portion. The percent amino

acid identity of the human BMP-12 protein in the cysteine-rich C-terminal domain (amino acids #3 - #104) to the corresponding region of human BMP proteins and other proteins within the TGF- β family is as follows: BMP-2, 55%; BMP-3, 43%; BMP-4, 53%; BMP-5, 49%; BMP-6, 49%; BMP-7, 50%; BMP-8, 57%; BMP-9, 48%; BMP-10, 57%; activin WC (BMP-11), 38%; Vg1, 46%; GDF-1, 47%; TGF- β 1, 36%; TGF- β 2, 36%; TGF- β 3, 39%; inhibin β (B), 36%; inhibin β (A), 41%.

The human BMP-12 DNA sequence (SEQ ID NO:1), or a portion thereof, can be used as a probe to identify a human cell line or tissue which synthesizes BMP-12 mRNA. Briefly described, RNA is extracted from a selected cell or tissue source and either electrophoresed on a formaldehyde agarose gel and transferred to nitrocellulose, or reacted with formaldehyde and spotted on nitrocellulose directly. The nitrocellulose is then hybridized to a probe derived from the coding sequence of human BMP-12.

Alternatively, the human BMP-12 sequence is used to design oligonucleotide primers which will specifically amplify a portion of the BMP-12 encoding sequence located in the region between the primers utilized to perform the specific amplification reaction. It is contemplated that these human BMP-12 derived primers would allow one to specifically amplify corresponding BMP-12 encoding sequences from mRNA, cDNA or genomic DNA templates. Once a positive source has been identified by one of the above described methods, mRNA is selected by oligo (dT) cellulose chromatography and cDNA is synthesized and cloned in λ gt10 or other λ bacteriophage vectors known to those skilled in the art, for example, λ ZAP by established techniques (Toole et al., supra). It is also possible to perform the oligonucleotide primer directed amplification reaction, described above, directly on a pre-established human cDNA or genomic library which has been cloned into a λ bacteriophage vector. In such cases, a library which yields a specifically amplified DNA product encoding a portion of the human BMP-12 protein could be screened directly, utilizing the fragment of amplified BMP-12 encoding DNA as a probe.

Oligonucleotide primers designed on the basis of the DNA sequence of the human BMP-12 genomic clone λHuG -48 are predicted to allow the specific amplification of human BMP-12 encoding DNA sequences from pre-established human cDNA libraries which are commercially available (ie. Stratagene, La Jolla,

CA or Clontech Laboratories, Inc., Palo Alto, CA). The following oligonucleotide primer is designed on the basis of nucleotides #571 to #590 of the DNA sequence set forth in SEO ID NO:1 and synthesized on an automated DNA synthesizer:

#4: TGCGGATCCAGCCGCTGCAGCCGCAAGCC (SEQ ID NO:21)

The first nine nucleotides of primer #4 (underlined) comprise the recognition sequence for the restriction endonuclease BamHI which can be used to facilitate the manipulation of a specifically amplified DNA sequence encoding the human BMP-12 protein of the invention and are thus not derived from the DNA sequence presented in SEQ ID NO:1.

The following oligonucleotide primer is designed on the basis of nucleotides #866 - #885 of the DNA sequence set forth in SEQ ID NO:1 and synthesized on an automated DNA synthesizer:

#5 GACTCTAGACTACCTGCAGCCGCAGGCCT (SEQ ID NO:22)

The first nine nucleotides of primer #5 (underlined) comprise the recognition sequence for the restriction endonuclease XbaI which can be used to facilitate the manipulation of a specifically amplified DNA sequence encoding the human BMP-12 protein of the invention and are thus not derived from the DNA sequence presented in SEQ ID NO:1.

The standard nucleotide symbols in the above identified primers are as follows: A, adenine; C, cytosine; G, guanine; T, thymine.

Primers #4 and #5 identified above are utilized as primers to allow the amplification of a specific BMP-12 encoding nucleotide sequence from preestablished cDNA libraries which may include the following: human fetal brain cDNA/λZAPII (Stratagene catalog #936206), human liver/λUNI-ZAP XR (Stratagene Catalog #937200), human lung/λUNI-ZAP XR (Stratagene catalog #937205).

Approximately 1 x 10⁸ pfu (plaque forming units) of λ bacteriophage libraries containing human cDNA inserts such as those detailed above are denatured at 95°C for five minutes prior to addition to a reaction mixture containing 200 μ M each deoxynucleotide triphosphates (dATP, dGTP, dCTP and dTTP) 10 mM Tris-HCl pH 8.3, 50 mM KCl, 1.5 mM MgCl₂, 0.001% gelatin, 1.25 units Taq DNA polymerase, 100 pM oligonucleotide primer #4 and 100 pM oligonucleotide primer

#5. The reaction mixture is then subjected to thermal cycling in the following manner: 1 minute at 94°C, 1 minute at 50°C, 1 minute at 72°C for thirty-nine cycles followed by 10 minutes at 72°C.

The DNA which is specifically amplified by this reaction would be expected to generate a BMP-12 encoding product of approximately 333 base pairs, the internal 315 bp of which correspond to nucleotides #571 to #885 of SEQ ID NO:1 and also including 9 bp at each end of the BMP-12 specific fragment which correspond to the restriction sites defined by nucleotides #1 - #9 of primers #4 and #5. The resulting 333 bp DNA product is digested with the restriction endonucleases BamHI and XbaI, phenol extracted, chloroform extracted and ethanol precipitated.

Alternatively, to ethanol precipitation, buffer exchange and removal of small fragments of DNA resulting from the BamHI/XbaI restriction digest is accomplished by dilution of the digested DNA product in 10 mM Tris-HCl pH 8.0, 1 mM EDTA followed by centrifugation through a CentriconTM 30 microconcentrator (W.R. Grace & Co., Beverly, MA; Product #4209). The resulting BamHI/XbaI digested amplified DNA product is subcloned into a plasmid vector (ie. pBluescript, pGEM-3 etc.) between the BamHI and XbaI sites of the polylinker region. DNA sequence analysis of the resulting subclones would be required to confirm the integrity of the BMP-12 encoding insert. Once a positive cDNA source has been identified in this manner, the corresponding cDNA library from which a 333 bp BMP-12 specific sequence was amplified could be screened directly with the 333 bp insert or other BMP-12 specific probes in order to identify and isolate cDNA clones encoding the full-length BMP-12 protein of the invention.

Additional methods known to those skilled in the art may be used to isolate other full-length cDNAs encoding human BMP-12 related proteins, or full length cDNA clones encoding BMP-12 related proteins of the invention from species other than humans, particularly other mammalian species.

The following examples demonstrate the use of the human BMP-12 sequence to isolate homologues from BMP-12 related proteins in a murine genomic DNA library.

The DNA sequence which encodes the human BMP-12 protein of the invention is predicted to be significantly homologous to BMP-12 and BMP-12 related

sequences from species other than humans that it could be utilized to specifically amplify DNA sequences from those other species which would encode the corresponding BMP-12 related proteins. Specifically, the following oligonucleotides are designed on the basis of the human BMP-12 sequence (SEQ ID NO:1) and are synthesized on an automated DNA synthesizer:

#6: GCGGATCCAAGGAGCTCGGCTGGGACGA (SEQ ID NO:23)

#7: GGAATTCCCCACCACCATGTCCTCGTAT (SEQ ID NO:24)

The first eight nucleotides of oligonucleotide primers #6 and #7 (underlined) comprise the recognition sequence for the restriction endonucleases BamHI and EcoRI, respectively. These sequences are utilized to facilitate the manipulation of a specifically amplified DNA sequence encoding a BMP-12 or BMP-12 related protein from a species other than human and are thus not derived from the DNA sequence presented in SEQ ID NO:1. Oligonucleotide primer #6 is designed on the basis of nucleotides #607-#626 of SEQ ID NO:1. Oligonucleotide primer #7 is designed on the basis of the reverse compliment of nucleotides #846-#865 of the DNA sequence set forth in SEQ ID NO:1.

Oligonucleotide primers #6 and #7 identified above are utilized as primers to allow the amplification of specific BMP-12 related sequences from genomic DNA derived from species other than humans. The amplification reaction is performed as follows:

Murine genomic DNA (source: strain Balb c) is sheared by repeated passage through a 25 gauge needle, denatured at 100° C for five minutes and then chilled on ice before adding to a reaction mixture containing 200 μM each deoxynucleotide triphosphates (dATP, DGTP, dCTP and dTTP) 10 mM Tris-HCl pH 8.3, 50 mM KCl, 1.5 mM MgCl₂, 0.001% gelatin, 1.25 units Taq DNA polymerase, 100 pM oligonucleotide primer #6 and 100 pM oligonucleotide primer #7. The reaction mixture is then subjected to thermal cycling in the following manner: 1 minute at 95°C, 1 minute at 55°C, 1 minute at 72°C for forty cycles followed by 10 minutes at 72°C.

The DNA which is specifically amplified by this reaction is ethanol precipitated, digested with the restriction endonucleases BamHI and EcoRI and subjected to agarose gel electrophoresis. A region of the gel, corresponding to the

predicted size of the murine BMP-12 or BMP-12 related encoding DNA fragment, is excised and the specifically amplified DNA fragments contained therein are extracted (by electroelution or by other methods known to those skilled in the art) and subcloned in to a plasmid vector, such as pGEM-3 or pBluescript between the BamHI and EcoRI sites of the polylinker. DNA sequence analysis of one of the resulting subclones named mV1, indicates that the specifically amplified DNA sequence contained therein encodes a portion of a protein which appears to be the murine homolog to either the BMP-12 or VL-1 sequence of the invention. The DNA sequence (SEQ ID NO:10) and derived amino acid sequence (SEQ ID NO:11) of this specifically amplified murine DNA fragment are shown in the sequence listings.

Nucleotides #1-#26 of SEQ ID NO:10 comprise a portion of oligonucleotide #6 and nucleotides #246-#272 comprise a portion of the reverse compliment of oligonucleotide #7 utilized to perform the specific amplification reaction. Nucleotide #27 of SEQ ID NO:10 appears to be the last nucleotide of a codon triplet, and nucleotides #244-#245 of SEQ ID NO:10 appear to be the first two nucleotides of a codon triplet. Therefore, nucleotides #28 to #243 of SEQ ID NO:10 correspond to a partial coding sequence of mV1. Due to the function of oligonucleotides #6 and #7 in initiating the amplification reaction, they may not correspond exactly to the actual sequence encoding the murine homolog to the human BMP-12 or VL-1 protein of the invention and are therefore not translated in the corresponding amino acid sequence derivation (SEQ ID NO:11).

Oligonucleotide probes designed on the basis of the specifically amplified murine BMP-12 or VL-1 DNA sequence set forth in SEQ ID NO:10 can be utilized by those skilled in the art to identify full-length murine BMP-12 or VL-1 encoding clones (either cDNA or genomic).

DNA sequence analysis of another of the resulting subclones named mV2, indicates that the specifically amplified DNA sequence contained therein encodes a portion of a murine BMP-12 related sequence of the invention. The DNA sequence (SEQ ID NO:12) and derived amino acid sequence (SEQ ID NO:13) of this specifically amplified murine DNA fragment are shown in the sequence listings.

Nucleotides #1-#26 of SEQ ID NO:12 comprise a portion of oligonucleotide #6 and nucleotides #246-#272 comprise a portion of the reverse compliment of

oligonucleotide #7 utilized to perform the specific amplification reaction. Nucleotide #27 of SEQ ID NO:12 appears to be the last nucleotide of a codon triplet, and nucleotides #244-#245 of SEQ ID NO:12 appear to be the first two nucleotides of a codon triplet. Therefore, nucleotides #28 to #243 of SEQ ID NO:12 correspond to a partial coding sequence of mV2. Due to the function of oligonucleotides #6 and #7 in initiating the amplification reaction, they may not correspond exactly to the actual sequence encoding the murine BMP-12 related protein of the invention and are therefore not translated in the corresponding amino acid sequence derivation (SEQ ID NO:13).

Oligonucleotide probes designed on the basis of the specifically amplified murine BMP-12 related DNA sequence set forth in SEQ ID NO:12 can be utilized by those skilled in the art to identify full-length murine BMP-12 related encoding clones (either cDNA or genomic).

DNA sequence analysis of another of the resulting subclones named mV9, indicates that the specifically amplified DNA sequence contained therein encodes a portion of a murine BMP-12 related sequence of the invention. This sequence appears to be the murine homolog to the human MP52 DNA sequence described at SEQ ID NO:3. The DNA sequence (SEQ ID NO:14) and derived amino acid sequence (SEQ ID NO:15) of this specifically amplified murine DNA fragment are shown in the sequence listings.

Nucleotides #1-#26 of SEQ ID NO:14 comprise a portion of oligonucleotide #6 and nucleotides #246-#272 comprise a portion of the reverse compliment of oligonucleotide #7 utilized to perform the specific amplification reaction. Nucleotide #27 of SEQ ID NO:14 appears to be the last nucleotide of a codon triplet, and nucleotides #244-#245 of SEQ ID NO:14 appear to be the first two nucleotides of a codon triplet. Therefore, nucleotides #28 to #243 of SEQ ID NO:14 correspond to a partial coding sequence of mV9. Due to the function of oligonucleotides #6 and #7 in initiating the amplification reaction, they may not correspond exactly to the actual sequence encoding the murine BMP-12 related protein of the invention and are therefore not translated in the corresponding amino acid sequence derivation (SEQ ID NO:15).

Oligonucleotide probes designed on the basis of the specifically amplified murine BMP-12 related DNA sequence set forth in SEQ ID NO:14 can be utilized by those skilled in the art to identify full-length murine BMP-12 related encoding clones (either cDNA or genomic).

Alternatively, oligonucleotide primers #6 and #7 identified above are utilized as primers to allow the specific amplification of a 275 base pair DNA probe, the internal 259 bp of which correspond to nucleotides #607 to #865 of SEQ ID NO:1, from the BMP-12 encoding plasmid subclone PCR1-1#2. This 275bp DNA probe was radioactively labelled with ³²P and employed to screen a murine genomic library constructed in the vector λ FIX II (Stratagene catalog #946306). recombinants of the murine genomic library are plated at a density of approximately 20,000 recombinants per plate on 50 plates. Duplicate nitrocellulose replicas of the recombinant bacteriophage plaques are hybridized, under reduced stringency conditions, to the specifically amplified 333 bp probe in standard hybridization buffer (SHB = 5X SSC, 0.1% SDS, 5X Denhardt's, 100 μ g/ml salmon sperm DNA) at 60°C overnight. The following day the radioactively labelled oligonucleotide containing hybridization solution is removed an the filters are washed, under reduced stringency conditions, with 2X SSC, 0.1% SDS at 60°C. Multiple positively hybridizing recombinants are identified and plaque purified. Fragments of the positively hybridizing murine genomic recombinant clones are subcloned into standard plasmid vectors (i.e. pGEM-3) and subjected to DNA sequence analysis.

DNA sequence analysis of one of these subclones named MVR3 indicates that it encodes a portion of the mouse gene corresponding to the PCR product mV1 (murine homolog of the human BMP-12 sequence set forth in SEQ ID NO:1) described above. The partial DNA sequence of this subclone and corresponding amino acid translation are set forth in SEQ ID NO: 29 and SEQ ID NO:30 respectively.

DNA sequence analysis of another one of these subclones named MVR32 indicates that it encodes a portion of the mouse gene corresponding to the PCR product mV2 (murine homolog of the human VL-1 sequence set forth in SEQ ID NO:7) described above. The partial DNA sequence of this subclone and

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corresponding amino acid translation are set forth in SEQ ID NO: 31 and SEQ ID NO:32 respectively.

DNA sequence analysis of another of these subclones named MVR23 indicates that it encodes a portion of the mouse gene corresponding to the PCR product mV9 (murine homolog of the MP-52 sequence set forth in SEQ ID NO:3) described above.

In a similar manner to that which is described above for identifying and isolating human genomic clones encoding the BMP-12 protein of the invention, oligonucleotide probe(s) corresponding to the VL-1 encoding sequence set forth in SEQ ID NO:7 can be designed and utilized to identify human genomic or cDNA sequences encoding the VL-1 (BMP-13) protein. These oligonucleotides would be designed to regions specific for VL-1 encoding sequences and would therefore be likely to be derived from regions of the lowest degree of nucleotide sequence identity between the specifically amplified VL-1 encoding sequence (SEQ ID NO:7) and the specifically amplified BMP-12 encoding sequence (SEQ ID NO:5).

Alternatively, oligonucleotide primers #4 and #5 identified above are utilized as primers to allow the specific amplification of a 333 base pair DNA probe, the internal 315 bp of which correspond to nucleotides #571 to #885 of SEQ ID NO:1, from the BMP-12 encoding plasmid subclone PCR1-1#2. This 333 bp DNA probe was radioactively labelled with ³²P and employed to screen a human genomic library constructed in the vector $\lambda DASH$ II (Stratagene catalog #945203). 1 million recombinants of the human genomic library are plated at a density of approximately 20,000 recombinants per plate on 50 plates. Duplicate nitrocellulose replicas of the recombinant bacteriophage plaques are hybridized, under reduced stringency conditions, to the specifically amplified 333 bp probe in standard hybridization buffer (SHB = 5X SSC, 0.1% SDS, 5X Denhardt's, 100 μ g/ml salmon sperm DNA) at 60°C overnight. The following day the radioactively labelled oligonucleotide containing hybridization solution is removed an the filters are washed, under reduced stringency conditions, with 2X SSC, 0.1% SDS at 60°C. Multiple (approximately 15) positively hybridizing recombinants are identified and plaque purified.

In order to distinguish positively hybridizing recombinants encoding the VL-1 protein of the invention from BMP-12 and other BMP-12-related encoding 34

recombinants which would be predicted to hybridize positively to the 333 bp DNA probe generated from the BMP-12 encoding plasmid PCR1-1#2 utilized in this screening procedure, the following oligonucleotide probe, based on the VL-1 sequence set forth in SEQ ID NO:7, is designed and synthesized on an automated DNA synthesizer:

#8: TGTATGCGACTTCCCGC [SEQUENCE ID NO: 35]

An oligonucleotide corresponding to nucleotides #60 to #76 of SEQ ID NO:7 which contains 5 nucleotide differences to the corresponding region of the BMP-12 encoding sequence set forth in SEQ ID NO:1 (nucleotides #672 to #689) One of the recombinant bacteriophage clones which hybridizes to the VL-1 oligonucleotide probe #8 is designated \(\lambda\)JLDc31. This recombinant bacteriophage clone is plaque purified, a bacteriophage plate stock is made and bacteriophage DNA is isolated from the \(\lambda JLDc31\) human genomic clone. The bacteriophage \(\lambda JLDc31\) has been deposited with the American Type Culture Collection, 12301 Parklawn Drive, Rockville, MD "ATCC" under the accession #75922 on October 20, 1994. This deposit meets the requirements of the Budapest Treaty of the International Recognition of the Deposit of Microorganisms for the Purpose of Patent Procedure and Regulations thereunder. The oligonucleotide hybridizing region of this recombinant, \(\lambda \) LDc31, is localized to a 2.5 kb Eco RI fragment. This fragment is subcloned into a plasmid vector (pGEM-3) and DNA sequence analysis is performed. This plasmid subclone is designated pGEMJLDc31/2.5 and has been deposited with the American Type Culture Collection, 12301 Parklawn Drive, Rockville, MD "ATCC" under the accession # 69710 on October 20, 1994. This deposit meets the requirements of the Budapest Treaty of the International Recognition of the Deposit of Microorganisms for the Purpose of Patent Procedure and Regulations thereunder.

The partial DNA sequence (SEQ ID NO:25) and derived amino acid sequence (SEQ ID NO:26) of a portion of the 2.5 kb DNA insert of the plasmid subclone pGEMJLDc31/2.5, derived from clone λJLDc31, are shown in the Sequence Listings

The DNA sequence of a portion of the 2.5 kb EcoRI insert of the plasmid pGEMJLDc31/2.5 is set forth in SEQ ID NO:25.

contains an 912 bp open reading frame, as defined by nucleotides #52 through #963 of SEQ ID NO:25. As this sequence is derived from a genomic clone it is difficult to determine the boundary between the 5' extent of coding sequence and the 3' limit of intervening sequence (intron/non-coding sequence). The entire open reading frame (nucleotides #52 through #963 of SEQ ID NO:25) encodes a portion of the VL-1 protein of the invention of up to 304 amino acids.

Based on the knowledge of other BMP proteins and other proteins within the TGF- β family, it is predicted that the precursor polypeptide would be cleaved at the multibasic sequence Arg-Arg-Arg in agreement with a proposed consensus proteolytic processing sequence of Arg-X-X-Arg. Cleavage of the VL-1 precursor polypeptide is expected to generate a 120 amino acid mature peptide beginning with the amino acid Thr at position #1 of SEQ ID NO:26. The processing of VL-1 into the mature form is expected to involve dimerization and removal of the N-terminal region in a manner analogous to the processing of the related protein TGF- β [Gentry et al., Molec & Cell. Biol., 8:4162 (1988); Derynck et al. Nature, 316:701 (1985)].

It is contemplated therefore that the mature active species of VL-1 comprises a homodimer of two polypeptide subunits, each subunit comprising amino acids #1 to #120 of SEQ ID NO:26 with a predicted molecular weight of approximately 12,000 daltons. Further active species are contemplated comprising at least amino acids #19 to #119 or #120 of SEQ ID NO:26, thereby including the first and last conserved cysteine residue.

Using such a method, a clone encoding the mature human VL-1 (BMP-13) was obtained. The nucleotide sequence and corresponding amino acid sequence encoded by this clone are listed in the Sequence Listings at SEQ ID NO: 25 and 26, respectively.

EXAMPLE 2

Expression of BMP-12

In order to produce human BMP-12 proteins, the DNA encoding it is transferred into an appropriate expression vector and introduced into mammalian cells or other preferred eukaryotic or prokaryotic hosts by conventional genetic engineering techniques.

In order to produce the human BMP-12 protein in bacterial cells, the following procedure is employed.

Expression of BMP-12 in E. coli

An expression plasmid pALV1-781, for production of BMP-12 in E. coli was constructed which contains the following principal features. Nucleotides 1-2060 contain DNA sequences originating from the plasmid pUC-18 [Norrander et al., Gene <u>26</u>:101-106 (1983)] including sequences containing the gene for β -lactamase which confers resistance to the antibiotic ampicillin in host E. coli strains, and a colE1-derived origin of replication. Nucleotides 2061-2221 contain DNA sequences for the major leftward promotor (pL) of bacteriophage λ [Sanger et al., J. Mol. Biol. 162:729-773 (1982)], including three operator sequences 0_11 , 0_12 and 0_13 . The operators are the binding sites for λcI repressor protein, intracellular levels of which control the amount of transcription initiation from pL. Nucleotides 2222-2723 contain a strong ribosome binding sequence included on a sequence derived from nucleotides 35566 to 35472 and 38137 to 38361 from bacteriophage lambda as described in Sanger et al., J. Mol. Biol. 162:729-773 (1982). Nucleotides 2724-3041 contain a DNA sequence encoding mature BMP-12 protein with all 3' untranslated sequence removed. The BMP-12 DNA sequences introduced into the pALV1-781 expression vector were modified at the 5'end to raise the A+T content without altering the coding capacity. These changes were made to increase the efficiency of translation initiated on the BMP-12 mRNA in E. coli. Nucleotides 3042-3058 provide a "Linker" DNA sequence containing restriction endonuclease sites. Nucleotides 3059-3127 provide a transcription termination sequence based on that of the E. coli asp A gene [Takagi et al., Nucl. Acids Res. 13:2063-2074 (1985)]. Nucleotides 3128-3532 are DNA sequences derived from pUC-18.

Plasmid pALV1-781 was transformed into the *E. coli* host strain GI724 (F, $\underline{lac}I^q$, $\underline{lac}p^{L8}$, ampC:: λcI^+) by the procedure of Dagert and Ehrlich, Gene 6:23 (1979). GI724 (ATCC accession No. 55151) contains a copy of the wild-type λcI repressor gene stably integrated into the chromosome at the $\underline{amp}C$ locus, where it has been placed under the transcriptional control of *Salmonella typhimurium* \underline{trp} promotor/operator sequences. In GI724, λCI protein is made only during growth in tryptophan-free media, such as minimal media or a minimal medium supplemented

with casamino acids such as IMC, described above. Addition of tryptophan to a culture of GI724 will repress the <u>trp</u> promoter and turn off synthesis of λcI , gradually causing the induction of transcription from pL promoters if they are present in the cell.

Transformants were selected on 1.5% w/v agar plates containing IMC medium, which is composed of M9 medium [Miller, "Experiments in Molecular Genetics," Cold Spring Harbor Laboratory, New York (1972)] containing 1 mM MgSO₄ and supplemented with 0.5% w/v glucose, 0.2% w/v casamino acids and 100 μ g/ml ampicillin. GI724 transformed with pALV1-781 was grown at 37°C to an A₅₅₀ of 0.5 in IMC medium containing 100 μ g/ml ampicillin. Tryptophan was then added to a final concentration of 100 μ g/ml and the culture incubated for a further 4 hours. During this time BMP-12 protein accumulates within the "inclusion body" fraction.

Preparation of Protein Monomer

18 g of frozen cells were weighed out and resuspended in 60ml of 100 mM Tris, 10 mM EDTA, 1 mM phenylmethylsulfonyl fluoride [PMSF], pH 8.3. Cells were lysed by 3 passes through a Microfluidizer™ [model #MCF 100 T]. The inclusion body pellet was obtained by centrifugation at 15,000g at 4°C for 20 minutes. The supernatant was decanted, and the pellet was washed with 100 ml of 100 mM Tris, 1.0 M NaCl, 10 mM EDTA, 1 mM PMSF, pH 8.3. The suspension was centrifuged again at 15,000g at 4°C for 10 minutes, and the supernatant decanted. The pellet was then washed with 100 ml of 100 mM Tris, 10 mM EDTA, 1% Triton X-100, 1 mM PMSF, pH 8.3. The suspension was centrifuged again at 15,000g at 4°C for 10 minutes, and the supernatant decanted. The pellet was resuspended with 50 ml of 20 mM Tris, 1 mM EDTA, 1 mM PMSF, pH 8.3, containing 1% DTT in a glass tissue homogenizer. Monomeric BMP-12 was then solubilized by acidification to pH 2.5 with glacial acetic acid. The soluble fraction was isolated by centrifugation at 15,000g for 20 minutes at 4°C.

The supernatant from this centrifugation was collected and chromatographed over a Sephacryl S- 100^{TM} size exclusion column (83 cm x 2.6 cm; \approx 440 ml bed) in 20 ml increments. The Sephacryl S- 100^{TM} column was run with a mobile phase of 1% acetic acid at a flow rate of 1.4 ml/min. Fractions corresponding to BMP-12

monomer were detected by absorbance at 280 nm, and using a computer calculated extinction coefficient of 18200M⁻¹cm⁻¹ and molecular weight (11667 daltons). This size exclusion column pooled material was used as starting material for refolding reactions.

As an alternative to the above, 1.0 g of cells stored at -80°C are measured. Solution (3.4 ml 100 mM TRIS, 10 mM EDTA, pH 8.5) is added. The solution is vortexed until cells are well suspended. 40 μ l 100 mM PMSF in isopropanol is added. The cells are lysed at 1000 psi in a French pressure cell. The inclusion bodies are centrifuged at 4°C for 20 minutes in an Eppendorf microfuge to form pellets. The supernatants are decanted. To one pellet (out of 4 total) 1.0 ml degassed 8.0 M guanidine hydrochloride, 0.5 M TRIS, 5 mM EDTA, pH 8.5, containing 250 mM DTT is added. The pellet is dissolved and argon is blown over the liquid for 30 seconds. Next the solution is incubated at 37°C for one hour. Insoluble material is pelleted for 2-3 minutes in an Eppendorf microfuge at 23°C. 0.5-1.0 ml of supernatant is injected onto a Supelco 2 cm guard cartridge (LC-304), and eluted with an acetonitrile gradient in 0.1% TFA from 1-70% over 35 minutes. BMP-12 elutes between 29 and 31 minutes. Fractions are pooled and the protein concentration determined by adsorbance at 280 nanometers versus 0.1% TFA, using the theoretical extinction coefficient based upon the amino acid content.

As a second alternate method to the above, frozen cell pellets obtained from the *E. coli* transformants as described above are thawed in 30 ml of TE8.3(100:10) buffer (100 mM Tris-HCl pH 8.3, 10 mM Na₂EDTA, 1 mM PMSF). Cells are lysed by three passes through a MicrofluidizerTM [model #MCF 100 T]. The initial inclusion body material pellet is dissolved in 8 M guanidine-HCl, TE8.5(100:10) buffer (100 mM Tris-HCl pH 8.5, 10 mM Na₂EDTA which contained 100 mM DTT, and incubated at 37°C for 1 hour. This material is centrifuged at 12,000 x g for 15 minutes at room temperature.

Refolding of BMP-12 protein using CHAPS system

A sufficient volume of the BMP-12 pool is lyophilized to give 10 μ g of protein. 5 μ l of glass distilled water is added to redissolve the residue, then 100 μ l of refold mix (50 mM Tris, 1.0 M NaCl, 2% 3-(3-chlolamido-37)

propyl)dimethylammonio-1-propane-sulfate (CHAPS), 5 mM EDTA, 2 mM glutathione (reduced) 1 mM glutathione (oxidized); at pH of approximately 8.5). The solution is gently mixed and stored at 23°C for 1-4 days. Dimer formation is assessed by running an aliquot on a Novex 16% tricine gel at 125 volts for 2.5 hours, followed by Coomassie Blue staining and destaining.

BMP-12 dimer was purified using a C4 analytical RP-HPLC (reversed phase-high performance liquid chromatography) column (Vydac 214TP54) which was equilibrated to 1% B buffer (diluted into A buffer) and was run over 35 minutes, during which the protein elutes, using the following gradient (A buffer = 0.1% trifluoroacetic acid, B buffer = 95% acetonitrile, 0.1% trifluoroacetic acid [TFA]), with a flow rate of 1 ml/min:

1-5 minutes 20% B buffer

5-10 minutes 20-30% B buffer

10-30 minutes 30-50% B buffer

30-35 minutes 50-100% B buffer

Protein was monitored by absorbance at 280nm. Peak BMP-12 fractions (eluting between 29 and 31 minutes) were pooled. Purity was assessed by SDS-PAGE. The concentration was determined by absorbance at 280nm, and using the computer calculated extinction coefficient and molecular weight as indicated above.

Expression of BMP-12 in mammalian cells:

Another contemplated preferred expression system for biologically active recombinant human BMP-12 is stably transformed mammalian cells.

One skilled in the art can construct mammalian expression vectors by employing the sequence of SEQ ID NO:1, or other DNA sequences encoding BMP-12 proteins or other modified sequences and known vectors, such as pCD [Okayama et al., Mol. Cell Biol., 2:161-170 (1982)], pJL3, pJL4 [Gough et al., EMBO J., 4:645-653 (1985)] and pMT2 CXM.

The mammalian expression vector pMT2 CXM is a derivative of p91023(b) (Wong et al., Science 228:810-815, 1985) differing from the latter in that it contains the ampicillin resistance gene in place of the tetracycline resistance gene and further contains a XhoI site for insertion of cDNA clones. The functional elements of pMT2 CXM have been described (Kaufman, R.J., 1985, Proc. Natl. Acad. Sci.

USA <u>82</u>:689-693) and include the adenovirus VA genes, the SV40 origin of replication including the 72 bp enhancer, the adenovirus major late promoter including a 5' splice site and the majority of the adenovirus tripartite leader sequence present on adenovirus late mRNAs, a 3' splice acceptor site, a DHFR insert, the SV40 early polyadenylation site (SV40), and pBR322 sequences needed for propagation in <u>E. coli</u>.

Plasmid pMT2 CXM is obtained by EcoRI digestion of pMT2-VWF, which has been deposited with the American Type Culture Collection (ATCC), Rockville, MD (USA) under accession number ATCC 67122. EcoRI digestion excises the cDNA insert present in pMT2-VWF, yielding pMT2 in linear form which can be ligated and used to transform <u>E. coli</u> HB 101 or DH-5 to ampicillin resistance. Plasmid pMT2 DNA can be prepared by conventional methods. pMT2 CXM is then constructed using loopout/in mutagenesis [Morinaga, et al., <u>Biotechnology 84</u>: 636 (1984). This removes bases 1075 to 1145 relative to the Hind III site near the SV40 origin of replication and enhancer sequences of pMT2. In addition it inserts a sequence containing the recognition site for the restriction endonuclease Xho I. A derivative of pMT2CXM, termed pMT23, contains recognition sites for the restriction endonucleases PstI, Eco RI, SalI and XhoI. Plasmid pMT2 CXM and pMT23 DNA may be prepared by conventional methods.

pEMC2β1 derived from pMT21 may also be suitable in practice of the invention. pMT21 is derived from pMT2 which is derived from pMT2-VWF. As described above EcoRI digestion excises the cDNA insert present in pMT-VWF, yielding pMT2 in linear form which can be ligated and used to transform E. Coli HB 101 or DH-5 to ampicillin resistance. Plasmid pMT2 DNA can be prepared by conventional methods.

pMT21 is derived from pMT2 through the following two modifications. First, 76 bp of the 5' untranslated region of the DHFR cDNA including a stretch of 19 G residues from G/C tailing for cDNA cloning is deleted. In this process, a XhoI site is inserted to obtain the following sequence immediately upstream from DHFR. Second, a unique ClaI site is introduced by digestion with EcoRV and XbaI, treatment with Klenow fragment of DNA polymerase I, and ligation to a ClaI linker (CATCGATG). This deletes a 250 bp segment from the adenovirus associated RNA

(VAI) region but does not interfere with VAI RNA gene expression or function. pMT21 is digested with EcoRI and XhoI, and used to derive the vector pEMC2B1.

A portion of the EMCV leader is obtained from pMT2-ECAT1 [S.K. Jung, et al, J. Virol 63:1651-1660 (1989)] by digestion with Eco RI and PstI, resulting in a 2752 bp fragment. This fragment is digested with TaqI yielding an Eco RI-TaqI fragment of 508 bp which is purified by electrophoresis on low melting agarose gel. A 68 bp adapter and its complementary strand are synthesized with a 5' TaqI protruding end and a 3' XhoI protruding end which has a sequence which matches the EMC virus leader sequence from nucleotide 763 to 827. It also changes the ATG at position 10 within the EMC virus leader to an ATT and is followed by a XhoI site. A three way ligation of the pMT21 Eco RI-XhoI fragment, the EMC virus EcoRI-TaqI fragment, and the 68 bp oligonucleotide adapter TaqI-XhoI adapter resulting in the vector pEMC2 β 1.

This vector contains the SV40 origin of replication and enhancer, the adenovirus major late promoter, a cDNA copy of the majority of the adenovirus tripartite leader sequence, a small hybrid intervening sequence, an SV40 polyadenylation signal and the adenovirus VA I gene, DHFR and β -lactamase markers and an EMC sequence, in appropriate relationships to direct the high level expression of the desired cDNA in mammalian cells.

The construction of vectors may involve modification of the BMP-12 DNA sequences. For instance, BMP-12 cDNA can be modified by removing the non-coding nucleotides on the 5' and 3' ends of the coding region. The deleted non-coding nucleotides may or may not be replaced by other sequences known to be beneficial for expression. These vectors are transformed into appropriate host cells for expression of BMP-12 proteins. Additionally, the sequence of SEQ ID NO:1 or other sequences encoding BMP-12 proteins can be manipulated to express BMP-12 protein by isolating the mature coding sequence of nucleotides 571 to 882 of SEQ ID NO:1 and adding at the 5' end sequences encoding the complete propeptides of other BMP proteins.

For example, one skilled in the art can make a fusion protein in which the propertide of BMP-2 is linked in operable fashion to the mature BMP-12 peptide by preparing a DNA vector in which the DNA sequence encoding the BMP-2

propertide is linked in proper reading frame to the DNA sequence encoding the mature BMP-12 peptide. The DNA sequence of such a fusion protein is shown in SEQUENCE ID NO:27.

One skilled in the art can manipulate the sequences of SEQ ID NO:1 by eliminating or replacing the mammalian regulatory sequences flanking the coding sequence with bacterial sequences to create bacterial vectors for intracellular or extracellular expression by bacterial cells, as described above. As another example, the coding sequences could be further manipulated (e.g. ligated to other known linkers or modified by deleting non-coding sequences therefrom or altering nucleotides therein by other known techniques). The modified BMP-12 coding sequence could then be inserted into a known bacterial vector using procedures such as described in T. Taniguchi et al., Proc. Natl Acad. Sci. USA, 77:5230-5233 (1980). This exemplary bacterial vector could then be transformed into bacterial host cells and a BMP-12 protein expressed thereby. For a strategy for producing extracellular expression of BMP-12 proteins in bacterial cells, see, e.g. European patent application EPA 177,343.

Similar manipulations can be performed for the construction of an insect vector [See, e.g. procedures described in published European patent application 155,476] for expression in insect cells. A yeast vector could also be constructed employing yeast regulatory sequences for intracellular or extracellular expression of the factors of the present invention by yeast cells. [See, e.g., procedures described in published PCT application WO86/00639 and European patent application EPA 123,289].

A method for producing high levels of a BMP-12 protein of the invention in mammalian cells may involve the construction of cells containing multiple copies of the heterologous BMP-12 gene. The heterologous gene is linked to an amplifiable marker, e.g. the dihydrofolate reductase (DHFR) gene for which cells containing increased gene copies can be selected for propagation in increasing concentrations of methotrexate (MTX) according to the procedures of Kaufman and Sharp, J. Mol. Biol., 159:601-629 (1982). This approach can be employed with a number of different cell types.

For example, a plasmid containing a DNA sequence for a BMP-12 of the invention in operative association with other plasmid sequences enabling expression thereof and the DHFR expression plasmid pAdA26SV(A)3 [Kaufman and Sharp, Mol. Cell. Biol., 2:1304 (1982)] can be co-introduced into DHFR-deficient CHO cells, DUKX-BII, by various methods including calcium phosphate coprecipitation and transfection, electroporation or protoplast fusion. DHFR expressing transformants are selected for growth in alpha media with dialyzed fetal calf serum, and subsequently selected for amplification by growth in increasing concentrations of MTX (e.g. sequential steps in 0.02, 0.2, 1.0 and 5uM MTX) as described in Kaufman et al., Mol Cell Biol., 5:1750 (1983). Transformants are cloned, and biologically active BMP-12 expression is monitored by the Rosen-modified Sampath-Reddi rat assay described below in Example 5. BMP-12 expression should increase with increasing levels of MTX resistance. BMP-12 polypeptides are characterized using standard techniques known in the art such as pulse labeling with [35S] methionine or cysteine and polyacrylamide gel electrophoresis. Similar procedures can be followed to produce other related BMP-12 proteins.

EXAMPLE 3

Preparation of BMP-2 propertide/BMP-12 mature peptide fusion

In order to construct a vector encoding the BMP-2 propeptide/BMP-12 mature peptide fusion, the following cloning procedure was used to fuse the two sequences together.

First, a DNA restriction enzyme fragment comprising the propeptide of human BMP-2 protein, comprising nucleotides 1 through 843 of SEQ ID NO:27 is cut from pBMP2 \$\triangle\$EMC. pBMP2 \$\triangle\$EMC is a plasmid derived from lambda U20S-39 (ATCC #40345) comprising the entire coding sequence for human BMP-2 protein with the non-translated 5' and 3' sequences of BMP-2 deleted from the vector. The 5' restriction enzyme used was Bgl II and it cuts pBMP2 \$\triangle\$EMC in the vector at nucleotide 979. The 3' restriction enzyme used was Mae II and it cuts pBMP2 \$\triangle\$EMC in the BMP-2 propeptide at nucleotide 1925, just short of the carboxy terminus. The resulting 954 base pair product was then gel isolated and gene cleaned. Second, a DNA restriction enzyme fragment comprising the 5' portion of the human BMP-12 mature peptide DNA sequence, is cut from pPCR1-1#2 V1-1

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(ATCC #69517). The 5' restriction enzyme used was Eae I and it cuts pPCR1-1#2 V1-1 just 3' of N-terminus of the human BMP-12 mature peptide sequence. The resulting 259 base pair product was gel isolated and gene cleaned. Third, two DNA oligos were designed and synthesized, so that when annealed would form a tiny DNA fragment comprising fusion sequence of the extreme 3' end of the human BMP-2 propeptide and the 5' end of BMP-12 mature peptide. The DNA fragment has a 5' Mae II complimentary sticky end which anneals to the 3' restriction enzyme fragment comprising the human BMP-2 propertide. The annealed oligo DNA fragment has a 3' Eae I complimentary sticky end which anneals to the 5' of the restriction enzyme fragment comprising the mature peptide of human BMP-12. The coding strand oligo is named B2/12 and is 13 base pairs long. Next, a DNA fragment encoding the 123 base pairs at the 3' end of the BMP-12 mature peptide fragment was obtained as follows. First, a DNA fragment comprising the propeptide of human BMP-2 protein, comprising nucleotides 1 through 846 is PCR amplified from pBMP2 \(EMC \). The 5' primer (oligo 655a) anneals just 5' of the polylinker. The 3' primer (BMPpro3) anneals to the BMP-2 propertide 3' end and introduces a Bgl II restriction enzyme site by silent sequence mutations. The resulting PCR product was cut with Sal I, which cleaves in the polylinker, and Bgl II. The 850 base pair restriction enzyme fragment (ending in amino acid sequence REKR) was gel isolated and gene cleaned. The BMP-12 mature peptide was PCR amplified using a 5' primer (oligo 5-1) encoding the Bgl II restriction enzyme site by silent sequence mutations, and annealing to the 5' end of a possible mature cleavage product, beginning with amino acid sequence SRCS. The 3' primer (V1-1 3) anneals to the BMP-12 mature peptide 3' end and introduces a Xba I restriction enzyme site after the stop codon. The resulting PCR product was cut with Bgl II and Xba I. The 321 base pair restriction enzyme fragment was gel isolated and gene cleaned.

The two restriction fragments were three-way ligated into a previously Sall and XbaI cut vector. The resultant construct was sequenced to check for PCR induced errors and a silent C to T mutation was observed at base pair 185 in the propeptide. This plasmid was designated pREKRSRC. Then pREKRSRC was cut with BglII and NgoMI, and the vector fragment encompassing the last 123 base pairs of the BMP12 mature sequence was thereby isolated. The three restriction fragments

and the annealed oligolinker were four-way ligated to yield pREKR-TAL with the BMP-2 propeptide with the mature cleavage site at the 3' end fused to the (TAL) 5' end of the BMP-12 mature peptide. The coding sequence of the resulting ligated vector is shown in SEQ ID NO:27.

EXAMPLE 4

Biological Activity of Expressed BMP-12

To measure the biological activity of the expressed BMP-12 proteins obtained in Example 2 above, the proteins are recovered from the cell culture and purified by isolating the BMP-12 proteins from other proteinaceous materials with which they are co-produced as well as from other contaminants. The purified protein may be assayed in accordance with the rat assay described below in Example 5.

Purification is carried out using standard techniques known to those skilled in the art.

Protein analysis is conducted using standard techniques such as SDS-PAGE acrylamide [Laemmli, Nature 227:680 (1970)] stained with Coomassie Blue or silver [Oakley, et al. Anal. Biochem. 105:361 (1980)] and by immunoblot [Towbin, et al. Proc. Natl. Acad. Sci. USA 76:4350 (1979)]

Example 5

ROSEN MODIFIED SAMPATH-REDDI ASSAY

A modified version of the rat ectopic implant assay described in Sampath and Reddi, Proc. Natl. Acad. Sci. USA, 80:6591-6595 (1983) is used to evaluate the activity of the BMP-12 proteins. This modified assay is herein called the Rosen-modified Sampath-Reddi assay. The assay has been widely used to evaluate the bone and cartilage-inducing activity of BMPs. The ethanol precipitation step of the Sampath-Reddi procedure is replaced by dialyzing (if the composition is a solution) or diafiltering (if the composition is a suspension) the fraction to be assayed against water. The solution or suspension is then equilibrated to 0.1% TFA. The resulting solution is added to 20 mg of rat matrix. A mock rat matrix sample not treated with the protein serves as a control. This material is frozen and lyophilized and the resulting powder enclosed in #5 gelatin capsules. The capsules are implanted subcutaneously in the abdominal thoracic area of 21-49 day old male Long Evans rats. The implants are removed after 10 days. A section of each implant is fixed

and processed for histological analysis. 1 μ m glycolmethacrylate sections are stained with Von Kossa and acid fuschin to score the amount of induced tendon/ligament-like tissue formation present in each implant.

BMP-12 was implanted in the rats in doses of 1, 5, 25 and 50 μ g per implant for 10 days. BMP-2 at a dose of 5 μ g was included as a positive control. For all doses of BMP-12 tested, no bone or cartilage formation was observed in the implants after ten days. Instead, the implants were filled with tissue resembling embryonic tendon, which is easily recognized by the presence of dense bundles of fibroblasts oriented in the same plane and packed tightly together. [Tendon/ligament-like tissue is described, for example, in Ham and Cormack, <u>Histology</u> (JB Lippincott Co. (1979), pp. 367-369, the disclosure of which is hereby incorporated by reference]. These findings were reproduced in a second set of assays in which tendon/ligament-like tissues was present in all BMP-12 containing implants. In contrast, the BMP-2 implants, as expected, showed cartilage and bone formation, but contained no tendon/ligament-like tissue.

The BMP-12 proteins and related proteins of this invention may be assessed for activity on this assay.

Example 6

Using methods in accordance with the above examples, with minor modifications within the skill of the art, human MP52 protein and the murine homologue of BMP-13 protein were expressed and assayed for tendon/ligament-like tissue inducing activity. All proteins showed comparable results, similar to those described above for human BMP-12.

The foregoing descriptions detail presently preferred embodiments of the present invention. Numerous modifications and variations in practice thereof are expected to occur to those skilled in the art upon consideration of these descriptions. Those modifications and variations are believed to be encompassed within the claims appended hereto. The disclosure of all references discussed herein are hereby incorporated by reference.

SEQUENCE LISTING

- (1) GENERAL INFORMATION:
 - (i) APPLICANT: GENETICS INSTITUTE, INC. PRESIDENT AND FELLOWS OF HARVARD COLLEGE
 - (ii) TITLE OF INVENTION: TENDON-INDUCING COMPOSITIONS
 - (iii) NUMBER OF SEQUENCES: 35
 - (iv) CORRESPONDENCE ADDRESS:
 - (A) ADDRESSEE: GENETICS INSTITUTE, INC.(B) STREET: 87 CambridgePark Drive

 - (C) CITY: Cambridge
 - (D) STATE: Massachusetts
 - (E) COUNTRY: USA (F) ZIP: 02140
 - (v) COMPUTER READABLE FORM:
 - (A) MEDIUM TYPE: Floppy disk
 - (B) COMPUTER: IBM PC compatible
 - (C) OPERATING SYSTEM: PC-DOS/MS-DOS
 - (D) SOFTWARE: PatentIn Release #1.0, Version #1.25
 - (vi) CURRENT APPLICATION DATA:

 - (A) APPLICATION NUMBER: (B) FILING DATE: Herewith
 - (C) CLASSIFICATION:
 - (vii) PRIOR APPLICATION DATA:
 - (A) APPLICATION NUMBER: US 08/164,103
 - (B) FILING DATE: 07-DEC-1993
 - (C) APPLICATION NUMBER: US 08/217,780 (D) FILING DATE: 25-MAR-1994

 - (E) APPLICATION NUMBER: US 08/333,576 (F) FILING DATE: 02-NOV-1994
 - (viii) ATTORNEY/AGENT INFORMATION:
 - (A) NAME: Lazar, Steven R.
 - (B) REGISTRATION NUMBER: 32,618
 - (C) REFERENCE/DOCKET NUMBER: 5202D-PCT
 - (ix) TELECOMMUNICATION INFORMATION:
 - (A) TELEPHONE: 617 498-8260
 - (B) TELEFAX: 617 876-5851
- (2) INFORMATION FOR SEQ ID NO:1:
 - (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 926 base pairs
 - (B) TYPE: nucleic acid
 - (C) STRANDEDNESS: single
 (D) TOPOLOGY: linear
 - (ii) MOLECULE TYPE: DNA (genomic)
 - (vi) ORIGINAL SOURCE:
 - (A) ORGANISM: Homo sapiens
 - (vii) IMMEDIATE SOURCE:
 - (B) CLONE: v1-1
 - (ix) FEATURE:
 - (A) NAME/KEY: mat_peptide
 - (B) LOCATION: 571..882
 - (ix) FEATURE:
 - (A) NAME/KEY: CDS

(B) LOCATION: 1..882

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:1:

Ile	Ile	Gln	Thr	Leu 55	Leu	Asn	Ser	Met	Ala 60	Pro	Asp	GCG Ala	GCG Ala	Pro 65	GCC	
TCC Ser	TGC Cys	TGT Cys	GTG Val 70	CCA Pro	GCG Ala	CGC Arg	CTC Leu	AGC Ser 75	CCC Pro	ATC Ile	AGC Ser	ATC Ile	CTC Leu 80	TAC Tyr	ATC Ile	
GAC Asp	GCC Ala	GCC Ala 85	AAC Asn	AAC Asn	GTT Val	GTC Val	TAC Tyr 90	AAG Lys	CAA Gln	TAC Tyr	GAG Glu	GAC Asp 95	ATG Met	GTG Val	GTG Val	
GAG Glu	GCC Ala 100	TGC Cys	GGC Gly	TGC Cys	AGG Arg	TAG	CGCG	CGG (GCCG	GGGA(GG G(GCA (GCCA(С		
GCG	GCCG	AGG 1	ATCC													
(2)	INF	ORMA'	TION	FOR	SEQ	ID I	NO:2	:								
		(i) S	(B)	LEI	NGTH PE: a	RACTI : 294 amino	am:	ino a id		s						
			MOLE													
			SEQUI													
-190	Arg	Asn	Thr	Thr	His -18	Tyr 85	Arg	Ala	Asn		Val 180	Arg	Gly	Pro		-175
Ser	Trp	Thr	Ser	Pro -170	Pro	Leu	Leu	Leu	Leu -16		Thr	Cys	Pro		Ala 160	
Ala	Arg	Ala	Pro -155	Arg	Leu	Leu	Tyr	Ser -15		Ala	Ala	Glu		Leu L45	Val	
Gly	Gln	Arg -140	Trp	Glu	Ala	Phe	Asp -13		Ala	Asp	Ala		Arg	Arg	His	
Arg	Arg -125	Glu	Pro	Arg	Pro	Pro -12	Arg 20	Ala	Phe	Cys		Leu .15	Leu	Arg	Ala	
Val -110	Ala	Gly	Pro	Val	Pro -10	Ser)5	Pro	Leu	Ala	Leu -1	Arg 100	Arg	Leu	Gly		-95
Gly	Trp	Pro	Gly	Gly -90	Gly	Gly	Ser	Ala	Ala -85	Glu	Glu	Arg	Ala	Val -80	Leu	
Val	Val	Ser	Ser -75	Arg	Thr	Gln	Arg	Lys -70	Glu	Ser	Leu	Phe	Arg -65	Glu	Ile	
Arg	Ala	Gln -60	Ala	Arg	Ala	Leu	Gly -55	Ala	Ala	Leu	Ala	Ser -50	Glu	Pro	Leu	
Pro	Asp -45	Pro	Gly	Thr	Gly	Thr	Ala	Ser	Pro	Arg	Ala -35	Val	Ile	Gly	Gly	
Arg -30	Arg	Arg	Arg	Arg	Thr -25	Ala	Leu	Ala	Gly	Thr -20	Arg	Thr	Ala	Gln	Gly -15	
Ser	Gly	Gly	Gly	Ala -10	Gly	Arg	Gly	His	Gly -5	Arg	Arg	Gly	Arg	Ser 1	Arg	

Cys Ser Arg Lys Pro Leu His Val Asp Phe Lys Glu Leu Gly Trp Asp

Asp Trp Ile Ile Ala Pro Leu Asp Tyr Glu Ala Tyr His Cys Glu Gly

Leu Cys Asp Phe Pro Leu Arg Ser His Leu Glu Pro Thr Asn His Ala

Ile Ile Gln Thr Leu Leu Asn Ser Met Ala Pro Asp Ala Ala Pro Ala

Ser Cys Cys Val Pro Ala Arg Leu Ser Pro Ile Ser Ile Leu Tyr Ile

Asp Ala Ala Asn Asn Val Val Tyr Lys Gln Tyr Glu Asp Met Val Val 90

Glu Ala Cys Gly Cys Arg 100

- (2) INFORMATION FOR SEQ ID NO:3:
 - (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 1207 base pairs(B) TYPE: nucleic acid

 - (C) STRANDEDNESS: single
 - (D) TOPOLOGY: linear
 - (ii) MOLECULE TYPE: DNA (genomic)
 - (vi) ORIGINAL SOURCE:
 - (A) ORGANISM: Homo sapiens
 - (vii) IMMEDIATE SOURCE:
 - (B) CLONE: MP52
 - (ix) FEATURE:
 - (A) NAME/KEY: CDS
 - (B) LOCATION: 845..1204
 - (xi) SEQUENCE DESCRIPTION: SEQ ID NO:3:

ACCGGGCGGC CCTGAACCCA AGCCAGGACA CCCTCCCCAA ACAAGGCAGG CTACAGCCCG 60 120 CCCCAGCTCC TTCCTGCTGA AGAAGGCCAG GGAGCCCGGG CCCCCACGAG AGCCCAAGGA 180 GCCGTTTCGC CCACCCCCA TCACACCCCA CGAGTACATG CTCTCGCTGT ACAGGACGCT 240 GTCCGATGCT GACAGAAAGG GAGGCAACAG CAGCGTGAAG TTGGAGGCTG GCCTGGCCAA 300 CACCATCACC AGCTTTATTG ACAAAGGGCA AGATGACCGA GGTCCCGTGG TCAGGAAGCA 360 GAGGTACGTG TTTGACATTA GTGCCCTGGA GAAGGATGGG CTGCTGGGGG CCGAGCTCCG 420 480 TGCCCAGCTG AAGCTGTCCA GCTGCCCCAG CGGCCGGCAG CCGGCCTCCT TGCTGGATGT 540 GCGCTCCGTG CCAGGCCTGG ACGGATCTGG CTGGGAGGTG TTCGACATCT GGAAGCTCTT 600 CCGAAACTTT AAGAACTCGG CCCAGCTGTG CCTGGAGCTG GAGGCCTGGG AACGGGGCAG 660 GGCCGTGGAC CTCCGTGGCC TGGGCTTCGA CCGCGCCGCC CGGCAGGTCC ACGAGAAGGC 720

CCT	TTCC	CTG C	STGT	TGG	CC G	CACC	AAGA	A ACC	GGA	CCTG	TTCT	(ATT)	ATG A	AGATT	raaggc	780
CCGC	CTCTC	GC (CAGG	CGA	OA AT	BACCO	TGT	A TG	AGTAC	CTG	TTC	AGCC#	AGC (GCG	AAAACG	840
GCGG		Pro			Thi						y Pro				C CTT Leu 15	889
				AGT Ser 20												937
				TGG Trp												985
				TGC Cys												1033
				ATC Ile												1081
				TGC Cys												1129
CTC Leu	TTC Phe	ATT Ile	GAC Asp	TCT Ser 100	GCC Ala	AAC Asn	AAC Asn	GTG Val	GTG Val 105	TAT Tyr	AAG Lys	CAG Gln	TAT Tyr	GAG Glu 110	GAC Asp	1177
				TCG Ser					TAG							1207

(2) INFORMATION FOR SEQ ID NO:4:

- (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 120 amino acids
 - (B) TYPE: amino acid
 - (D) TOPOLOGY: linear
- (ii) MOLECULE TYPE: protein
- (xi) SEQUENCE DESCRIPTION: SEQ ID NO:4:

Ala Pro Leu Ala Thr Arg Gln Gly Lys Arg Pro Ser Lys Asn Leu Lys

Ala Arg Cys Ser Arg Lys Ala Leu His Val Asn Phe Lys Asp Met Gly
20 25 30

Trp Asp Asp Trp Ile Ile Ala Pro Leu Glu Tyr Glu Ala Phe His Cys
35 40 45

Glu Gly Leu Cys Glu Phe Pro Leu Arg Ser His Leu Glu Pro Thr Asn 50 55 60

His Ala Val Ile Gln Thr Leu Met Asn Ser Met Asp Pro Glu Ser Thr 65 70 75 80

Pro Pro Thr Cys Cys Val Pro Thr Arg Leu Ser Pro Ile Ser Ile Leu 85 90 95

Phe Ile Asp Ser Ala Asn Asn Val Val Tyr Lys Gln Tyr Glu Asp Met 105 Val Val Glu Ser Cys Gly Cys Arg 115 (2) INFORMATION FOR SEQ ID NO:5: (i) SEQUENCE CHARACTERISTICS: (A) LENGTH: 128 base pairs (B) TYPE: nucleic acid (C) STRANDEDNESS: single (D) TOPOLOGY: linear (ii) MOLECULE TYPE: DNA (genomic) (vi) ORIGINAL SOURCE: (A) ORGANISM: Homo Sapiens (vii) IMMEDIATE SOURCE: (B) CLONE: V1-1 fragment (ix) FEATURE: (A) NAME/KEY: CDS (B) LOCATION: 28..102 (xi) SEQUENCE DESCRIPTION: SEQ ID NO:5: GGATCCTGGA AGGATTGGAT CATTGCG CCG CTG GAC TAC GAG GCG TAC CAC 51 Pro Leu Asp Tyr Glu Ala Tyr His TGC GAG GGC CTT TGC GAC TTC CCT TTG CGT TCG CAC CTC GAG CCC ACC Cys Glu Gly Leu Cys Asp Phe Pro Leu Arg Ser His Leu Glu Pro Thr 99 15 AAC CACGCTATAG TCCAAACCTT TCTAGA 128 Asn 25 (2) INFORMATION FOR SEQ ID NO:6: (i) SEQUENCE CHARACTERISTICS: (A) LENGTH: 25 amino acids (B) TYPE: amino acid (D) TOPOLOGY: linear (ii) MOLECULE TYPE: protein (xi) SEQUENCE DESCRIPTION: SEQ ID NO:6: Pro Leu Asp Tyr Glu Ala Tyr His Cys Glu Gly Leu Cys Asp Phe Pro Leu Arg Ser His Leu Glu Pro Thr Asn 20 (2) INFORMATION FOR SEQ ID NO:7: (i) SEQUENCE CHARACTERISTICS: (A) LENGTH: 128 base pairs (B) TYPE: nucleic acid (C) STRANDEDNESS: single (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: DNA (genomic)	•
(vi) ORIGINAL SOURCE: (A) ORGANISM: Homo Sapiens	
(vii) IMMEDIATE SOURCE: (B) CLONE: VL-1	:
(ix) FEATURE: (A) NAME/KEY: CDS (B) LOCATION: 28102	
(xi) SEQUENCE DESCRIPTION: SEQ ID NO:7:	
GGATCCTGGG ATGACTGGAT TATGGCG CCG CTG GAC TAC GAG GCG TAC CAC Pro Leu Asp Tyr Glu Ala Tyr His 1 5	51
TGC GAG GGT GTA TGC GAC TTC CCG CTG CGC TCG CAC CTG GAG CCC ACC Cys Glu Gly Val Cys Asp Phe Pro Leu Arg Ser His Leu Glu Pro Thr 10 15 20	99
AAC CACGCCATGC TACAAACGCT TCTAGA Asn 25	128
(2) INFORMATION FOR SEQ ID NO:8:	
(i) SEQUENCE CHARACTERISTICS: (A) LENGTH: 25 amino acids (B) TYPE: amino acid (D) TOPOLOGY: linear	
(ii) MOLECULE TYPE: protein	
(xi) SEQUENCE DESCRIPTION: SEQ ID NO:8:	
Pro Leu Asp Tyr Glu Ala Tyr His Cys Glu Gly Val Cys Asp Phe Pro 1 5 10 15	
Leu Arg Ser His Leu Glu Pro Thr Asn 20 25	
(2) INFORMATION FOR SEQ ID NO:9:	
 (i) SEQUENCE CHARACTERISTICS: (A) LENGTH: 3585 base pairs (B) TYPE: nucleic acid (C) STRANDEDNESS: single (D) TOPOLOGY: linear 	
(ii) MOLECULE TYPE: DNA (genomic)	
(vii) IMMEDIATE SOURCE: (B) CLONE: pALV1-781	
(xi) SEQUENCE DESCRIPTION: SEQ ID NO:9:	
CTAACTACCC AACTCAAAAA AAAAAAAAA AAAAACCCCC TCTAACCCCC ATTGACGAAA	60
GGGCCTCGTG ATACGCCTAT TTTTATAGGT TAATGTCATG ATAATAATGG TTTCTTAGAC	120
GTCAGGTGGC ACTTTTCGGG GAAATGTGCG CGGAACCCCT ATTTGTTTAT TTTTCTAAAT	180

ACATTCA	TAP	ATGTATCCGC	TCATGAGACA	ATAACCCTGA	TAAATGCTTC	AATAATATTG	240
AAAAAGGA	A AG	AGTATGAGTA	TTCAACATTT	CCGTGTCGCC	CTTATTCCCT	TTTTTGCGGC	300
ATTTTGCC	CTT	CCTGTTTTTG	CTCACCCAGA	AACGCTGGTG	AAAGTAAAAG	ATGCTGAAGA	. 360
TCAGTTGG	3 GT	GCACGAGTGG	GTTACATCGA	ACTGGATCTC	AACAGCGGTA	AGATCCTTGA	420
GAGTTTTC	CGC	CCCGAAGAAC	GTTTTCCAAT	GATGAGCACT	TTTAAAGTTC	TGCTATGTGG	480
CGCGGTAT	CTA	TCCCGTATTG	ACGCCGGGCA	AGAGCAACTC	GGTCGCCGCA	TACACTATTC	540
TCAGAATO	BAC	TTGGTTGAGT	ACTCACCAGT	CACAGAAAAG	CATCTTACGG	ATGGCATGAC	600
AGTAAGAG	AAE	TTATGCAGTG	CTGCCATAAC	CATGAGTGAT	AACACTGCGG	CCAACTTACT	660
TCTGACAA	4CG	ATCGGAGGAC	CGAAGGAGCT	AACCGCTTTT	TTGCACAACA	TGGGGGATCA	720
TGTAACTC	CGC	CTTGATCGTT	GGGAACCGGA	GCTGAATGAA	GCCATACCAA	ACGACGAGCG	780
TGACACCA	ACG	ATGCCTGTAG	CAATGGCAAC	AACGTTGCGC	AAACTATTAA	CTGGCGAACT	840
ACTTACTO	CTA	GCTTCCCGGC	AACAATTAAT	AGACTGGATG	GAGGCGGATA	AAGTTGCAGG	900
ACCACTTO	CTG	CGCTCGGCCC	TTCCGGCTGG	CTGGTTTATT	GCTGATAAAT	CTGGAGCCGG	960
TGAGCGTG	GG	TCTCGCGGTA	TCATTGCAGC	ACTGGGGCCA	GATGGTAAGC	CCTCCCGTAT	1020
CGTAGTTA	ATC	TACACGACGG	GGAGTCAGGC	AACTATGGAT	GAACGAAATA	GACAGATCGC	1080
TGAGATAG	GT	GCCTCACTGA	TTAAGCATTG	GTAACTGTCA	GACCAAGTTT	ACTCATATAT	1140
ACTTTAGA	TT	GATTTAAAAC	TTCATTTTTA	ATTTAAAAGG	ATCTAGGTGA	AGATCCTTTT	1200
TGATAATO	CTC	ATGACCAAAA	TCCCTTAACG	TGAGTTTTCG	TTCCACTGAG	CGTCAGACCC	1260
CGTAGAAA	AAG	ATCAAAGGAT	CTTCTTGAGA	TCCTTTTTTT	CTGCGCGTAA	TCTGCTGCTT	1320
GCAAACAA	AA	AAACCACCGC	TACCAGCGGT	GGTTTGTTTG	CCGGATCAAG	AGCTACCAAC	1380
TCTTTTTC	CG	AAGGTAACTG	GCTTCAGCAG	AGCGCAGATA	CCAAATACTG	TCCTTCTAGT	1440
GTAGCCGT	CAG	TTAGGCCACC	ACTTCAAGAA	CTCTGTAGCA	CCGCCTACAT	ACCTCGCTCT	1500
GCTAATCC	TG	TTACCAGTGG	CTGCTGCCAG	TGGCGATAAG	TCGTGTCTTA	CCGGGTTGGA	1560
CTCAAGAC	GA	TAGTTACCGG	ATAAGGCGCA	GCGGTCGGGC	TGAACGGGGG	GTTCGTGCAC	1620
ACAGCCCA	\GC	TTGGAGCGAA	CGACCTACAC	CGAACTGAGA	TACCTACAGC	GTGAGCATTG	1680
AGAAAGCG	CC	ACGCTTCCCG	AAGGGAGAAA	GGCGGACAGG	TATCCGGTAA	GCGGCAGGGT	1740
CGGAACAG	GA	GAGCGCACGA	GGGAGCTTCC	AGGGGGAAAC	GCCTGGTATC	TTTATAGTCC	1800
TGTCGGGT	TT	CGCCACCTCT	GACTTGAGCG	TCGATTTTTG	TGATGCTCGT	CAGGGGGGCG	1860
GAGCCTAT	GG	AAAAACGCCA	GCAACGCGGC	CTTTTTACGG	TTCCTGGCCT	TTTGCTGGCC	1920
TTTTGCTC	AC	ATGTTCTTTC	CTGCGTTATC	CCCTGATTCT	GTGGATAACC	GTATTACCGC	1980
CTTTGAGT	GA	GCTGATACCG	CTCGCCGCAG	CCGAACGACC	GAGCGCAGCG	AGTCAGTGAG	2040
CGAGGAAG	CG	GAAGAGCGCC	CAATACGCAA	ACCGCCTCTC	CCCGCGCGTT	GGCCGATTCA	2100
TTAATGCA	GA	ATTGATCTCT	CACCTACCAA	ACAATGCCCC	CCTGCAAAAA	ATAAATTCAT	2160
ATAAAAAA	CA	TACAGATAAC	CATCTGCGGT	GATAAATTAT	CTCTGGCGGT	GTTGACATAA	2220

ATACCACTGG	CGGTGATACT	GAGCACATCA	GCAGGACGCA	CTGACCACCA	TGAAGGTGAC	2280
GCTCTTAAAA	ATTAAGCCCT	GAAGAAGGC	AGCATTCAAA	GCAGAAGGCT	TTGGGGTGTG	2340
TGATACGAAA	CGAAGCATTG	GCCGTAAGTG	CGATTCCGGA	TTAGCTGCCA	ATGTGCCAAT	2400
CGCGGGGGGT	TTTCGTTCAG	GACTACAACT	GCCACACACC	ACCAAAGCTA	ACTGACAGGA	2460
GAATCCAGAT	GGATGCACAA	ACACGCCGCC	GCGAACGTCG	CGCAGAGAAA	CAGGCTCAAT	2520
GGAAAGCAGC	AAATCCCCTG	TTGGTTGGGG	TAAGCGCAAA	ACCAGTTCCG	AAAGATTTTT	2580
TTAACTATAA	ACGCTGATGG	AAGCGTTTAT	GCGGAAGAGG	TAAAGCCCTT	CCCGAGTAAC	2640
ааааааасаа	CAGCATAAAT	AACCCCGCTC	TTACACATTC	CAGCCCTGAA	AAAGGGCATC	2700
AAATTAAACC	ACACCTATGG	TGTATGCATT	TATTTGCATA	CATTCAATCA	ATTGTTATCT	2760
AAGGAAATAC	TTACATATGT	CTCGTTGTTC	TCGTAAACCA	CTGCATGTAG	ATTTTAAAGA	2820
GCTCGGCTGG	GACGACTGGA	TCATCGCGCC	GCTGGACTAC	GAGGCGTACC	ACTGCGAGGG	2880
CCTTTGCGAC	TTCCCTTTGC	GTTCGCACCT	CGAGCCCACC	AACCATGCCA	TCATTCAGAC	2940
GCTGCTCAAC	TCCATGGCAC	CAGACGCGGC	GCCGGCCTCC	TGCTGTGTGC	CAGCGCGCCT	3000
CAGCCCCATC	AGCATCCTCT	ACATCGACGC	CGCCAACAAC	GTTGTCTACA	AGCAATACGA	3060
GGACATGGTG	GTGGAGGCCT	GCGGCTGCAG	GTAGTCTAGA	GTCGACCTGC	AGTAATCGTA	3120
CAGGGTAGTA	САААТАААА	AGGCACGTCA	GATGACGTGC	CTTTTTTCTT	GTGAGCAGTA	3180
AGCTTGGCAC	TGGCCGTCGT	TTTACAACGT	CGTGACTGGG	AAAACCCTGG	CGTTACCCAA	3240
CTTAATCGCC	TTGCAGCACA	TCCCCCTTTC	GCCAGCTGGC	GTAATAGCGA	AGAGGCCCGC	3300
ACCGATCGCC	CTTCCCAACA	GTTGCGCAGC	CTGAATGGCG	AATGGCGCCT	GATGCGGTAT	3360
TTTCTCCTTA	CGCATCTGTG	CGGTATTTCA	CACCGCATAT	ATGGTGCACT	CTCAGTACAA	3420
TCTGCTCTGA	TGCCGCATAG	TTAAGCCAGC	CCCGACACCC	GCCAACACCC	GCTGACGCGC	3480
CCTGACGGGC	TTGTCTGCTC	CCGGCATCCG	CTTACAGACA	AGCTGTGACC	GTCTCCGGGA	3540
GCTGCATGTG	TCAGAGGTTT	TCACCGTCAT	CACCGAAACG	CGCGA		3585

(2) INFORMATION FOR SEQ ID NO:10:

- (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 272 base pairs
 (B) TYPE: nucleic acid
 (C) STRANDEDNESS: single

 - (D) TOPOLOGY: linear
- (ii) MOLECULE TYPE: DNA (genomic)
- (vi) ORIGINAL SOURCE:
 - (A) ORGANISM: mouse
- (vii) IMMEDIATE SOURCE:
 - (B) CLONE: mV1
- (ix) FEATURE:

 - (A) NAME/KEY: CDS
 (B) LOCATION: 28..243

	(xi)	SEC	QUENC	E DE	SCRI	PTIC	ON: S	SEQ I	D NO	0:10:						
GGAT	CCA	AGG A	AGCTO	GGC1	G GG	SACGA				rc go le Al						51
			CAC His													99
			ACC Thr													147
			GCT Ala													195
			ATT Ile 60													243
CAAT	racg)	AGG A	ACATO	GTG	er Go	GGA	ATTC									272
(2)	INF	ORMA!	rion	FOR	SEQ	ID 1	NO:1	l:								
		(i) \$	(B)	ENCE LEI TYI	NGTH PE: a	: 72	amin ac:	no ao id								
			MOLE			_										
_			SEQUI						-			_				
Trp 1	He	IIe	Ala	Pro 5	Leu	Asp	Tyr	GIu	Ala 10	Tyr	His	Cys	GIU	15	vaı	
Cys	Asp	Phe	Pro 20	Leu	Arg	Ser	His	Leu 25	Glu	Pro	Thr	Asn	His 30	Ala	Ile	
Ile	Gln	Thr 35	Leu	Leu	Asn	Ser	Met 40	Ala	Pro	Asp	Ala	Ala 45	Pro	Ala	Ser	
Cys	Сув 50	Val	Pro	Ala	Arg	Leu 55	Ser	Pro	Ile	Ser	Ile 60	Leu	Tyr	Ile	Asp	
Ala 65	Ala	Asn	Asn	Val	Val 70	Tyr	Lys									
(2)	INF	ORMA'	TION	FOR	SEQ	ID I	NO:1	2:								
	(i	()	QUENCA) Li B) T C) S D) T	ENGT YPE : TRAN	nuc DEDN	72 b leic ESS:	ase aci sin	pair d	S							

(ii) MOLECULE TYPE: DNA (genomic)

(vi) ORIGINAL SOURCE:
 (A) ORGANISM: mouse

(vii) IMMEDIATE SOURCE: (B) CLONE: mV2

	(ix)	(2	ATURI A) NA 3) LO	ME/F			. 243										
	(xi)	SEC	QUENC	E DE	SCR	PTIC	ON: S	SEQ 1	D NO):12:	:						
GGAT	CCAF	AGG 1	AGCT	CGGCT	rg go	SACGI						CC CT ro Le				5	1
			CAC His													9	9
			ACT Thr													14	7
			TCC Ser													19	5
			ATC Ile 60													24	3
CAAT	racg <i>i</i>	AGG 1	ACATO	GTG	ST GO	GGA/	ATTC									27	2
(2)	INFO	ORMA!	rion	FOR	SEQ	ID I	NO:1	3:									
,		(i) :	(B)	ENCE LEI TYI	NGTH	: 72	amin	no ao id									
	()	ii) 1	MOLE	CULE	TYPI	: p:	rote:	in									
	()	ki) :	SEQUI	ENCE	DES	CRIP'	rion	: SE(Q ID	NO:	13:						
Trp 1	Ile	Ile	Ala	Pro 5	Leu	Glu	Tyr	Glu	Ala 10	Tyr	His	Cys	Glu	Gly 15	Val		
Cys	Asp	Phe	Pro 20	Leu	Arg	Ser	His	Leu 25	Glu	Pro	Thr	Asn	His 30	Ala	Ile		
Ile	Gln	Thr 35	Leu	Met	Asn	Ser	Met 40	Asp	Pro	Gly	Ser	Thr 45	Pro	Pro	Ser		
cys	Cys 50	Val	Pro	Thr	Lys	Leu 55	Thr	Pro	Ile	Ser	Ile 60	Leu	Tyr	Ile	Asp		
Ala 65	Gly	Asn	Asn	Val	Val 70	Tyr	Lys										
(2)	INFO	ORMA'	rion	FOR	SEQ	ID I	NO:1	4:									
	(i)	() ()	c) s:	ENGTI YPE :	H: 2' nuci	72 ba leic ESS:	ase j acie sin	pair: d	s								

	(vi)			L SC GANI			e									
((vii)			TE S		-										
	(ix)	(P		:: ME/K CATI			243									·
	(xi)	SEC	UENC	E DE	SCRI	PTIC	N: S	SEQ J	D NO	:14:						
GGAT	CCAA	AGG P	GCTC	CGGCT	rg ge	ACG						CT CT CO Le				51
				TGC Cys												99
				AAC Asn												147
				ACA Thr 45												195
				CTC Leu												243
CAAT	racga	AGG A	ACATO	GTG	T GO	GGA	ATTC									272
(2)	INFO	ORMAT	rion	FOR	SEQ	ID 1	10:1	5:								
	ı	(i) s	(A) (B)	ENCE LEI TYI	NGTH:	72 min	amin ac:	no ao id								
	(:	li) P	OLE	CULE	TYP	E: p	rote:	in								
	()	ci) S	SEQUE	ENCE	DESC	RIP	rion	: SE	Q ID	NO:	15:					
Trp 1	Ile	Ile	Ala	Pro 5			-	Glu		Phe	His	Cys	Glu	Gly 15	Leu	
Сув	Glu	Phe	Pro 20	Leu	Arg	Ser	His	Leu 25	Glu	Pro	Thr	Asn	His 30	Ala	Val	
Ile	Gln	Thr 35	Leu	Met	Asn	Ser	Met 40	Asp	Pro	Glu	Ser	Thr 45	Pro	Pro	Thr	
Cys	Суs 50	Val	Pro	Thr	Arg	Leu 55	Ser	Pro	Ile	Ser	Ile 60	Leu	Phe	Ile	Asp	
Ser 65	Ala	Asn	Asn	Val	Val 70	Tyr	Lys									
(2)	INF	ORMA!	rion	FOR	SEQ	ID :	NO:1	6 :								
	(i)	()	Ã) Li	CE CI ENGTI YPE:	H: 7	ami	no a		59							

```
(C) STRANDEDNESS: single
          (D) TOPOLOGY: linear
    (ii) MOLECULE TYPE: peptide
    (vi) ORIGINAL SOURCE:
          (A) ORGANISM: BMP/TGF-beta consensus sequence
    (xi) SEQUENCE DESCRIPTION: SEQ ID NO:16:
     Trp Xaa Asp Trp Ile Xaa Ala
(2) INFORMATION FOR SEQ ID NO:17:
     (i) SEQUENCE CHARACTERISTICS:
          (A) LENGTH: 27 base pairs
          (B) TYPE: nucleic acid
(C) STRANDEDNESS: single
          (D) TOPOLOGY: linear
    (ii) MOLECULE TYPE: DNA (genomic)
   (vii) IMMEDIATE SOURCE:
          (B) CLONE: oligonucleotide #1
    (xi) SEQUENCE DESCRIPTION: SEQ ID NO:17:
CGGATCCTGG VANGAYTGGA THRTNGC
(2) INFORMATION FOR SEQ ID NO:18:
     (i) SEQUENCE CHARACTERISTICS:
          (A) LENGTH: 6 amino acids (B) TYPE: amino acid
          (C) STRANDEDNESS: single
          (D) TOPOLOGY: linear
    (ii) MOLECULE TYPE: peptide
   (vii) IMMEDIATE SOURCE:
          (B) CLONE: BMP/TGF-beta consensus sequence
    (xi) SEQUENCE DESCRIPTION: SEQ ID NO:18:
     His Ala Ile Xaa Gln Thr
                      5
(2) INFORMATION FOR SEQ ID NO:19:
     (i) SEQUENCE CHARACTERISTICS:
          (A) LENGTH: 28 base pairs
           (B) TYPE: nucleic acid
          (C) STRANDEDNESS: single
          (D) TOPOLOGY: linear
    (ii) MOLECULE TYPE: DNA (genomic)
```

(vii) IMMEDIATE SOURCE:

(B) CLONE: oligonucleotide #2

27

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:19:	
TTTCTAGAAR NGTYTGNACD ATNGCRTG	28
(2) INFORMATION FOR SEQ ID NO:20:	٠
(i) SEQUENCE CHARACTERISTICS: (A) LENGTH: 40 base pairs (B) TYPE: nucleic acid (C) STRANDEDNESS: single (D) TOPOLOGY: linear	•
(ii) MOLECULE TYPE: DNA (genomic)	
<pre>(vii) IMMEDIATE SOURCE: (B) CLONE: oligonucleotide #3</pre>	
(xi) SEQUENCE DESCRIPTION: SEQ ID NO:20:	
CCACTGCGAG GGCCTTTGCG ACTTCCCTTT GCGTTCGCAC	40
(2) INFORMATION FOR SEQ ID NO:21:	
 (i) SEQUENCE CHARACTERISTICS: (A) LENGTH: 29 base pairs (B) TYPE: nucleic acid (C) STRANDEDNESS: single (D) TOPOLOGY: linear 	
(ii) MOLECULE TYPE: DNA (genomic)	
(vii) IMMEDIATE SOURCE: (A) LIBRARY: oligonucleotide #4	
(xi) SEQUENCE DESCRIPTION: SEQ ID NO:21:	
TGCGGATCCA GCCGCTGCAG CCGCAAGCC	29
(2) INFORMATION FOR SEQ ID NO:22:	
(i) SEQUENCE CHARACTERISTICS: (A) LENGTH: 29 base pairs (B) TYPE: nucleic acid (C) STRANDEDNESS: single (D) TOPOLOGY: linear	
(ii) MOLECULE TYPE: DNA (genomic)	
(vii) IMMEDIATE SOURCE: (B) CLONE: oligonucleotide #5	
(xi) SEQUENCE DESCRIPTION: SEQ ID NO:22:	
GACTCTAGAC TACCTGCAGC CGCAGGCCT	29
(2) INFORMATION FOR SEQ ID NO:23:	
(i) SEQUENCE CHARACTERISTICS: (A) LENGTH: 28 base pairs (B) TYPE: nucleic acid (C) STRANDEDNESS: single	

	(D) TOPOLOGY: linear	
(ii)	MOLECULE TYPE: DNA (genomic)	
(vii)	IMMEDIATE SOURCE: (A) LIBRARY: oligonucleotide #6	<i>:</i> -
(xi)	SEQUENCE DESCRIPTION: SEQ ID NO:23:	
GCGGATCC	AA GGAGCTCGGC TGGGACGA	28
(2) INFO	RMATION FOR SEQ ID NO:24:	
(i)	SEQUENCE CHARACTERISTICS: (A) LENGTH: 28 base pairs (B) TYPE: nucleic acid (C) STRANDEDNESS: single (D) TOPOLOGY: linear	
(ii)	MOLECULE TYPE: DNA (genomic)	
(vii)	IMMEDIATE SOURCE: (B) CLONE: oligonucleotide #7	
(xi)	SEQUENCE DESCRIPTION: SEQ ID NO:24:	
GGAATTCC	CC ACCACCATGT CCTCGTAT	28
(2) INFO	RMATION FOR SEQ ID NO:25:	
(i)	SEQUENCE CHARACTERISTICS: (A) LENGTH: 1171 base pairs (B) TYPE: nucleic acid (C) STRANDEDNESS: single (D) TOPOLOGY: linear	
(ii)	MOLECULE TYPE: DNA (genomic)	
(vii)	IMMEDIATE SOURCE: (B) CLONE: Human VL-1 protein	
(ix)	FEATURE: (A) NAME/KEY: CDS (B) LOCATION: 2964	
(ix)	FEATURE: (A) NAME/KEY: mat_peptide (B) LOCATION: 605964	
(xi)	SEQUENCE DESCRIPTION: SEQ ID NO:25:	
G AAT TCC Asn Ser -201-20	G GAT CTC TCG CAC ACT CCT CTC CGG AGA CAG AAG TAT TTG r Asp Leu Ser His Thr Pro Leu Arg Arg Gln Lys Tyr Leu -195 -190	46
TTT GAT (Phe Asp V	FTG TCC ATG CTC TCA GAC AAA GAA GAG CTG GTG GGC GCG GAG Val Ser Met Leu Ser Asp Lys Glu Glu Leu Val Gly Ala Glu -180 -175	94

CTG Leu -17	Arg	CTC Leu	TTT Phe	CGC Arg	CAG Gln -16	Ala	CCC Pro	TCA Ser	GCG Ala	CCC Pro -16	Trp	GGG Gly	CCA Pro	CCA Pro	GCC Ala -155	142
GGG Gly	CCG Pro	CTC Leu	CAC His	GTG Val -15	Gln	CTC Leu	TTC Phe	CCT Pro	TGC Cys -14	Leu	TCG Ser	CCC Pro	CTA Leu	CTG Leu -14	Leu	190
-GAC Asp	GCG Ala	CGG Arg	ACC Thr -13	Leu	GAC Asp	CCG Pro	CAG Gln	GGG Gly -13	Ala	CCG Pro	CCG Pro	GCC Ala	GGC Gly -12	Trp	GAA Glu	238
GTC Val	TTC Phe	GAC Asp -12	Val	TGG Trp	CAG Gln	GGC Gly	CTG Leu -11!	Arg	CAC His	CAG Gln	CCC Pro	TGG Trp -11	Lys	CAG Gln	CTG Leu	286
TGC Cys	TTG Leu -10	Glu	CTG Leu	CGG Arg	GCC Ala	GCA Ala -10	Trp	GGC Gly	GAG Glu	CTG Leu	GAC Asp -95	GCC Ala	GGG Gly	GAG Glu	GCC Ala	334
GAG Glu -90	GCG Ala	CGC Arg	GCG Ala	CGG Arg	GGA Gly -85	CCC Pro	CAG Gln	CAA Gln	CCG Pro	CCG Pro -80	CCC Pro	CCG Pro	GAC Asp	CTG Leu	CGG Arg -75	382
AGT Ser	CTG Leu	GGC Gly	TTC Phe	GGC Gly -70	CGG Arg	AGG Arg	GTG Val	CGG Arg	CCT Pro -65	CCC Pro	CAG Gln	GAG Glu	CGG Arg	GCC Ala -60	CTG Leu	430
CTG Leu	GTG Val	GTA Val	TTC Phe -55	ACC Thr	AGA Arg	TCC Ser	CAG Gln	CGC Arg -50	AAG Lys	AAC Asn	CTG Leu	TTC Phe	GCA Ala -45	GAG Glu	ATG Met	478
CGC Arg	GAG Glu	CAG Gln -40	CTG Leu	GGC Gly	TCG Ser	GCC Ala	GAG Glu -35	GCT Ala	GCG Ala	GGC Gly	CCG Pro	GGC Gly -30	GCG Ala	GGC Gly	GCC Ala	526
GAG Glu	GGG Gly -25	TCG Ser	TGG Trp	CCG Pro	CCG Pro	CCG Pro -20	TCG Ser	GGC Gly	GCC Ala	CCG Pro	GAT Asp -15	GCC Ala	AGG Arg	CCT Pro	TGG Trp	574
CTG Leu -10	CCC Pro	TCG Ser	CCC Pro	GGC Gly	CGC Arg -5	CGG Arg	CGG Arg	CGG Arg	CGC Arg	ACG Thr 1	GCC Ala	TTC Phe	GCC Ala	AGT Ser 5	CGC Arg	622
CAT His	GGC Gly	AAG Lys	CGG Arg 10	CAC His	GGC Gly	AAG Lys	AAG Lys	TCC Ser 15	AGG Arg	CTA Leu	CGC Arg	TGC Cys	AGC Ser 20	AAG Lys	AAG Lys	670
•	CTG Leu	CAC His 25	GTG Val	AAC Asn	TTC Phe	AAG Lys	GAG Glu 30	CTG Leu	GGC Gly	TGG Trp	GAC Asp	GAC Asp 35	TGG Trp	ATT Ile	ATC Ile	718
	C	CTG Leu	GAG Glu	TAC Tyr	GAG Glu	GCC Ala 45	TAT Tyr	CAC His	TGC Cys	GAG Glu	GGT Gly 50	GTA Val	TGC Cys	GAC Asp	TTC Phe	766
		OF T	TCG Ser	CAC His	CTG Leu 60	Glu	CCC Pro	ACC Thr	AAC Asn	CAC His 65	GCC Ala	ATC Ile	ATC Ile	CAG Gln	ACG Thr 70	814
			,cc	ATG Met 75	GAC Asp	CCC Pro	GGC Gly	TCC Ser	ACC Thr 80	CCG Pro	CCC Pro	AGC. Ser	TGC Cys	TGC Cys 85	GTG Val	862
				JCT TCT	CCC Pro	ATC Ile	AGC Ser	ATT Ile 95	Leu	TAC Tyr	ATC Ile	GAC Asp	GCG Ala 100	GGC Gly	AAT Asn	910

			TAC Tyr												
	AGG Arg 120	TAG	CGGT	GCC :	rttc(CCCC	CG C	CTTG	GCCC(g gaj	ACCAI	AGGT	GGG	CCAA	G T
CCG	CCGCCTTGCA GGGGAGGCCT GGCTGCAGAG AGGCGGAGGA GGAAGCTGGC GCTGGGGGAG														
GCT(SCTGAGGGTG AGGGAACAGC CTGGATGTGA GAGCCGGTGG GAGAGAAGGG AGCGCACCTT														
CCC	CCCAGTAACT TCTACCTGCC AGCCCAGAGG GAAATAT														
(2)	(2) INFORMATION FOR SEQ ID NO:26:														
	(i) SEQUENCE CHARACTERISTICS:(A) LENGTH: 321 amino acids(B) TYPE: amino acid(D) TOPOLOGY: linear														
	(:	ii) 1	MOLE	CULE	TYPI	E: p	rote	in							
	(xi) SEQUENCE DESCRIPTION: SEQ ID NO:26:														
	Ser 1 -20		Leu	Ser	His		Pro 195	Leu	Arg	Arg		Lys -190	Tyr	Leu	Phe
Asp -18!	Val	Ser	Met	Leu	Ser		Lys	Glu	Glu		Val L75	Gly	Ala	Glu	Leu -170
Arg	Leu	Phe	Arg	Gln -169		Pro	Ser	Ala	Pro -16		Gly	Pro	Pro		Gly 155
Pro	Leu	His	Val -150	Gln	Leu	Phe	Pro	Cys -14	Leu 15	Ser	Pro	Leu		Leu 140	Asp
Ala	Arg	Thr -13	Leu 5	Asp	Pro	Gln	Gly -13	Ala 30	Pro	Pro	Ala		Trp 125	Glu	Val
Phe	Asp -120	Val	Trp	Gln	Gly	Leu -1		His	Gln	Pro		Lys 10	Gln	Leu	Cys
Leu -105	Glu	Leu	Arg	Ala	Ala -10	Trp	Gly	Glu	Leu	Asp -9		Gly	Glu	Ala	Glu -90
Ala	Arg	Ala	Arg	Gly -85	Pro	Gln	Gln	Pro	Pro -80	Pro	Pro	Asp	Leu	Arg -75	Ser
Leu	Gly	Phe	Gly -70	Arg	Arg	Val	Arg	Pro -65	Pro	Gln	Glu	Arg	Ala -60	Leu	Leu
Val	Val	Phe -55	Thr	Arg	Ser	Gln	Arg -50	Lys	Asn	Leu	Phe	Ala -45	Glu	Met	Arg
3lu	Gln -40	Leu	Gly	Ser	Ala	Glu -35	Ala	Ala	Gly	Pro	Gly -30	Ala	Gly	Ala	Glu
Gly -25	Ser	Trp	Pro	Pro	Pro -20	Ser	Gly	Ala	Pro	Asp -15	Ala	Arg	Pro	Trp	Leu -10
Pro	Ser	Pro	Gly	Arg -5	Arg	Arg	Arg	Arg	Thr 1	Ala	Phe	Ala	Ser 5	Arg	His
Gly	Lys	Arg 10	His	Gly	Lys	Lys	Ser 15	Arg ص		Arg	Cys	Ser 20		Lys	Pro

Leu His Val Asn Phe Lys Glu Leu Gly Trp Asp Asp Trp Ile Ile Ala 30

Pro Leu Glu Tyr Glu Ala Tyr His Cys Glu Gly Val Cys Asp Phe Pro

Leu Arg Ser His Leu Glu Pro Thr Asn His Ala Ile Ile Gln Thr Leu

Met Asn Ser Met Asp Pro Gly Ser Thr Pro Pro Ser Cys Cys Val Pro

Thr Lys Leu Thr Pro Ile Ser Ile Leu Tyr Ile Asp Ala Gly Asn Asn

Val Val Tyr Lys Gln Tyr Glu Asp Met Val Val Glu Ser Cys Gly Cys 110

Arq 120

(2) INFORMATION FOR SEQ ID NO:27:

- (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 1233 base pairs
 - (B) TYPE: nucleic acid
 - (C) STRANDEDNESS: single
 - (D) TOPOLOGY: linear
- (ii) MOLECULE TYPE: DNA (genomic)
- (vii) IMMEDIATE SOURCE:
 - (B) CLONE: DNA encoding BMP2 propeptide/BMP-12 mature peptide
- (ix) FEATURE:
 - (A) NAME/KEY: CDS
 - (B) LOCATION: 1..1233
- (ix) FEATURE:
 - (A) NAME/KEY: mat_peptide (B) LOCATION: 847..1233
- (xi) SEQUENCE DESCRIPTION: SEQ ID NO:27:
- ATG GTG GCC GGG ACC CGC TGT CTT CTA GCG TTG CTG CTT CCC CAG GTC Met Val Ala Gly Thr Arg Cys Leu Leu Ala Leu Leu Pro Gln Val -282
- CTC CTG GGC GGC GCT GGC CTC GTT CCG GAG CTG GGC CGC AGG AAG 96 Leu Leu Gly Gly Ala Ala Gly Leu Val Pro Glu Leu Gly Arg Arg Lys -265 -260 -255
- TTC GCG GCG GCG TCG TCG GGC CGC CCC TCA TCC CAG CCC TCT GAC GAG 144 Phe Ala Ala Ala Ser Ser Gly Arg Pro Ser Ser Gln Pro Ser Asp Glu ·-245 -240
- GTC CTG AGC GAG TTC GAG TTG CGG CTG CTC AGC ATG TTC GGC CTG AAA 192 Val Leu Ser Glu Phe Glu Leu Arg Leu Leu Ser Met Phe Gly Leu Lys -230 -225
- CAG AGA CCC ACC CCC AGC AGG GAC GCC GTG GTG CCC CCC TAC ATG CTA 240 Gln Arg Pro Thr Pro Ser Arg Asp Ala Val Val Pro Pro Tyr Met Leu -205 -210 -215

GAC Asp	CTG Leu	TAT Tyr	. Arg	AGG Arg	CAC His	TCA Ser	GGT Gly -19	Gln	CCG Pro	GGC	TCA Ser	CCC Pro		CCA Pro	GAC Asp		288
CAC His	CGG Arg -18	Leu	GAG Glu	AGG Arg	GCA Ala	GCC Ala -18	Ser	CGA Arg	GCC Ala	AAC Asn	ACT Thr -17	Val	CGC	AGC Ser	TTC Phe		336
CAC His -17	His	GAA Glu	GAA Glu	TCT Ser	TTG Leu -16	Glu	GAA Glu	CTA Leu	CCA Pro	GAA Glu -16	Thr	AGT Ser	GGG Gly	AAA Lys	ACA Thr	5	384
ACC Thr	CGG Arg	AGA Arg	TTC	TTC Phe -15	Phe	AAT Asn	TTA Leu	AGT Ser	TCT Ser	Ile	CCC Pro	ACG Thr	GAG Glu	GAG Glu -14	Phe		432
ATC Ile	ACC Thr	TCA Ser	GCA Ala -13	Glu	CTT Leu	CAG Gln	GTT Val	TTC Phe -13	Arg	GAA Glu	CAG Gln	ATG Met	CAA Gln -12	Asp	GCT Ala		480
TTA Leu	GGA Gly	AAC Asn -12	Asn	AGC Ser	AGT Ser	TTC Phe	CAT His	His	CGA Arg	ATT Ile	AAT Asn	ATT Ile -11	TAT Tyr 0	GAA Glu	ATC Ile		528
ATA Ile	AAA Lys -10	Pro	GCA Ala	ACA Thr	GCC Ala	AAC Asn -10	Ser	AAA Lys	TTC Phe	CCC Pro	GTG Val -95	ACC Thr	AGA Arg	CTT Leu	TTG Leu		576
GAC Asp -90	Thr	AGG Arg	TTG Leu	GTG Val	AAT Asn -85	CAG Gln	AAT Asn	GCA Ala	AGC Ser	AGG Arg -80	TGG Trp	GAA Glu	AGT Ser	TTT Phe	GAT Asp -75		624
GTC Val	ACC Thr	CCC Pro	GCT Ala	GTG Val -70	ATG Met	CGG Arg	TGG Trp	ACT Thr	GCA Ala -65	CAG Gln	GGA Gly	CAC His	GCC Ala	AAC Asn -60	CAT His		672
GGA Gly	TTC Phe	GTG Val	GTG Val -55	GAA Glu	GTG Val	GCC Ala	CAC His	TTG Leu -50	GAG Glu	GAG Glu	AAA Lys	CAA Gln	GGT Gly -45	GTC Val	TCC Ser		720
AAG Lys	AGA Arg	CAT His -40	GTT Val	AGG Arg	ATA Ile	AGC Ser	AGG Arg -35	TCT Ser	TTG Leu	CAC His	CAA Gln	GAT Asp -30	GAA Glu	CAC His	AGC Ser		768
TGG Trp	TCA Ser -25	CAG Gln	ATA Ile	AGG Arg	CCA Pro	TTG Leu -20	CTA Leu	GTA Val	ACT Thr	TTT Phe	GGC Gly -15	CAT His	GAT Asp	GGA Gly	AAA Lys		816
GGG Gly -10	CAT His	CCT Pro	CTC Leu	CAC His	AAA Lys -5	AGA Arg	GAA Glu	AAA Lys	CGT Arg	ACG Thr 1	GCG Ala	TTG Leu	GCC Ala	GGG Gly 5	ACG Thr		864
CGG Arg	ACA Thr	GCG Ala	CAG Gln 10	GGC Gly	AGC Ser	GGC Gly	GGG Gly	GGC Gly 15	GCG Ala	GGC Gly	CGG Arg	GGC Gly	CAC His 20	GGG Gly	CGC Arg		912
AGG Arg	GGC Gly	CGG Arg 25	AGC Ser	CGC Arg	TGC Cys	AGC Ser	CGC Arg 30	AAG Lys	CCG Pro	TTG Leu	CAC His	GTG Val 35	GAC Asp	TTC Phe	AAG Lys		960
GAG Glu	CTC Leu 40	GGC Gly	TGG Trp	GAC Asp	Aap GAC	TGG Trp 45	ATC Ile	ATC Ile	GCG Ala	CCG Pro	CTG Leu 50	GAC Asp	TAC Tyr	GAG Glu	GCG Ala	1	800
TAC Tyr 55	CAC His	TGC Cys	GAG Glu	GGC Gly	CTT Leu 60	TGC Cys	GAC Asp	Phe	CCT Pro	TTG Leu 65	CGT Arg	TCG Ser	CAC His	CTC Leu	GAG Glu 70	1	056

		GCC Ala 75						CCA Pro	1104
		GCC Ala							·1152
		ATC Ile					_		1200
		GTG Val	 	 	 				1233

(2) INFORMATION FOR SEQ ID NO:28:

- (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 411 amino acids
 - (B) TYPE: amino acid
 - (D) TOPOLOGY: linear
- (ii) MOLECULE TYPE: protein
- (xi) SEQUENCE DESCRIPTION: SEQ ID NO:28:

Met Val Ala Gly Thr Arg Cys Leu Leu Ala Leu Leu Leu Pro Gln Val
-282 -280 -275 -270

Leu Leu Gly Gly Ala Ala Gly Leu Val Pro Glu Leu Gly Arg Arg Lys
-265 -260 -255

Phe Ala Ala Ala Ser Ser Gly Arg Pro Ser Ser Gln Pro Ser Asp Glu
-250 -245 -240 -235

Val Leu Ser Glu Phe Glu Leu Arg Leu Leu Ser Met Phe Gly Leu Lys
-230 -225 -220

Gln Arg Pro Thr Pro Ser Arg Asp Ala Val Val Pro Pro Tyr Met Leu
-215 -210 -205

Asp Leu Tyr Arg Arg His Ser Gly Gln Pro Gly Ser Pro Ala Pro Asp
-200 -195 -190

His Arg Leu Glu Arg Ala Ala Ser Arg Ala Asn Thr Val Arg Ser Phe
-185 -180 -175

His His Glu Glu Ser Leu Glu Glu Leu Pro Glu Thr Ser Gly Lys Thr
-170 -165 -160 -155

Thr Arg Arg Phe Phe Phe Asn Leu Ser Ser Ile Pro Thr Glu Glu Phe
-150 -145 -140

Ile Thr Ser Ala Glu Leu Gln Val Phe Arg Glu Gln Met Gln Asp Ala
-135 -130 -125

Leu Gly Asn Asn Ser Ser Phe His His Arg Ile Asn Ile Tyr Glu Ile
-120 -115 -110

Ile Lys Pro Ala Thr Ala Asn Ser Lys Phe Pro Val Thr Arg Leu Leu
-105 -100 -95

Asp Thr Arg Leu Val Asn Gln Asn Ala Ser Arg Trp Glu Ser Phe Asp
-90 -85 -80 -75

Val Thr Pro Ala Val Met Arg Trp Thr Ala Gln Gly His Ala Asn His
-70 -65 -60

Gly Phe Val Val Glu Val Ala His Leu Glu Glu Lys Gln Gly Val Ser
-55 -50 -45

Lys Arg His Val Arg Ile Ser Arg Ser Leu His Gln Asp Glu His Ser
-40 -35 -30

Trp Ser Gln Ile Arg Pro Leu Leu Val Thr Phe Gly His Asp Gly Lys
-25 -15

Gly His Pro Leu His Lys Arg Glu Lys Arg Thr Ala Leu Ala Gly Thr
-10 -5 1 5

Arg Thr Ala Gln Gly Ser Gly Gly Gly Ala Gly Arg Gly His Gly Arg
10 15 20

Arg Gly Arg Ser Arg Cys Ser Arg Lys Pro Leu His Val Asp Phe Lys
25 30 35

Glu Leu Gly Trp Asp Asp Trp Ile Ile Ala Pro Leu Asp Tyr Glu Ala
40 45 50

Tyr His Cys Glu Gly Leu Cys Asp Phe Pro Leu Arg Ser His Leu Glu 55 60 65 70

Pro Thr Asn His Ala Ile Ile Gln Thr Leu Leu Asn Ser Met Ala Pro
75 80 85

Asp Ala Ala Pro Ala Ser Cys Cys Val Pro Ala Arg Leu Ser Pro Ile 90 95 100

Ser Ile Leu Tyr Ile Asp Ala Ala Asn Asn Val Val Tyr Lys Gln Tyr 105 110 115

Glu Asp Met Val Val Glu Ala Cys Gly Cys Arg 120 125

- (2) INFORMATION FOR SEQ ID NO:29:
 - (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 1203 base pairs
 - (B) TYPE: nucleic acid
 - (C) STRANDEDNESS: single
 - (D) TOPOLOGY: linear
 - (ii) MOLECULE TYPE: DNA (genomic)
 - (vii) IMMEDIATE SOURCE:
 - (B) CLONE: murine MV1
 - (ix) FEATURE:
 - (A) NAME/KEY: CDS
 - (B) LOCATION: 2..721
 - (xi) SEQUENCE DESCRIPTION: SEQ ID NO:29:
- A AAG TTC TGC CTG GTG CTG GNG NCG GTG ACG GCC TCG GAG AGC AGN Lys Phe Cys Leu Val Leu X01 X02 Val Thr Ala Ser Glu Ser X03 1 5 10
- CNG CTG GCC CTG AGA CGA CTG GGC TTC GGC TGN CCG GGC GGT GGC GAC X04 Leu Ala Leu Arg Arg Leu Gly Phe Gly X05 Pro Gly Gly Gly Asp

94

46

200

GGC Gly	GGC Gly	GGC Gly	ACT Thr 35	GCG Ala	GNC X06	GAG Glu	GAG Glu	CGC Arg 40	GCG Ala	CTG Leu	TTG Leu	GTG Val	ATC Ile 45	TCC Ser	TCC Ser	142
CGT Arg	ACG Thr	CAA Gln 50	AGG Arg	AAA Lys	GAG Glu	AGT Ser	CTG Leu 55	TTC Phe	CGG Arg	GAG Glu	ATC Ile	CGA Arg 60	GCC Ala	CAG Gln	GCC Ala	. 190
CGT Arg	GCT Ala 65	CTC Leu	CGG Arg	GCC Ala	GCT Ala	GCA Ala 70	GAG Glu	CCG Pro	CCA Pro	CCG Pro	GAT Asp 75	CCA Pro	GGA Gly	CCA Pro	GGC Gly	238
GCT Ala 80	GGG Gly	TCA Ser	CGC Arg	AAA Lys	GCC Ala 85	AAC Asn	CTG Leu	GGC Gly	GGT Gly	CGC Arg 90	AGG Arg	CGG Arg	CAG Gln	CGG Arg	ACT Thr 95	286
GCG Ala	CTG Leu	GCT Ala	GGG Gly	ACT Thr 100	CGG Arg	GGA Gly	GNG X07	NAG X08	GGA Gly 105	AGC Ser	GGT Gly	GGT Gly	GGC Gly	GGC Gly 110	GGT Gly	334
GGC Gly	GGT Gly	GGC Gly	GGC Gly 115	GGC Gly	GGC Gly	GGC Gly	GGC Gly	GGC Gly 120	GGC Gly	GGC Gly	GGC Gly	GGC Gly	GGC Gly 125	GGC Gly	GCA Ala	382
GGC Gly	AGG Arg	GGC Gly 130	CAC His	GGG Gly	CGC Arg	AGA Arg	GGC Gly 135	CGG Arg	AGC Ser	CGC Arg	TGC Cys	GGT Gly 140	CGC Arg	AAG Lys	TCA Ser	430
CTG Leu	CAC His 145	GTG Val	GAC Asp	TTT Phe	AAG Lys	GAG Glu 150	CTG Leu	GGC Gly	TGG Trp	GAC Asp	GAC Asp 155	TGG Trp	ATC Ile	ATC Ile	GCG Ala	478
CCA Pro 160	TTA Leu	GAC Asp	TAC Tyr	GAG Glu	GCA Ala 165	TAC Tyr	CAC His	TGC Cys	GAG Glu	GGC Gly 170	GTT Val	TGC Cys	GAC Asp	TTT Phe	CCT Pro 175	526
CTG Leu	CGC Arg	TCG Ser	CAC His	CTG Leu 180	GAG Glu	CCT Pro	ACC Thr	AAC Asn	CAC His 185	GCC Ala	ATC Ile	ATT Ile	CAG Gln	ACG Thr 190	CTG Leu	574
CTC Leu	AAC Asn	TCC Ser	ATG Met 195	GCG Ala	CCC Pro	GAC Asp	GCT Ala	GCG Ala 200	CCA Pro	GCC Ala	TCC Ser	TGC Cys	TGC Cys 205	GTG Val	CCC Pro	622
GCA Ala	AGG Arg	CTC Leu 210	AGT Ser	CCC Pro	ATC Ile	AGC Ser	ATT Ile 215	CTC Leu	TAC Tyr	ATC Ile	GAT Asp	GCC Ala 220	GCC Ala	AAC Asn	AAC Asn	670
GTG Val	GTC Val 225	TAC Tyr	AAG Lys	CAG Gln	TAC Tyr	GAA Glu 230	GAC Asp	ATG Met	GTG Val	GTG Val	GAG Glu 235	GCC Ala	TGC Cys	GGC Gly	TGC Cys	718
AGG Arg 240	TAGO	ATGO	GG 1	CTG	GGAG	G GI	CTGC	CCGC	CCA	\GGA(CCT	AGCI	CAAC	EAG		771
CAGG	TGTC	AT C	AGGC	CCGA	G GG	ACG	GCGG	A CTA	TGGC	CTC	TGCC	AGC	ACA C	AGGA	GAGCA	831
CACA	GTTA	AC A	CTC	CATI	T AC	CACAC	TCCI	TCF	CTCA	CGC	ACAT	GTTI	TAC C	GTGG	ACGGC	891
AGGC	GCTA	AA A	GÇC1	TGCI	T A	TTGC	TACC	C ATT	GATA	CAA	ACCI	CTGI	rcc 1	TTTC	CGGGAG	951
AGGG	AAGG	GC A	TCTG	TGT	T A	GTT	CAGI	CAA 1	TGGC	ACT	LAAA	CCA	AGT A	GAAA	TGGGT	1011
TAGO	ATTO	GA I	TCTC	CTTI	T AC	TTG	AGGC	GG1	GTGG	CTG	GATT	CCT	SAC C	TTGG	SATATG	1071
GAGI	GCAC	TG C	AGGG	CTGG	G AT	ACCO	AGAT	TCI	CTGG	AGT	GGGC	ATTO	GG A	ACCI	TCAAA	1131

AGTAAGGAGC CACTGGGGCT TGGGAGGGAG CACCCGGTTC CTAAACAAGT CTGATGTGTA 1191 1203 CTGCTCAGTT TG

(2) INFORMATION FOR SEQ ID NO:30:

- (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 240 amino acids
 - (B) TYPE: amino acid (D) TOPOLOGY: linear
- (ii) MOLECULE TYPE: protein
- (xi) SEQUENCE DESCRIPTION: SEQ ID NO:30:

Lys Phe Cys Leu Val Leu X01 X02 Val Thr Ala Ser Glu Ser X03 X04

Leu Ala Leu Arg Arg Leu Gly Phe Gly X05 Pro Gly Gly Asp Gly

Gly Gly Thr Ala X06 Glu Glu Arg Ala Leu Leu Val Ile Ser Ser Arg

Thr Gln Arg Lys Glu Ser Leu Phe Arg Glu Ile Arg Ala Gln Ala Arg

Ala Leu Arg Ala Ala Ala Glu Pro Pro Pro Asp Pro Gly Pro Gly Ala

Gly Ser Arg Lys Ala Asn Leu Gly Gly Arg Arg Arg Gln Arg Thr Ala

Leu Ala Gly Thr Arg Gly X07 X08 Gly Ser Gly Gly Gly Gly Gly

Arg Gly His Gly Arg Arg Gly Arg Ser Arg Cys Gly Arg Lys Ser Leu

His Val Asp Phe Lys Glu Leu Gly Trp Asp Asp Trp Ile Ile Ala Pro

Leu Asp Tyr Glu Ala Tyr His Cys Glu Gly Val Cys Asp Phe Pro Leu

Arg Ser His Leu Glu Pro Thr Asn His Ala Ile Ile Gln Thr Leu Leu

Asn Ser Met Ala Pro Asp Ala Ala Pro Ala Ser Cys Cys Val Pro Ala 200

Arg Leu Ser Pro Ile Ser Ile Leu Tyr Ile Asp Ala Ala Asn Asn Val

Val Tyr Lys Gln Tyr Glu Asp Met Val Val Glu Ala Cys Gly Cys Arg 230

(2) INFORMATION FOR SEQ ID NO:31:

- (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 1046 base pairs (B) TYPE: nucleic acid 70

(C) STRANDEDNESS: single

		(I) TC	POLC	GY:	line	ear									
	(ii)	MOI	ECUI	E T	PE:	DNA	(ger	omic	2)							•
((iii)	HYI	POTHE	TICA	AL: N	10										2
	(iv)	ANT	ri-se	ENSE :	NO											
((vii)				OURC MUF		MV2									
	(ix)	(2		ME/I	ŒY: ION:		790									
	(xi)	SEC	QUENC	CE DI	ESCRI	PTIC	ON: S	SEQ :	ID NO):31	:					
					GC AT /S I] 5				ly Pi					ly Se		46
					AGG Arg											94
GGC Gly	CTG Leu	CGC Arg	CCT Pro 35	CAG Gln	CCT Pro	TGG Trp	AAG Lys	CAG Gln 40	CTG Leu	TGC Cys	CTG Leu	GAG Glu	TTG Leu 45	CGG Arg	GCA Ala	142
					GAC Asp											190
CCC Pro	CAG Gln 65	CAG Gln	CCA Pro	CCG Pro	CCT Pro	CTG Leu 70	GAC Asp	CTG Leu	CGG Arg	AGT Ser	CTG Leu 75	GGC Gly	TTC Phe	GGT Gly	CGG Arg	238
					CAG Gln 85											286
					CTG Leu											334
					GCC Ala				Cys							382
					TGG Trp											430
					CGT Arg											478
					AAG Lys 165											526
					ATC Ile											574

GAG Glu	GGC Gly	GTG Val	TGC Cys 195	GAC Asp	TTT Phe	CCG Pro	CTG Leu	CGC Arg 200	TCG Ser	CAC His	CTT Leu	GAG Glu	CCC Pro 205	ACT Thr	AAC Asn	622
CAT His	GCC Ala	ATC Ile 210	ATT Ile	CAG Gln	ACG Thr	CTG Leu	ATG Met 215	AAC Asn	TCC Ser	ATG Met	GAC Asp	CCG Pro 220	GGC Gly	TCC Ser	ACC Thr	. 670
CCG Pro	CCT Pro 225	AGC Ser	TGC Cys	TGC Cys	GTT Val	CCC Pro 230	ACC Thr	AAA Lys	CTG Leu	ACT Thr	CCC Pro 235	ATT Ile	AGC Ser	ATC Ile	CTG Leu	718
TAC Tyr 240	ATC Ile	GAC Asp	GCG Ala	GGC Gly	AAT Asn 245	AAT Asn	GTN X02	GTC Val	TAC Tyr	AAG Lys 250	CAG Gln	TAT Tyr	GAG Glu	GAC Asp	ATG Met 255	766
				TGC Cys 260				TAG	CGGT	GCT (STCC	CGCC	AC C	rggg(CCAGG	820
GAC	CATG	GAG (GGAG	GCCT(GA C	rgcco	GAGAI	A AGO	BAGC	AGGA	GCT	GCC.	rtg (GAAG	AGGCCA	880
CAG	GTGG	GGG 2	ACAG	CCTG	AA AA	GTAG	GAGC	A CA	AATE	GAAG	CAG	CCA	GCC '	TTCC	CAGAAC	940
CTT	CCAA!	rcc (CCA	ACCC	AG A	AGCA	3CTA	A GG	GTT	rcac	AAC	r tt t(GC (CTTG	CCAGCC	1000
TGG	AAAG	ACT I	AGAC	AAGA	GG G	ATTC:	rtct(C TT	TTA:	TAT	GGC'	rtg				1046

(2) INFORMATION FOR SEQ ID NO:32:

- (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 263 amino acids
 - (B) TYPE: amino acid
 - (D) TOPOLOGY: linear
- (ii) MOLECULE TYPE: protein
- (xi) SEQUENCE DESCRIPTION: SEQ ID NO:32:

Arg Lys Gln Ala Cys Ile Pro Ala Gly Pro Thr Leu Arg Gly Ser Ser 1 10 15

Gly Thr Gln Pro Arg Pro Ala Gly Lys Ser Phe Asp Val Trp Gln Gly

Leu Arg Pro Gln Pro Trp Lys Gln Leu Cys Leu Glu Leu Arg Ala Ala
35 40 45

Trp Gly Glu Leu Asp X01 Gly Asp Thr Gly Ala Arg Ala Arg Gly Pro 50 60

Gln Gln Pro Pro Leu Asp Leu Arg Ser Leu Gly Phe Gly Arg Arg 65 70 75 80

Val Arg Pro Pro Gln Glu Arg Ala Leu Leu Val Val Phe Thr Arg Ser 85 90 95

Gln Arg Lys Asn Leu Phe Thr Glu Met His Glu Gln Leu Gly Ser Ala 100 105 110

Glu Ala Ala Gly Ala Glu Gly Ser Cýs Pro Ala Pro Ser Gly Ser Pro 115 120 125

Asp Thr Gly Ser Trp Leu Pro Ser Pro Gly Arg Arg Arg Arg Arg Thr 130 135 140

Ala 145	Phe	Ala	Ser	Arg	His 150	Gly	Lys	Arg	His	Gly 155	Lys	Lys	Ser	Arg	Leu 160	
Arg	Cys	Ser	Arg	Lys 165	Pro	Leu	His	Val	Asn 170	Phe	Lys	Glu	Leu	Gly 175	Trp	
Asp	Asp	Trp	Ile 180	Ile	Ala	Pro	Leu	Glu 185	Tyr	Glu	Ala	Tyr	His 190	Cys	Glu	
Gly	Val	Cys 195	Asp	Phe	Pro	Leu	Arg 200	Ser	His	Leu	Glu	Pro 205	Thr	Asn	His	
Ala	Ile 210	Ile	Gln	Thr	Leu	Met 215	Asn	Ser	Met	Asp	Pro 220	Gly	Ser	Thr	Pro	
Pro 225	Ser	Cys	Cys	Val	Pro 230	Thr	Lys	Leu	Thr	Pro 235	Ile	Ser	Ile	Leu	Tyr 240	
Ile	Asp	Ala	Gly	Asn 245	Asn	X02	Val	Tyr	Lys 250	Gln	туг	Glu	Asp	Met 255	Val	
Val	Glu	Ser	Cys 260	Gly	Cys	Arg										
(2)	INF	ORMAT	rion	FOR	SEQ	ID I	NO:3	3:								
	(ii) (iii) (iv)	(1) (1) (1) (1) (1) (1) (1) (1) (1) (1)	A) Li 3) T C) S C) S C) T C D T C P O T H F I - S I M T M T M M	CE CIENGTI YPE: TRANI OPOLO LE TI ETICI ENSE	H: 11 nucl DEDNI DGY: YPE: AL: NO	Massine DNA	base acid sing ear (gen	pai: d gle								
	(ix)) FEA	ATURI A) NI		KEY:	CDS		01								
	(ix)) FE2 (2 (1	A) N	E: AME/I OCAT:	KEY: ION:	mat 990	_pepi	tide 01								
	(xi)	SEC	QUEN	CE DI	escr:	PTIC	ON:	SEQ :	ID N	0:33	:					
AAC'	TATA	GCA (CTG	CAGT	cc c	rggt	CTTG	G GT	GTAG	GGT	GCG	CTCC	rgg '	rccc	3CGGC	т б
CAG	GAT?	ATG (CAGT	BACC	T AA	GGT	rgtt(G GC	CTGA:	rggg	ACT'	TTTG	GCT '	rgct	AAACC	A 12
AAG	CTCG	GTT (CGGA!		CCC (Pro (-284	ily i		Arg :					rp i			17
				AGG Arg Q					Arg					Trp		21

CAA Gln	CAG Gln	GCC Ala	TGG Trp	CTC Leu	CCA Pro	CAT His	CGA Arg	AGA Arg	CAG Gln	CTG Leu	GGC Gly	CAT His	Leu	CTG Leu	TTA Leu	266
GGA	GGC	-255	GCG	CTG	ACA	GTG	-250	AGG	ATT	TGC	тст	-245	ACA	GCT	CTT	· 314
Gly	Gly -240		Ala	Leu	Thr	-235		Arg	ITE	Cys	-23(Tyr)	IIII	Ara	пеп	
Ser -225	Leu	TGT Cys	CCC Pro	TGC Cys	CGG Arg -220	Ser	CCC Pro	GCA Ala	GAC Asp	GAA Glu -215	Ser	GCA Ala	GCC Ala	GAA Glu	ACA Thr -210	362
GGC Gly	CAG Gln	AGC Ser	TTC Phe	CTG Leu -205	Phe	GAC Asp	GTG Val	TCC Ser	AGC Ser -200	Leu	AAC Asn	GAC Asp	GCA Ala	GAC Asp -195	Glu	410
GTG Val	GTG Val	GGT Gly	GCC Ala -190	Glu	CTG Leu	CGC Arg	GTG Val	CTG Leu -18	Arg	CGG Arg	GGA Gly	TCT Ser	CCA Pro -180	Glu	TCG Ser	458
GGC Gly	CCA Pro	GGC Gly -17	Ser	TGG Trp	ACT Thr	TCT Ser	CCG Pro -17	Pro	TTG Leu	CTG Leu	CTG Leu	CTG Leu -16	Ser	ACG Thr	TGC Cys	506
CCG Pro	GGC Gly -16	Ala	GCC Ala	CGA Arg	GCG Ala	CCA Pro -15	Arg	CTG Leu	CTG Leu	TAC Tyr	TCG Ser	Arg	GCA Ala	GCT Ala	GAG Glu	554
CCC Pro -14	Leu	GTC Val	GGT Gly	CAG Gln	CGC Arg	Trp	GAG Glu	GCG Ala	TTC Phe	GAC Asp -13	Val	GCG Ala	GAC Asp	GCC Ala	ATG Met -130	602
AGG Arg	CGC Arg	CAC His	CGT Arg	CGT Arg -12	Glu	CCG Pro	CGC Arg	CCC Pro	CCC Pro -12	Arg	GCG Ala	TTC Phe	TGC Cys	CTC Leu -11	TTG Leu 5	650
CTG Leu	CGC Arg	GCA Ala	GTG Val -11	Ala	GGC Gly	CCG Pro	GTG Val	CCG Pro -10	Ser	CCG Pro	TTG Leu	GCA Ala	CTG Leu -10	Arg	CGA Arg	698
CTG Leu	GGC	TTC Phe -95	GGC Gly	TGG Trp	CCG Pro	GGC Gly	GGA Gly -90	GGG Gly	GGC Gly	TCT Ser	GCG Ala	GCA Ala -85	GAG Glu	GAG Glu	CGC Arg	746
GCG Ala	GTG Val -80	CTA Leu	GTC Val	GTC Val	TCC Ser	TCC Ser -75	CGC Arg	ACG Thr	CAG Gln	AGG Arg	AAA Lys -70	GAG Glu	AGC Ser	TTA Leu	TTC Phe	794
CGG Arg -65	Glu	ATC Ile	CGC Arg	GCC Ala	CAG Gln -60	GCC Ala	CGC Arg	GCG Ala	CTC Leu	GGG Gly -55	GCC Ala	GCT Ala	CTG Leu	GCC Ala	TCA Ser -50	842
GAG Glu	CCG Pro	CTG Leu	CCC Pro	GAC Asp -45	Pro	GGA Gly	ACC Thr	GGC Gly	ACC Thr	GCG Ala	TCG Ser	CCA Pro	AGG Arg	GCA Ala -35	GTC Val	890
ATT Ile	GGC Gly	GGC Gly	CGC Arg -30	Arg	CGG Arg	AGG Arg	AGG Arg	ACG Thr -25	Ala	TTG Leu	GCC Ala	GGG Gly	ACG Thr -20	CGG Arg	ACA Thr	938
GCG Ala	CAG Gln	GGC Gly -15	Ser	GGC Gly	GGG Gly	GGC	GCG Ala -10	Gly	CGG Arg	GGC Gly	CAC His	GGG Gly -5	Arg	AGG Arg	GGC Gly	986
CGG Arg	AGC Ser	Arg	TGC Cys	AGC Ser	CGC Arg	Lys	CCG	TTG Leu	His	GTG Val	Asp	TTC Phe	AAG Lys	GAG Glu	CTC Leu 15	1034

GGC Gly	TGG Trp	GAC Asp	GAC Asp	TGG Trp 20	ATC Ile	ATC Ile	GCG Ala	CCG Pro	CTG Leu 25	GAC Asp	TAC Tyr	GAG Glu	GCG Ala	TAC Tyr 30	CAC His	1082
TGC Cys	GAG Glu	GGC Gly	CTT Leu 35	TGC Cys	GAC Asp	TTC Phe	CCT Pro	TTG Leu 40	CGT Arg	TCG Ser	CAC His	CTC Leu	GAG Glu 45	CCC Pro	ACC Thr	·1130
				ATT Ile												1178
				TGC Cys												1226
				GCC Ala												1274
-				GCC Ala 100					TAGO	CGCG	CGG (GCCG(E GGA(G		1321
GGG	CAGCO	CAC (CGG	CCGAC	G A	rcc										1345

(2) INFORMATION FOR SEQ ID NO:34:

- (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 388 amino acids
 - (B) TYPE: amino acid (D) TOPOLOGY: linear
- (ii) MOLECULE TYPE: protein
- (xi) SEQUENCE DESCRIPTION: SEQ ID NO:34:

Pro Gly Arg Arg Pro Leu Leu Trp Ala Arg Leu Ala Ala Phe Arg -284 -280 -275

Leu Gly Gln Arg Arg Gly Val Gly Arg Trp Leu Gln Gln Ala Trp Leu

Pro His Arg Arg Gln Leu Gly His Leu Leu Gly Gly Pro Ala Leu -245

Thr Val Cys Arg Ile Cys Ser Tyr Thr Ala Leu Ser Leu Cys Pro Cys -230 -225

Arg Ser Pro Ala Asp Glu Ser Ala Ala Glu Thr Gly Gln Ser Phe Leu -220 -215 -205 -210

Phe Asp Val Ser Ser Leu Asn Asp Ala Asp Glu Val Val Gly Ala Glu -200

Leu Arg Val Leu Arg Arg Gly Ser Pro Glu Ser Gly Pro Gly Ser Trp

Thr Ser Pro Pro Leu Leu Leu Ser Thr Cys Pro Gly Ala Ala Arg -170 -165

Ala Pro Arg Leu Leu Tyr Ser Arg Ala Ala Glu Pro Leu Val Gly Gln -150

Arg Trp Glu Ala Phe Asp Val Ala Asp Ala Met Arg Arg His Arg Arg -140 -135 -130 75

Glu Pro Arg Pro Pro Arg Ala Phe Cys Leu Leu Leu Arg Ala Val Ala

- Gly Pro Val Pro Ser Pro Leu Ala Leu Arg Arg Leu Gly Phe Gly Trp -100
- Pro Gly Gly Gly Ser Ala Ala Glu Glu Arg Ala Val Leu Val Val
- Ser Ser Arg Thr Gln Arg Lys Glu Ser Leu Phe Arg Glu Ile Arg Ala
- Gln Ala Arg Ala Leu Gly Ala Ala Leu Ala Ser Glu Pro Leu Pro Asp
- Pro Gly Thr Gly Thr Ala Ser Pro Arg Ala Val Ile Gly Gly Arg Arg
- Arg Arg Arg Thr Ala Leu Ala Gly Thr Arg Thr Ala Gln Gly Ser Gly
- Gly Gly Ala Gly Arg Gly His Gly Arg Arg Gly Arg Ser Arg Cys Ser
- Arg Lys Pro Leu His Val Asp Phe Lys Glu Leu Gly Trp Asp Asp Trp
- Ile Ile Ala Pro Leu Asp Tyr Glu Ala Tyr His Cys Glu Gly Leu Cys
- Asp Phe Pro Leu Arg Ser His Leu Glu Pro Thr Asn His Ala Ile Ile
- Gln Thr Leu Leu Asn Ser Met Ala Pro Asp Ala Ala Pro Ala Ser Cys
- Cys Val Pro Ala Arg Leu Ser Pro Ile Ser Ile Leu Tyr Ile Asp Ala
- Ala Asn Asn Val Val Tyr Lys Gln Tyr Glu Asp Met Val Val Glu Ala

Cys Gly Cys Arg

(2) INFORMATION FOR SEQ ID NO:35:

- (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 17 base pairs

 - (B) TYPE: nucleic acid (C) STRANDEDNESS: single
 - (D) TOPOLOGY: linear
- (ii) MOLECULE TYPE: DNA (genomic)
- (iii) HYPOTHETICAL: NO
- (iv) ANTI-SENSE: NO
- (vi) ORIGINAL SOURCE:
 - (C) INDIVIDUAL ISOLATE: primer number 8
- (xi) SEQUENCE DESCRIPTION: SEQ ID NO:35:

TGTATGCGAC TTCCCGC

What is claimed is:

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1. A DNA molecule comprising an isolated DNA sequence encoding a BMP-12 related protein.

- 5 2. A DNA molecule according to claim 1, wherein said DNA sequence is selected from the group consisting of:
 - (a) nucleotides #496, #571 or #577 to #882 of SEQ ID NO:1;
 - (b) nucleotides #605 or #659 to #964 of SEQ ID NO:25; and
 - (c) sequences which hybridize to (a) or (b) under stringent hybridization conditions and encode a BMP-12 related protein which exhibits the ability to form tendon/ligament-like tissue.
 - 3. A DNA molecule comprising the DNA sequence of claim 1 wherein said DNA sequence is selected from the group consisting of:
 - (a) nucleotides encoding for amino acids #-25, #1 or #3 to #104 of SEQ ID NO:2;
 - (b) in a 5' to 3' direction, nucleotides encoding a propertide selected from the group consisting of native BMP-12 propertide and a BMP protein propertide; and nucleotides encoding for amino acids #-25, #1 or #3 to #104 of SEQ ID NO:2; and
- 20 (c) nucleotides encoding for amino acids #1 or #19 to #120 of SEQ ID NO:26:
 - (d) in a 5' to 3' direction, nucleotides encoding a propeptide selected from the group consisting of native BMP-12 propeptide and a BMP protein propeptide; and nucleotides encoding for amino acids #1 or #19 to #120 of SEQ ID NO:26;
 - (e) sequences which hybridize to any of (a) through (d) under stringent hybridization conditions and encode a BMP-12 related protein which exhibits the ability to form cartilage and/or bone.
 - 4. A host cell transformed with a DNA molecule according to claim 1.
 - 5. A host cell transformed with the DNA molecule of claim 2.
 - 6. A host cell transformed with the DNA molecule of claim 3.

7. An isolated DNA molecule having a sequence encoding a BMP-12 protein which is characterized by the ability to induce the formation of tendon/ligament-like tissue, said DNA molecule comprising a DNA sequence selected from the group consisting of:

- 5
- (a) nucleotide #496, #571 or #577 to #882 of SEQ ID NO:1;
- (b) nucleotide #605 or #659 to #964 of SEQ ID NO:25; and
- (c) naturally occurring allelic sequences and equivalent degenerative codon sequences of (a) or (b).
 - 8. A host cell transformed with the DNA molecule of claim 7.
- 10
- 9. A vector comprising a DNA molecule of claim 7 in operative association with an expression control sequence therefor.
 - 10. A host cell transformed with the vector of claim 9.
- 11. A method for producing a purified BMP-12 protein, said method comprising the steps of:
- 15
 - (a) culturing a host cell transformed with a DNA molecule according to claim 2, comprising a nucleotide sequence encoding a BMP-12 related protein; and
 - (b) recovering and purifying said BMP-12 related protein from the culture medium.
- 20
- 12. A method for producing a purified BMP-12 related protein said method comprising the steps of:
- (a) culturing a host cell transformed with a DNA molecule according to claim 3, comprising a nucleotide sequence encoding a BMP-12 related protein; and
- 25 (b) recovering and purifying said BMP-12 related protein from the culture medium.
 - 13. A method for producing a purified BMP-12 related protein said method comprising the steps of:
- (a) culturing a host cell transformed with a DNA molecule according to claim 7, comprising a nucleotide sequence encoding a BMP-12 related protein; and

(b) recovering and purifying said BMP-12 related protein from the culture medium.

- 14. A purified polypeptide comprising an amino acid sequence selected from the following group:
- 5 (a) from amino acid #-25 to amino acid #104 as set forth in SEQ ID NO:2;
 - (b) from amino acid #1 to amino acid #104 as set forth in SEQ ID NO:2.
 - (c) from amino acid #3 to amino acid #104 as set forth in SEQ ID NO:2.
- (d) from amino acid #1 to amino acid #120 as set forth in SEQ ID NO:26; and
 - (d) from amino acid #19 to amino acid #120 as set forth in SEQ ID NO:26.
 - 15. A purified polypeptide wherein said polypeptide is in the form of a dimer comprised of two subunits, each with the amino acid sequence of claim 14.
 - 16. A purified protein produced by the steps of

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- (a) culturing a cell transformed with a DNA molecule comprising the nucleotide sequence from nucleotide #496, #571 or #577 to #882 as shown in SEQ ID NO:1; and
- 20 (b) recovering and purifying from said culture medium a protein comprising the amino acid sequence from amino acid #-25, amino acid #1 or amino acid #3 to amino acid #104 as shown in SEQ ID NO:2.
 - 17. A purified BMP-12 related protein characterized by the ability to induce the formation of tendon/ligament-like tissue.
- 25 18. A pharmaceutical composition comprising an effective amount of the BMP-12 related protein of claim 17 in admixture with a pharmaceutically acceptable vehicle.
 - 19. A method for inducing tendon/ligament-like tissue formation in a patient in need of same comprising administering to said patient an effective amount of the composition of claim 18.

20. A pharmaceutical composition for tendon/ligament-like tissue healing and tissue repair said composition comprising an effective amount of the protein of a BMP-12 related protein in a pharmaceutically acceptable vehicle.

- 21. A method for treating tendinitis, or other tendon or ligament defect in a patient in need of same, said method comprising administering to said patient an effective amount of the composition of claim 20.
- 22. A chimeric DNA molecule comprising a DNA sequence encoding a propertide from a member of the TGF- β superfamily of proteins linked in correct reading frame to a DNA sequence encoding a BMP-12 related polypeptide.
- 23. A chimeric DNA molecule according to claim 22, wherein the propeptide is the propeptide from BMP-2.
- 24. A heterodimeric protein molecule comprising one monomer having the amino acid sequence of the polypeptide of claim 14, and one monomer having the amino acid sequence of a protein of the $TGF-\beta$ superfamily.
- 25. A method for inducing tendon/ligament-like tissue formation in a patient in need of same comprising administering to said patient an effective amount of a composition comprising a protein encoded by a DNA sequence selected from the group consisting of:

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- (a) nucleotides #496, #571 or #577 to #882 of SEQ ID NO:1;
- (b) nucleotides #845 or #899 to #1204 of SEQ ID NO:3;
- (c) nucleotides #605 or #659 to #964 of SEQ ID NO:25; and
- (d) sequences which hybridize to (a), (b) or (c) under stringent hybridization conditions and encode a protein which exhibits the ability to form tendon/ligament-like tissue.

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26. A method for inducing tendon/ligament-like tissue formation in a patient in need of same comprising administering to said patient an effective amount of the composition comprising a tendon/ligament-like tissue inducing protein having an amino acid sequence selected from the group consisting of:

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- (a) amino acids #-25, #1 or #3 to #104 of SEQ ID NO:2;
- (b) amino acids #1 or #19 to #120 of SEQ ID NO:4;
- (c) amino acids #1 or #19 to #120 of SEQ ID NO:26; and

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(d) mutants and/or variants of (a), (b) or (c) which exhibit the ability to form tendon and/or ligament.

- 27. A pharmaceutical composition for tendon/ligament-like tissue repair, said composition comprising an effective amount of a BMP-12 related protein in a pharmaceutically acceptable vehicle.
- 28. A method for treating tendinitis, or other tendon or ligament defect in a patient in need of same, said method comprising administering to said patient an effective amount of the composition of claim 27.

FIG 1/1
COMPARISON OF HUMAN V1-1 VS. HUMAN MP-52

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V1-1 Ser Arg Cys Ser Arg Lys Pro Leu His Val Asp Phe Lys Glu Leu
1 AGC CGC TGC AGC CGC AAG CCG TTG CAC GTG GAC TTC AAG GAG CTC
MP52 GCT CGC TGC AGT CGG AAG GCA CTG CAT GTC AAC TTC AAG GAC ATG
  1 Ala Arg Cys Ser Arg Lys Ala Leu His Val Asn Phe Lys Asp Met
     Gly Trp Asp Asp Trp Ile Ile Ala Pro Leu Asp Tyr Glu Ala Tyr
     GGC TGG GAC GAC TGG ATC ATC GCG CCG CTG GAC TAC GAG GCG TAC
     GGC TGG GAC GAC TGG ATC ATC GCA CCC CTT GAG TAC GAG GCT TTC
     Gly Trp Asp Asp Trp Ile Ile Ala Pro Leu Glu Tyr Glu Ala Phe
     His Cys Glu Gly Leu Cys Asp Phe Pro Leu Arg Ser His Leu Glu CAC TGC GAG GGC CTT TGC GAC TTC CCT TTG CGT TCG CAC CTC GAG
 31
     His Cys Glu Gly Leu Cys Glu Phe Pro Leu Arg Ser His Leu Glu
     Pro Thr Asn His Ala Ile Ile Gln Thr Leu Leu Asn Ser Met Ala
121
     CCC ACC AAC CAT GCC ATC ATT CAG ACG CTG CTC AAC TCC ATG GCA
     Pro Thr Asn His Ala Val Ile Gln Thr Leu Met Asn Ser Met Asp
     Pro Asp Ala Ala Pro Ala Ser Cys Cys Val Pro Ala Arg Leu Ser CCA GAC GCG GCG CCG GCC TCC TGC TGT GTG CCA GCG CGC CTC AGC
     CCC GAG TCC ACA CCA CCC ACC TGC TGT GTG CCC ACG CGG CTG AGT
181
     Pro Glu Ser Thr Pro Pro Thr Cys Cys Val Pro Thr Arg Leu Ser
 76
     Pro Ile Ser Ile Leu Tyr Ile Asp Ala Ala Asn Asn Val Val Tyr
     CCC ATC AGC ATC CTC TAC ATC GAC GCC GCC AAC AAC GTT GTC TAC
226
     Pro Ile Ser Ile Leu Phe Ile Asp Ser Ala Asn Asn Val Val Tyr
     Lys Gln Tyr Glu Asp Met Val Val Glu Ala Cys Gly Cys Arg
     AAG CAA TAC GAG GAC ATG GTG GTG GAG GCC TGC GGC TGC AGG
     AAG CAG TAT GAG GAC ATG GTC GTG GAG TCG TGT GGC TGC AGG
 91 Lys Gln Tyr Glu Asp Met Val Val Glu Ser Cys Gly Cys Arg
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Homology at the nucleotide level: 249/312 = 79.8% Homology at the amino acid level: 84/104 = 80.8%

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(51) International Patent Classis			(11) International Publication Number:	WO 95/16035
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(21) International Application N (22) International Filing Date:	Number: PCT/US 6 December 1994 (RU, European patent (AT, BE,	CH, DE, DK, ES, FR, GB, SE), OAPI patent (BF, BJ,
08/217,780 25 08/333,576 2 N	OWS OF HARVARD Coet, Cambridge, MA 0213 Ony, J.; 86 Packard Street, EY, John, M.; 59 Old Bolo. ROSEN, Vicki, A.; 12 02146 (US). WOLFMAPER, MA 02030 (US). TH Terrace, Port Jefferson, No. A.; 22 Slocum Road, L. R.; Genetics Institute,	/US]; { ## (US); { ##	With international search report Before the expiration of the tirclaims and to be republished in amendments. (88) Date of publication of the internation	ne limit for amending the the event of the receipt of

(54) Title: BMP-12, BMP-13 AND TENDON-INDUCING COMPOSITIONS THEREOF

(57) Abstract

Bone morphogenetic proteins BMP-12 and BMP-13 have been cloned. Compositions of these proteins with tendon/ligament-like tissue inducing activity are disclosed. The compositions are useful in the treatment of tendinitis and tendon or ligament defects and in related tissue repair.

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FR	France	MN	Mongolia	VN	Vict Nam
GA	Gabon				

INTERNATIONAL SEARCH REPORT

Inter nal Application No
PCT/US 94/14030

A. CLASSIFICATION OF SUBJECT MATTER
IPC 6 C12N15/12 C12N15/70 C12N15/62 C12N1/21 C07K14/51 A61K38/17 According to International Patent Classification (IPC) or to both national classification and IPC **B. FIELDS SEARCHED** Minimum documentation searched (classification system followed by classification symbols) IPC 6 C12N C07K A61K Documentation searched other than minimum documentation to the extent that such documents are included in the fields searched Electronic data base consulted during the international search (name of data base and, where practical, search terms used) C. DOCUMENTS CONSIDERED TO BE RELEVANT Category * Citation of document, with indication, where appropriate, of the relevant passages Relevant to claim No. X WO,A,93 16099 (BIOPHARM GESSELLSCHAFT ZUR 25,26 BIOTECHNOLOGISCHEN ENTWICKLUNG VON PHARMAKA) 19 August 1993 cited in the application see page 4, paragraph 3 see page 7, paragraph 3 see page 9, paragraph 2 WO, A, 91 18047 (GENENTECH, INC.) 28 22,23 November 1991 cited in the application see page 4, line 4 - line 22 WO, A, 93 00432 (GENETICS INSTITUTE, INC.) 7 1-28 January 1993 cited in the application see page 4, line 21 - line 33 see page 8, line 15 - page 10, line 2 Patent family members are listed in annex. Further documents are listed in the continuation of box C. X * Special categories of cited documents: T later document published after the international filing date or priority date and not in conflict with the application but cited to understand the principle or theory underlying the "A" document defining the general state of the art which is not considered to be of particular relevance invention "E" earlier document but published on or after the international "X" document of particular relevance; the claimed invention cannot be considered novel or cannot be considered to involve an inventive step when the document is taken alone filing date "L" document which may throw doubts on priority claim(s) or which is cited to establish the publication date of another citation or other special reason (as specified) "Y" document of particular relevance; the claimed invention cannot be considered to involve an inventive step when the document is combined with one or more other such docu-ments, such combination being obvious to a person skilled in the art. "O" document referring to an oral disclosure, use, exhibition or document published prior to the international filing date but later than the priority date claimed '&' document member of the same patent family Date of the actual completion of the international search Date of mailing of the international search report 19 May 1995 Authorized officer Name and mailing address of the ISA European Patent Office, P.B. 5818 Patentlaan 2 NL - 2280 HV Rijswijk Td. (+31-70) 340-2040, Tx. 31 651 epo nl, Montero Lopez, B Fax: (+31-70) 340-3016

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INTERNATIONAL SEARCH REPORT

ernational application No.

PCT/US 94/14030

Box I	Observations where certain claims were found unsearchable (Continuation of item 1 of first sheet)
This inte	ernational search report has not been established in respect of certain claims under Article 17(2)(a) for the following reasons:
1. X	Claims Nos.: because they relate to subject matter not required to be searched by this Authority, namely: Remark: Although claims 19,21,25,26 and 28 are directed to a mehtod of treatment of the human/animal body the search has been carried out and based on the alleged effects of the composition.
2.	Claims Nos.: because they relate to parts of the international application that do not comply with the prescribed requirements to such an extent that no meaningful international search can be carried out, specifically:
3.	Claims Nos.: because they are dependent claims and are not drafted in accordance with the second and third sentences of Rule 6.4(a).
Box II	Observations where unity of invention is lacking (Continuation of item 2 of first sheet)
This Int	ernational Searching Authority found multiple inventions in this international application, as follows:
1.	As all required additional search fees were timely paid by the applicant, this international search report covers all searchable claims.
2.	As all searchable claims could be searches without effort justifying an additional fee, this Authority did not invite payment of any additional fee.
3.	As only some of the required additional search fees were timely paid by the applicant, this international search report covers only those claims for which fees were paid, specifically claims Nos
4.	No required additional search fees were timely paid by the applicant. Consequently, this international search report is restricted to the invention first mentioned in the claims; it is covered by claims Nos.:
Remari	The additional search fees were accompanied by the applicant's protest. No protest accompanied the payment of additional search fees.

INTERNATIONAL SEARCH REPORT

information on patent family members

Inten sal Application No
PCT/US 94/14030

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(21) International Application Number: PCT/US			(81) Designated States: AU, CA, CN, RU, European patent (AT, BE,	CH, DE, DK, ES, FR, GB,				
(22) International Filing Date: 6 December 1994 ((06.12.9	SE), OAPI patent (BF, BJ, , MR, NE, SN, TD, TG).						
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CambridgePark Drive, Cambridge, MA 0214 PRESIDENT AND FELLOWS OF HARVARD C [US/US]; 17 Quincy Street, Cambridge, MA 0213	OLLEC	JE	(88) Date of publication of the intern	ational search report: 13 July 1995 (13.07.95)				
(72) Inventors: CELESTE, Anthony, J.; 86 Packard Street MA 01749 (US). WOZNEY, John, M.; 59 Old Bol Hudson, MA 01749 (US). ROSEN, Vicki, A.; 12 Road #7, Brookline, MA 02146 (US). WOLFML M.; 30 Rolling Lane, Dover, MA 02030 (US). TH Gerald, H.; 201 Bayview Terrace, Port Jefferson, N (US). MELTON, Douglas, A.; 22 Slocum Road, L MA 02173 (US).	ton Ros 7 Kilsy AN, No IOMSE NY 117	ed, /th :il, N, 77						
(74) Agent: LAZAR, Steven, R.; Genetics Institute, CambridgePark Drive, Cambridge, MA 02140 (US		87						
(54) Title: BMP-12, BMP-13 AND TENDON-INDUCIN	IG COM	/IPOS	SITIONS THEREOF					

(57) Abstract

Bone morphogenetic proteins BMP-12 and BMP-13 have been cloned. Compositions of these proteins with tendon/ligament-like tissue inducing activity are disclosed. The compositions are useful in the treatment of tendimitis and tendon or ligament defects and in related tissue repair.

^{* (}Referred to in PC1 Gazette No. 39/1995, Section II) .

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TITLE OF THE INVENTION

BMP-12, BMP-13 AND TENDON-INDUCING COMPOSITIONS THEREOF

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RELATED APPLICATIONS

The present invention is a continuation-in-part of application serial number 08/217,780, filed March 25, 1994, 08/164,103, filed on December 7, 1993 and 08\333,576, filed on November 2, 1994.

FIELD OF THE INVENTION

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The present invention relates to a novel family of purified proteins, and compositions containing such proteins, which compositions are useful for the induction of tendon/ligament-like tissue formation, wound healing and ligament and other tissue repair. These proteins may also be used in compositions for augmenting the activity of bone morphogenetic proteins.

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BACKGROUND OF THE INVENTION

The search for the molecule or molecules responsible for formation of bone, cartilage, tendon and other tissues present in bone and other tissue extracts has led to the discovery of a novel set of molecules called the Bone Morphogenetic Proteins (BMPs). The structures of several proteins, designated BMP-1 through BMP-11, have previously been elucidated. The unique inductive activities of these proteins, along with their presence in bone, suggests that they are important regulators of bone repair processes, and may be involved in the normal maintenance of bone tissue. There is a need to identify additional proteins which play a role in forming other vital tissues. The present invention relates to the identification of a family of proteins, which have tendon/ligament-like tissue inducing activity, and which are useful in compositions for the induction of tendon/ligament-like tissue formation and repair.

SUMMARY OF THE INVENTION

In one embodiment, the present invention comprises DNA molecules encoding a tendon/ligament-like inducing protein which the inventors have named V1-1. This novel protein is now called BMP-12. The present invention also includes DNA molecules encoding BMP-12 related proteins.

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BMP-12 related proteins are a subset of the BMP/TGF-β/Vg-1 family of proteins, including BMP-12 and VL-1, which are defined as tendon/ligament-like tissue inducing proteins encoded by DNA sequences which are cloned and identified, e.g., using PCR, using BMP-12 specific primers, such as primers #6 and #7 described below, with reduced stringency conditions. It is preferred that the DNA sequences encoding BMP-12 related proteins share at least about 80% homology at the amino acid level from amino acids with amino acids #3 to #103 of SEQ ID NO:1.

The DNA molecules preferably have a DNA sequence encoding the BMP-12 protein, the sequence of which is provided in SEQ ID NO:1, or a BMP-12 related protein as further described herein. Both the BMP-12 protein and BMP-12 related proteins are characterized by the ability to induce the formation of tendon/ligament-like tissue in the assay described in the examples.

The DNA molecules of the invention preferably comprise a DNA sequence, as described in SEQUENCE ID NO:1; more preferably nucleotides #496 to #882, #571 to #882 or #577 to #882 of SEQ ID NO:1; or DNA sequences which hybridize to the above under stringent hybridization conditions and encode a protein which exhibits the ability to form tendon/ligament-like tissue. The DNA molecules of the invention may also comprise a DNA sequence as described in SEQ ID NO:25; more preferably nucleotides #604 or #658 to #964 of SEQ ID NO:25.

The DNA molecules of the invention also include DNA molecules comprising a DNA sequence encoding a BMP-12 related protein with the amino acid sequence shown in SEQ ID NO:2 or SEQ ID NO:26, as well as naturally occurring allelic sequences and equivalent degenerative codon sequences of SEQ ID NO:2 or SEQ ID NO:26. Preferably, the DNA sequence of the present invention encodes amino acids #-25 to # 104, #1 to # 104 or #3 to #103 of SEQ ID NO:2; or amino acids #1 to #120 or #19 to #120 of SEQ ID NO:26. The DNA sequence may comprise, in a 5' to 3' direction, nucleotides encoding a propeptide, and nucleotides encoding for amino acids #-25 to #104, #1 to #104 or #3 to #103 of SEQ ID NO:2; or amino acids #1 to #120 or #19 to #120 of SEQ ID NO:26. The propeptide useful in the above embodiment is preferably selected from the group consisting of native BMP-12 propeptide and a protein propeptide from a different member of the TGF-B

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superfamily or BMP family. The invention further comprises DNA sequences which hybridize to the above DNA sequences under stringent hybridization conditions and encode a BMP-12 related protein which exhibits the ability to induce formation of tendon/ligament-like tissue.

In other embodiments, the present invention comprises host cells and vectors which comprise a DNA molecule encoding the BMP-12 protein, or a BMP-12 related protein. The host cells and vectors may further comprise the coding sequence in operative association with an expression control sequence therefor.

In another embodiment, the present invention comprises a method for producing a purified BMP-12 related protein, said method comprising the steps of culturing a host cell transformed with the above DNA molecule or vector comprising a nucleotide sequence encoding a BMP-12 related protein; and (b) recovering and purifying said BMP-12 related protein from the culture medium. In a preferred embodiment, the method comprises (a) culturing a cell transformed with a DNA molecule comprising the nucleotide sequence from nucleotide #496, #571 or #577 to #879 or #882 as shown in SEQ ID NO:1; or the nucleotide sequence from #604 or #658 to #963 of SEQ ID NO:25; and

(b) recovering and purifying from said culture medium a protein comprising the amino acid sequence from amino acid #-25, #1 or #3 to amino acid #103 or #104 as shown in SEQ ID NO:2; or from amino acid #1 or #19 to amino acid #120 as shown in SEQ ID NO:26. The present invention also includes a purified protein produced by the above methods.

The present invention further comprises purified BMP-12 related protein characterized by the ability to induce the formation of tendon/ligament-like tissue. The BMP-12 related polypeptides preferably comprise an amino acid sequence as shown in SEQ ID NO:2. The polypeptide more preferably comprise amino acids #-25, #1 or #3 to #103 or #104 as set forth in SEQ ID NO:2; or amino acids #1 or #19 to #120 as set forth in SEQ ID NO:26. In a preferred embodiment, the purified polypeptide may be in the form of a dimer comprised of two subunits, each with the amino acid sequence of SEQ ID NO:2.

In another embodiment, the present invention comprises compositions comprising an effective amount of the above-described BMP-12 related proteins.

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In the compositions, the protein may be admixed with a pharmaceutically acceptable vehicle.

The invention also includes methods for tendon/ligament-like tissue healing and tissue repair, for treating tendinitis, or other tendon or ligament defects, and for inducing tendon/ligament-like tissue formation in a patient in need of same, comprising administering to said patient an effective amount of the above composition.

Other embodiments include chimeric DNA molecules comprising a DNA sequence encoding a propertide from a member of the TGF- β superfamily of proteins linked in correct reading frame to a DNA sequence encoding a BMP-12 related polypeptide. One suitable propertide is the propertide from BMP-2. The invention also includes heterodimeric protein molecules comprising one monomer having the amino acid sequence shown in SEQ ID NO:2, and one monomer having the amino acid sequence of another protein of the TGF- β subfamily.

Finally, the present invention comprises methods for inducing tendon/ligament-like tissue formation in a patient in need of same comprising administering to said patient an effective amount of a composition comprising a protein which exhibits the ability to induce formation of tendon/ligament-like tissue, said protein having an amino acid sequence shown in SEQ ID NO:2 or SEQ ID NO:4 or SEQ ID NO:26. The amino acid sequences are more preferably one of the following: (a) amino acids #-25, #1 or #3 to #103 or #104 of SEQ ID NO:2; (b) amino acids #1 or #19 to #119 or #120 of SEQ ID NO:4; (c) amino acids #1 or #19. to #119 or #120 of SEQ ID NO:26; (d) mutants and/or variants of (a), (b) or (c) which exhibit the ability to form tendon and/or ligament. In other embodiments of the above method, the protein is encoded by a DNA sequence of SEQ ID NO:1, SEQ ID NO:3 or SEQ ID NO:25, more preferably one of the following: (a) nucleotides #496, #571 or #577 to #879 or #882 of SEQ ID NO:1; (b) nucleotides #845 or #899 to #1201 or #1204 of SEQ ID NO:3; (c) nucleotides #605 or #659 to #961 or #964 of SEQ ID NO:25; and (d) sequences which hybridize to (a) or (b) under stringent hybridization conditions and encode a protein which exhibits the ability to form tendon/ligament-like tissue.

Description of the Sequences

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SEQ ID NO:1 is the nucleotide sequence encoding the human BMP-12.

SEQ ID NO:2 is the amino acid sequence comprising the mature human BMP-12 polypeptide.

SEQ ID NO:3 is the nucleotide sequence encoding the protein MP52.

SEQ ID NO:4 is the amino acid sequence comprising the mature MP52 polypeptide.

SEQ ID NO:5 is the nucleotide sequence of a specifically amplified portion of the human BMP-12 encoding sequence.

SEQ ID NO:6 is the amino acid sequence encoded by the nucleotide sequence of SEQ ID NO:5.

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SEQ ID NO:7 is the nucleotide sequence of a specifically amplified portion of the human VL-1 encoding sequence.

SEQ ID NO:8 is the amino acid sequence encoded by the nucleotide sequence of SEQ ID NO:7.

SEQ ID NO:9 is the nucleotide sequence of the plasmid pALV1-781, used for expression of BMP-12 in E. coli.

SEQ ID NO:10 is the nucleotide sequence of a fragment of the murine clone, mV1.

SEQ ID NO:11 is the amino acid sequence of a fragment of the murine protein encoded by mV1.

SEQ ID NO:12 is the nucleotide sequence of a fragment of the murine clone, mV2.

SEQ ID NO:13 is the amino acid sequence of a fragment of the murine protein encoded by mV2.

SEQ ID NO:14 is the nucleotide sequence of a fragment of the murine clone, mV9.

SEQ ID NO:15 is the amino acid sequence of a fragment of the murine protein encoded by mV9.

SEQ ID NO:16 is the amino acid sequence of a BMP/TGF-β/Vg-1 protein consensus sequence. The first Xaa represents either Gln or Asn; the second Xaa represents either Val or Ile.

SEQ ID NO:17 is the nucleotide sequence of oligonucleotide #1.

SEQ ID NO:18 is the amino acid sequence of a BMP/TGF- β /Vg-1 protein consensus sequence. The Xaa represents either Val or Leu.

SEQ ID NO:19 is the nucleotide sequence of oligonucleotide #2.

SEQ ID NO:20 is the nucleotide sequence of oligonucleotide #3.

SEQ ID NO:21 is the nucleotide sequence of oligonucleotide #4.

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SEQ ID NO:22 is the nucleotide sequence of oligonucleotide #5

SEQ ID NO:23 is the nucleotide sequence of oligonucleotide #6.

SEQ ID NO:24 is the nucleotide sequence of oligonucleotide #7.

SEQ ID NO:25 is the nucleotide sequence of the human VL-1 (BMP-13) encoding sequence.

SEQ ID NO:26 is the amino acid sequence encoded by the nucleotide sequence of SEQ ID NO:25.

SEQ ID NO:27 is the nucleotide sequence encoding a fusion of BMP-2 propeptide and the mature coding sequence of BMP-12.

SEQ ID NO:28 is the amino acid sequence encoded by the nucleotide sequence of SEQ ID NO:27.

SEQ ID NO:29 is the nucleotide sequence encoding the murine mV1 protein. X01 is Val, Ala, Glu or Gly; X02 is Ser, Pro Thr or Ala; X03 is Ser or Arg; X04 is Leu, Pro, Gln or Arg; X05 is Cys or Trp; X06 is Val, Ala, Asp or Gly; X07 is Val, Ala, Glu or Gly; X08 is Gln, Lys or Glu.

SEQ ID NO:30 is the amino acid sequence encoded by the nucleotide sequence of SEQ ID NO:29. X01 through X08 are the same as in SEQ ID NO: 29.

SEQ ID NO:31 is the nucleotide sequence encoding the murine mV2 protein. X01 is Pro or Thr; X02 is Val.

SEQ ID NO:32 is the amino acid sequence encoded by the nucleotide sequence of SEQ ID NO:31. X01 and X02 are the same as in SEQ ID NO:31.

SEQ ID NO:33 is the nucleotide sequence encoding human BMP-12 protein.

SEQ ID NO:34 is the amino acid sequence encoded by the nucleotide sequence of SEQ ID NO:33.

SEQ ID NO:35 is the nucleotide sequence of oligonucleotide #8.

Brief Description of the Figures

Figure 1 is a comparison of the human BMP-12 and human MP52 sequences.

Detailed Description of the Invention

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The DNA sequences of the present invention are useful for producing proteins which induce the formation of tendon/ ligament-like tissue, as described further below. The DNA sequences of the present invention are further useful for isolating and cloning further DNA sequences encoding BMP-12 related proteins with similar activity. These BMP-12 related proteins may be homologues from other species, or may be related proteins within the same species.

Still, a further aspect of the invention are DNA sequences coding for expression of a tendon/ligament-like tissue inducing protein. Such sequences include the sequence of nucleotides in a 5' to 3' direction illustrated in SEQ ID NO:1 or SEQ ID NO:25, DNA sequences which, but for the degeneracy of the genetic code, are identical to the DNA sequence SEQ ID NO:1 or 25, and encode the protein of SEQ ID NO:2 or 26. Further included in the present invention are DNA sequences which hybridize under stringent conditions with the DNA sequence of SEQ ID NO:1 or 25 and encode a protein having the ability to induce the formation of tendon or ligament. Preferred DNA sequences include those which hybridize under stringent conditions as described in Maniatis et al, Molecular Cloning (A Laboratory Manual), Cold Spring Harbor Laboratory (1982), pages 387 to 389. Finally, allelic or other variations of the sequences of SEQ ID NO:1 or 25, whether such nucleotide changes result in changes in the peptide sequence or not, but where the peptide sequence still has tendon/ligament-like tissue inducing activity, are also included in the present invention.

The human BMP-12 DNA sequence (SEQ ID NO:1) and amino acid sequence (SEQ ID NO:2) are set forth in the Sequence Listings. Another protein that is useful for the compositions and methods of the present invention is VL-1. VL-1 is a BMP-12 related protein which was cloned using sequences from BMP-12. The inventors have now designated VL-1 as BMP-13. A partial DNA sequence of VL-1 (SEQ ID NO:7) and the encoded amino acid sequence (SEQ ID NO:8); as well as a DNA sequence encoding the mature VL-1 (SEQ ID NO:25) and the encoded amino acid sequence (SEQ ID NO:26) are set forth in the Sequence Listings. Although further descriptions are made with reference to the BMP-12 sequence of SEQ ID NO:1 and 2, it will be recognized that the invention includes similar modifications and

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improvements which may be made to other BMP-12 related sequences, such as the VL-1 sequence shown in SEQ ID NO:25 and 26.

The sequence of BMP-12 shown in SEQ ID NO. 1 includes the entire mature sequence and approximately 190 amino acids of the propeptide. The coding sequence of the mature human BMP-12 protein appears to begin at nucleotide #496 or #571 and continues through nucleotide #882 of SEQ ID NO:1. The first cysteine in the seven cysteine structure characteristic of TGF-β proteins begins at nucleotide #577. The last cysteine ends at #879. Thus, it is expected that DNA sequences encoding active BMP-12 species will comprise nucleotides #577 to #879 of SEQ ID NO:1.

It is expected that BMP-12, as expressed by mammalian cells such as CHO cells, exists as a heterogeneous population of active species of BMP-12 protein with varying N-termini. It is expected that all active species will contain the amino acid sequence beginning with the cysteine residue at amino acid #3 of SEQ ID NO:2 and continue through at least the cysteine residue at amino acid 103 or until the stop codon after amino acid 104. Other active species contain additional amino acid sequence in the N-terminal direction. As described further herein, the N-termini of active species produced by mammalian cells are expected to begin after the occurrence of a consensus cleavage site, encoding a peptide sequence Arg-X-X-Arg. Thus, it is expected that DNA sequences encoding active BMP-12 proteins will have a nucleotide sequence comprising the nucleotide sequence beginning at any of nucleotides #196, 199, 208, 217, 361, 388, 493, 496 or 571 to nucleotide #879 or 882 of SEQ ID NO:1.

The N-terminus of one active species of human BMP-12 has been experimentally determined by expression in E. coli to be as follows: [M]SRXSRKPLHVDF, wherein X designates an amino acid residue with no clear signal, which is consistent with a cysteine residue at that location. Thus, it appears that the N-terminus of this species of BMP-12 is at amino acid #1 of SEQ ID NO:1, and a DNA sequence encoding said species of BMP-12 would start at nucleotide #571 of SEQ ID NO:1. The apparent molecular weight of this species of human BMP-12 dimer was determined by SDS-PAGE to be approximately 20-22 kd on a

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Novex 16% tricine gel. The human BMP-12 protein exists as a clear, colorless solution in 0.1% trifluoroacetic acid.

As described earlier, BMP-12 related proteins are a subset of the BMP/TGF- β /Vg-1 family of proteins, including BMP-12 and VL-1, which can be defined as tendon/ligament-like tissue inducing proteins encoded by DNA sequences which can be cloned and identified, e.g., using PCR, using BMP-12 specific primers, such as primers #6 and #7 described below, with reduced stringency conditions. It is preferred that DNA sequences of the present invention share at least about 80% homology at the amino acid level from amino acids with the DNA encoding amino acids #3 to #103 of SEQ ID NO:1. For the purposes of the present invention, the term BMP-12 related proteins does not include the human MP52 protein. Using the sequence information of SEQ ID NO:1 and SEQ ID NO:3, and the comparison provided in Figure 1, it is within the skill of the art to design primers to the BMP-12 sequence which will allow for the cloning of genes encoding BMP-12 related proteins.

One example of the BMP-12-related proteins of the present invention is VL-1, presently referred to as BMP-13. The sequence of the full mature BMP-13 sequence and at least a part of the propeptide of BMP-13 is given in SEQ ID NO:25. Like BMP-12, it is expected that BMP-13, as expressed by mammalian cells such as CHO cells, exists as a heterogeneous population of active species of BMP-13 protein with varying N-termini. It is expected that all active species will contain the amino acid sequence beginning with the cysteine residue at amino acid #19 of SEQ ID NO:26 and continue through at least the cysteine residue at amino acid 119 or until the stop codon after amino acid 120. Other active species contain additional amino acid sequence in the N-terminal direction. As described further herein, the N-termini of active species produced by mammalian cells are expected to begin after the occurrence of a consensus cleavage site, encoding a peptide sequence Arg-X-X-Arg. Thus, it is expected that DNA sequences encoding active BMP-13 proteins will have a nucleotide sequence comprising the nucleotide sequence beginning at any of nucleotides #410, 458, 602, 605 or 659, to nucleotide #961 or 964 of SEQ ID NO:25.

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In order to produce the purified tendon/ligament-like tissue inducing proteins useful for the present invention, a method is employed comprising culturing a host cell transformed with a DNA sequence comprising a suitable coding sequence, particularly the DNA coding sequence from nucleotide #496, #571 or #577 to #879 or #882 of SEQ ID NO:1; and recovering and purifying from the culture medium a protein which contains the amino acid sequence or a substantially homologous sequence as represented by amino acids #-25, #1 or #3 to #103 or #104 of SEQ ID NO:2. In another embodiment, the method employed comprises culturing a host cell transformed with a DNA sequence comprising a suitable coding sequence, particularly the DNA coding sequence from nucleotide #605 or #659 to #961 or #964 of SEQ ID NO:25; and recovering and purifying from the culture medium a protein which contains the amino acid sequence or a substantially homologous sequence as represented by amino acids #1 or #19 to #119 or #120 of SEQ ID NO:26.

The human MP52 DNA is described in WO93/16099, the disclosure of which is incorporated herein by reference. However, this document does not disclose the ability of the protein to form tendon/ligament-like tissue, or its use in compositions for induction of tendon/ligament-like tissue. Human MP52 was originally isolated using RNA from human embryo tissue. The human MP52 nucleotide sequence (SEQ ID NO:3) and the encoded amino acid sequences (SEQ ID NO:4) are set forth in the Sequence Listings herein. The MP52 protein appears to begin at nucleotide #845 of SEQ ID NO:3 and continues through nucleotide #1204 of SEQ ID NO:3. The first cysteine of the seven cysteine structure characteristic of TGF-β proteins begins at nucleotide #899. The last cysteine ends at #1201. Other active species of MP52 protein may have additional nucleotides at the N-terminal direction from nucleotide #845 of SEQ ID NO:3.

Purified human MP52 proteins of the present invention may be produced by culturing a host cell transformed with a DNA sequence comprising the DNA coding sequence of SEQ ID NO:3 from nucleotide #845 to #1204, and recovering and purifying from the culture medium a protein which contains the amino acid sequence or a substantially homologous sequence as represented by amino acids #1 to #120 of SEQ ID NO:4. It is also expected that the amino acid sequence from amino acids

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#17 or #19 to #119 or #120 of SEQ ID NO:4 will retain activity. Thus, the DNA sequence from nucleotides #845, #893 or #899 to #1201 or #1204 are expected to encode active proteins.

For expression of the protein in mammalian host cells, the host cell is transformed with a coding sequence encoding a propertide suitable for the secretion. of proteins by the host cell is linked in proper reading frame to the coding sequence for the mature protein. For example, see United States Patent 5,168,050, the disclosure of which is hereby incorporated by reference, in which a DNA encoding a precursor portion of a mammalian protein other than BMP-2 is fused to the DNA encoding a mature BMP-2 protein. Thus, the present invention includes chimeric DNA molecules comprising a DNA sequence encoding a propeptide from a member of the TGF- β superfamily of proteins, is linked in correct reading frame to a DNA sequence encoding a tendon/ligament-like tissue inducing polypeptide. The term "chimeric" is used to signify that the propeptide originates from a different polypeptide than the encoded mature polypeptide. Of course, the host cell may be transformed with a DNA sequence coding sequence encoding the native propeptide linked in correct reading frame to a coding sequence encoding the mature protein shown in SEQ ID NO:2, SEQ ID NO:4, or SEQ ID NO:26. The full sequence of the native propertide may be determined through methods known in the art using the sequences disclosed in SEQ ID NO:1, SEQ ID NO:3, or SEO ID NO:25 to design a suitable probe for identifying and isolating the entire clone.

The present invention also encompasses the novel DNA sequences, free of association with DNA sequences encoding other proteinaceous materials, and coding for expression of tendon/ligament-like tissue inducing proteins. These DNA sequences include those depicted in SEQ ID NO:1 in a 5' to 3' direction and those sequences which hybridize thereto under stringent hybridization conditions [for example, 0.1X SSC, 0.1% SDS at 65°C; see, T. Maniatis et al, Molecular Cloning (A Laboratory Manual), Cold Spring Harbor Laboratory (1982), pages 387 to 389] and encode a protein having tendon/ligament-like tissue inducing activity.

Similarly, DNA sequences which code for proteins coded for by the sequences of SEQ ID NO:1 or SEQ ID NO:25, or proteins which comprise the amino acid sequence of SEQ ID NO:2 or SEQ ID NO:26, but which differ in codon

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sequence due to the degeneracies of the genetic code or allelic variations (naturally-occurring base changes in the species population which may or may not result in an amino acid change) also encode the tendon/ligament-like tissue inducing proteins described herein. Variations in the DNA sequences of SEQ ID NO:1 or SEQ ID NO:25 which are caused by point mutations or by induced modifications (including insertion, deletion, and substitution) to enhance the activity, half-life or production of the polypeptides encoded are also encompassed in the invention.

Another aspect of the present invention provides a novel method for producing tendon/ligament-like tissue inducing proteins. The method of the present invention involves culturing a suitable cell line, which has been transformed with a DNA sequence encoding a protein of the invention, under the control of known regulatory sequences. The transformed host cells are cultured and the proteins recovered and purified from the culture medium. The purified proteins are substantially free from other proteins with which they are co-produced as well as from other contaminants.

Suitable cells or cell lines may be mammalian cells, such as Chinese hamster ovary cells (CHO). As described above, expression of protein in mammalian cells requires an appropriate propeptide to assure secretion of the protein. The selection of suitable mammalian host cells and methods for transformation, culture, amplification, screening, product production and purification are known in the art. See, e.g., Gething and Sambrook, Nature, 293:620-625 (1981), or alternatively, Kaufman et al, Mol. Cell. Biol., 5(7):1750-1759 (1985) or Howley et al, U.S. Patent 4,419,446. Another suitable mammalian cell line, which is described in the accompanying examples, is the monkey COS-1 cell line. The mammalian cell CV-1 may also be suitable.

Bacterial cells may also be suitable hosts. For example, the various strains of <u>E. coli</u> (e.g., HB101, MC1061) are well-known as host cells in the field of biotechnology. Various strains of <u>B. subtilis</u>, <u>Pseudomonas</u>, other bacilli and the like may also be employed in this method. For expression of the protein in bacterial cells, DNA encoding a propeptide is not necessary.

Bacterial expression of mammalian proteins, including members of the TGF- β family is known to produce the proteins in a non-glycosylated form, and in the form

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of insoluble pellets, known as inclusion bodies. Techniques have been described in the art for solubilizing these inclusion bodies, denaturing the protein using a chaotropic agent, and refolding the protein sufficiently correctly to allow for their production in a soluble form. For example, see EP 0433225, the disclosure of which is hereby incorporated by reference.

Alternatively, methods have been devised which circumvent inclusion body formation, such as expression of gene fusion proteins, wherein the desired protein is expressed as a fusion protein with a fusion partner. The fusion protein is later subjected to cleavage to produce the desired protein. One example of such a gene fusion expression system for <u>E. coli</u> is based on use of the <u>E. coli</u> thioredoxin gene as a fusion partner, LaVallie et al., <u>Bio/Technology</u>, 11:187-193 (1993), the disclosure of which is hereby incorporated by reference.

Many strains of yeast cells known to those skilled in the art may also be available as host cells for expression of the polypeptides of the present invention. Additionally, where desired, insect cells may be utilized as host cells in the method of the present invention. See, e.g. Miller et al, <u>Genetic Engineering</u>, <u>8</u>:277-298 (Plenum Press 1986) and references cited therein.

Another aspect of the present invention provides vectors for use in the method of expression of these tendon/ligament-like tissue inducing proteins. Preferably the vectors contain the full novel DNA sequences described above which encode the novel factors of the invention. Additionally, the vectors contain appropriate expression control sequences permitting expression of the protein sequences. Alternatively, vectors incorporating modified sequences as described above are also embodiments of the present invention. Additionally, the sequence of SEQ ID NO:1 or SEQ ID NO:3 or SEQ ID NO:25 could be manipulated to express a mature protein by deleting propeptide sequences and replacing them with sequences encoding the complete propeptides of BMP proteins or members of the TGF- β superfamily. Thus, the present invention includes chimeric DNA molecules encoding a propeptide from a member of the TGF- β superfamily linked in correct reading frame to a DNA sequence encoding a protein having the amino acid sequence of SEQ ID NO:2 or SEQ ID NO:4 or SEQ ID NO:26. The vectors may be employed in the method of transforming cell lines and contain selected regulatory sequences in operative

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association with the DNA coding sequences of the invention which are capable of directing the replication and expression thereof in selected host cells. Regulatory sequences for such vectors are known to those skilled in the art and may be selected depending upon the host cells. Such selection is routine and does not form part of the present invention.

A protein of the present invention, which induces tendon/ligament-like tissue or other tissue formation in circumstances where such tissue is not normally formed, has application in the healing of tendon or ligament tears, deformities and other tendon or ligament defects in humans and other animals. Such a preparation employing a tendon/ligament-like tissue inducing protein may have prophylactic use in preventing damage to tendon or ligament tissue, as well as use in the improved fixation of tendon or ligament to bone or other tissues, and in repairing defects to tendon or ligament tissue. De novo tendon/ligament-like tissue formation induced by a composition of the present invention contributes to the repair of congenital, trauma induced, or other tendon or ligament defects of other origin, and is also useful in cosmetic plastic surgery for attachment or repair of tendons or ligaments. The compositions of the invention may also be useful in the treatment of tendinitis, carpal tunnel syndrome and other tendon or ligament defects. The compositions of the present invention can also be used in other indications wherein it is desirable to heal or regenerate tendon and/or ligament tissue. Such indications include, without limitation, regeneration or repair of injuries to the periodontal ligament, such as occurs in tendonitis, and regeneration or repair of the tendon-to-bone attachment. The compositions of the present invention may provide an environment to attract tendon- or ligament-forming cells, stimulate growth of tendon- or ligament-forming cells or induce differentiation of progenitors of tendon- or ligament-forming cells.

The BMP-12 related proteins may be recovered from the culture medium and purified by isolating them from other proteinaceous materials from which they are co-produced and from other contaminants present. The proteins of the present invention are capable of inducing the formation of tendon/ligament-like tissue. These proteins may be further characterized by the ability to demonstrate tendon/ligament-like tissue formation activity in the rat ectopic implant assay

described below. It is contemplated that these proteins may have ability to induce the formation of other types of tissue, such as ligaments, as well.

The tendon/ligament-like tissue inducing proteins provided herein also include factors encoded by the sequences similar to those of SEQ ID NO:1 or SEQ ID NO:25, but into which modifications are naturally provided (e.g. allelic variations in the nucleotide sequence which may result in amino acid changes in the polypeptide) or deliberately engineered. For example, synthetic polypeptides may wholly or partially duplicate continuous sequences of the amino acid residues of SEQ ID NO:2. These sequences, by virtue of sharing primary, secondary, or tertiary structural and conformational characteristics with tendon/ligament-like tissue growth factor polypeptides of SEQ ID NO:2 may possess tendon/ligament-like or other tissue growth factor biological properties in common therewith. Thus, they may be employed as biologically active substitutes for naturally-occurring tendon/ligament-like tissue inducing polypeptides in therapeutic compositions and processes.

Other specific mutations of the sequences of tendon/ligament-like tissue inducing proteins described herein involve modifications of glycosylation sites. These modifications may involve O-linked or N-linked glycosylation sites. For instance, the absence of glycosylation or only partial glycosylation results from amino acid substitution or deletion at asparagine-linked glycosylation recognition sites. The asparagine-linked glycosylation recognition sites comprise tripeptide sequences which are specifically recognized by appropriate cellular glycosylation enzymes. These tripeptide sequences may be asparagine-X-threonine, asparagine-X-serine or asparagine-X-cysteine, where X is usually any amino acid except proline. A variety of amino acid substitutions or deletions at one or both of the first or third amino acid positions of a glycosylation recognition site (and/or amino acid deletion at the second position) results in non-glycosylation at the modified tripeptide sequence. Additionally, bacterial expression of protein will also result in production of a non-glycosylated protein, even if the glycosylation sites are left unmodified.

The compositions of the present invention comprise a purified BMP-12 related protein which may be produced by culturing a cell transformed with the DNA sequence of SEQ ID NO:1 or SEQ ID NO:25 and recovering and purifying protein having the amino acid sequence of SEQ ID NO:2 or SEQ ID NO:26 from the culture

medium. The purified expressed protein is substantially free from other proteinaceous materials with which it is co-produced, as well as from other contaminants. The recovered purified protein is contemplated to exhibit tendon/ligament-like tissue formation activity, and other tissue growth activity, such as ligament regeneration. The proteins of the invention may be further characterized by the ability to demonstrate tendon/ligament-like tissue formation activity in the rat assay described below.

The compositions for inducing tendon/ligament-like tissue formation of the present invention may comprise an effective amount of a tendon/ligament-like tissue inducing protein, wherein said protein comprises the amino acid sequence of SEQ ID NO:2, preferably amino acids #-25. #1 or #3 to #103 or #104 of SEQ ID NO:2; or amino acids #1 or #19 to #120 of SEQ ID NO:26; as well as mutants and/or variants of SEQ ID NO:2 or SEQ ID NO:26, which exhibit the ability to form tendon and/or ligament like tissue.

Compositions of the present invention may further comprise additional proteins, such as additional members of the TGF- β superfamily of proteins, such as activins. Another aspect of the invention provides pharmaceutical compositions containing a therapeutically effective amount of a tendon/ligament-inducing protein, such as BMP-12 or VL-1, in a pharmaceutically acceptable vehicle or carrier. These compositions may be used to induce the formation of tendon/ligament-like tissue or other tissue. It is contemplated that such compositions may also be used for tendon and ligament repair, wound healing and other tissue repair, such as skin repair. It is further contemplated that proteins of the invention may increase neuronal survival and therefore be useful in transplantation and treatment of conditions exhibiting a decrease in neuronal survival. Compositions of the invention may further include at least one other therapeutically useful agent, such as the BMP proteins BMP-1, BMP-2, BMP-3, BMP-4, BMP-5, BMP-6 and BMP-7, disclosed for instance in United States Patents 5,108,922; 5,013,649; 5,116,738; 5,106,748; 5,187,076; and 5,141,905; BMP-8, disclosed in PCT publication WO91/18098; BMP-9, disclosed in PCT publication WO93/00432; and BMP-10 or BMP-11, disclosed in co-pending patent applications, serial number 08/061,695 and 08/061,464, filed on May 12,

1993. The disclosure of the above documents are hereby incorporated by reference herein.

The compositions of the invention may comprise, in addition to a tendon/ligament-inducing protein such as BMF-12 or VL-1 (BMP-13), other therapeutically useful agents including MP52, epidermal growth factor (EGF), fibroblast growth factor (FGF), platelet derived growth factor (PDGF), transforming growth factors (TGF- α and TGF- β), and fibroblast growth factor-4 (FGF-4), parathyroid hormone (PTH), leukemia inhibitory factor (LIF/HILDA/DIA), insulinlike growth factors (IGF-I and IGF-II). Portions of these agents may also be used in compositions of the present invention. For example, a composition comprising both BMP-2 and BMP-12 implanted together gives rise to both bone and tendon/ligament-like tissue. Such a composition may be useful for treating defects of the embryonic joint where tendon, ligaments, and bone form simultaneously at contiguous anatomical locations, and may be useful for regenerating tissue at the site of tendon attachment to bone. It is contemplated that the compositions of the invention may also be used in wound healing, such as skin healing and related tissue repair. The types of wounds include, but are not limited to burns, incisions and ulcers. (See, e.g. PCT Publication WO84/01106 for discussion of wound healing and related tissue repair).

It is expected that the proteins of the invention may act in concert with or perhaps synergistically with other related proteins and growth factors. Further therapeutic methods and compositions of the invention therefore comprise a therapeutic amount of at least one protein of the invention with a therapeutic amount of at least one of the BMP proteins described above. Such compositions may comprise separate molecules of the BMP proteins or heteromolecules comprised of different BMP moieties. For example, a method and composition of the invention may comprise a disulfide linked dimer comprising a BMP-12 related protein subunit and a subunit from one of the "BMP" proteins described above. Thus, the present invention includes compositions comprising a purified BMP-12 related polypeptide which is a heterodimer wherein one subunit comprises the amino acid sequence from amino acid #1 to amino acid #104 of SEQ ID NO:2, and one subunit comprises an amino acid sequence for a bone morphogenetic protein selected from the group

consisting of BMP-1, BMP-2, BMP-3, BMP-4, BMP-5, BMP-6, BMP-7, BMP-8, BMP-9, BMP-10 and BMP-11. A further embodiment may comprise a heterodimer of disulfide bonded tendon/ligament-like tissue inducing moieties such as BMP-12, VL-1 (BMP-13) or MP52. For example the heterodimer may comprise one subunit comprising an amino acid sequence from #1 to # 104 of SEQ ID NO:2 and the other subunit may comprise an amino acid sequence from #1 to #120 of SEQ ID NO:4 or #1 to #120 of SEQ ID NO:26. Further, compositions of the present invention may be combined with other agents beneficial to the treatment of the defect, wound, or tissue in question.

The preparation and formulation of such physiologically acceptable protein compositions, having due regard to pH, isotonicity, stability and the like, is within the skill of the art. The therapeutic compositions are also presently valuable for veterinary applications due to the lack of species specificity in TGF- β proteins. Particularly domestic animals and thoroughbred horses in addition to humans are desired patients for such treatment with the compositions of the present invention.

The therapeutic method includes administering the composition topically, systemically, or locally as an implant or device. When administered, the therapeutic composition for use in this invention is, of course, in a pyrogen-free, physiologically acceptable form. Further, the composition may desirably be encapsulated or injected in a viscous form for delivery to the site of tissue damage. Topical administration may be suitable for wound healing and tissue repair. Therapeutically useful agents other than the proteins which may also optionally be included in the composition as described above, may alternatively or additionally, be administered simultaneously or sequentially with the composition in the methods of the invention.

The compositions may also include an appropriate matrix and/or sequestering agent as a carrier. For instance, the matrix may support the composition or provide a surface for tendon/ligament-like tissue formation and/or other tissue formation. The matrix may provide slow release of the protein and/or the appropriate environment for presentation thereof. The sequestering agent may be a substance which aids in ease of administration through injection or other means, or may slow the migration of protein from the site of application.

The choice of a carrier material is based on biocompatibility, biodegradability, mechanical properties, cosmetic appearance and interface properties. The particular application of the compositions will define the appropriate formulation. Potential matrices for the compositions may be biodegradable and chemically defined. Further matrices are comprised of pure proteins or extracellular matrix components. Other potential matrices are nonbiodegradable and chemically defined. Preferred matrices include collagen-based materials, such as Helistat sponge (Integra LifeSciences, Plainsboro, N.J.), or collagen in an injectable form, as well as sequestering agents, which may also be biodegradable, and which may include alkylcellulosic materials.

Another preferred class of carrier are porous particulate polymer matrices, including polymers of poly(lactic acid), poly(glycolic acid) and copolymers of lactic acid and glycolic acid. These matrices may also include a sequestering agent. Suitable polymer matrices are described, for example, in WO 93/00050, the disclosure of which is incorporated herein by reference.

A preferred family of sequestering agents is cellulosic materials such as alkylcelluloses (including hydroxyalkylcelluloses), including methylcellulose, ethylcellulose, hydroxyethylcellulose, hydroxypropylcellulose, hydroxypropylmethylcellulose, and carboxymethylcellulose, the most preferred being cationic salts of carboxymethylcellulose (CMC). Other preferred sequestering agents include hyaluronic acid, sodium alginate, poly(ethylene glycol), polyoxyethylene oxide, carboxyvinyl polymer and poly(vinyl alcohol). The amount of sequestering agent useful herein is 0.5-20 wt%, preferably 1-10 wt% based on total formulation weight, which represents the amount necessary to prevent desorbtion of the protein from the polymer matrix and to provide appropriate handling of the composition, yet not so much that the progenitor cells are prevented from infiltrating the matrix, thereby providing the protein the opportunity to assist the activity of the progenitor cells.

Additional optional components useful in the practice of the subject application include, e.g. cryogenic protectors such as mannitol, sucrose, lactose, glucose, or glycine (to protect the protein from degradation during lyophilization), antimicrobial preservatives such as methyl and propyl parabens and benzyl alcohol;

antioxidants such as EDTA, citrate and BHT (butylated hydroxytoluene); and surfactants such as poly(sorbates) and poly(oxyethylenes); etc.

As described above, the compositions of the invention may be employed in methods for treating a number of tendon defects, such as the regeneration of tendon/ligament-like tissue in areas of tendon or ligament damage, to assist in repair of tears of tendon tissue, ligaments, and various other types of tissue defects or wounds. These methods, according to the invention, entail administering to a patient needing such tendon/ligament-like tissue or other tissue repair, a composition comprising an effective amount of a tendon/ligament-like tissue inducing protein, such as described in SEQ ID NO:2, SEQ ID NO:4 and/or SEQ ID NO:26. These methods may also entail the administration of a tendon/ligament-like tissue inducing protein in conjunction with at least one of the BMP proteins described above.

In another embodiment, the methods may entail administration of a heterodimeric protein in which one of the monomers is a tendon/ligament-like tissue inducing polypeptide, such as BMP-12, VL-1 (BMP-13) or MP52, and the second monomer is a member of the TGF- β superfamily of growth factors. In addition, these methods may also include the administration of a tendon/ligament-like tissue inducing protein with other growth factors including EGF, FGF, TGF- α , TGF- β , and IGF.

Thus, a further aspect of the invention is a therapeutic method and composition for repairing tendon/ligament-like tissue, for repairing tendon or ligament as well as treating tendinitis and other conditions related to tendon or ligament defects. Such compositions comprise a therapeutically effective amount of one or more tendon/ligament-like tissue inducing proteins, such as BMP-12, a BMP-12 related protein, or MP52, in admixture with a pharmaceutically acceptable vehicle, carrier or matrix.

The dosage regimen will be determined by the attending physician considering various factors which modify the action of the composition, e.g., amount of tendon or ligament tissue desired to be formed, the site of tendon or ligament damage, the condition of the damaged tendon or ligament, the size of a wound, type of damaged tissue, the patient's age, sex, and diet, the severity of any infection, time of administration and other clinical factors. The dosage may vary with the type of

matrix used in the reconstitution and the types of additional proteins in the composition. The addition of other known growth factors, such as IGF-I (insulin like growth factor I), to the final composition, may also affect the dosage.

Progress can be monitored by periodic assessment of tendon/ligament-like tissue formation, or tendon or ligament growth and/or repair. The progress can be monitored by methods known in the art, for example, X-rays, arthroscopy, histomorphometric determinations and tetracycline labeling.

The following examples illustrate practice of the present invention in recovering and characterizing human tendon/ligament-like tissue inducing protein and employing them to recover the other tendon/ligament-like tissue inducing proteins, obtaining the human proteins, expressing the proteins via recombinant techniques, and demonstration of the ability of the compositions of the present invention to form tendon/ligament-like tissue in an in vivo model. Although the examples demonstrate the invention with respect to BMP-12, with minor modifications within the skill of the art, the same results are believed to be attainable with MP52 and VL-1.

EXAMPLE 1

Isolation of DNA

DNA sequences encoding BMP-12 and BMP-12 related proteins may be isolated by various techniques known to those skilled in the art. As described below, oligonucleotide primers may be designed on the basis of amino acid sequences present in other BMP proteins, Vg-1 related proteins and other proteins of the TGF- β superfamily. Regions containing amino acid sequences which are highly conserved within the BMP family of proteins and within other members of the TGF- β superfamily of proteins can be identified and consensus amino acid sequences of these highly conserved regions can be constructed based on the similarity of the corresponding regions of individual BMP/TGF- β /Vg-1 proteins. An example of such a consensus amino acid sequence is indicated below.

Consensus amino acid sequence (1):

Trp-Gln/Asn-Asp-Trp-Ile-Val/Ile-Ala (SEQ ID NO:16)

Where X/Y indicates that either amino acid residue may appear at that position.

The following oligonucleotide is designed on the basis of the above identified consensus amino acid sequence (1):

#1: CGGATCCTGGVANGAYTGGATHRTNGC (SEQ ID NO:17)

This oligonucleotide sequence is synthesized on an automated DNA synthesizer. The standard nucleotide symbols in the above identified oligonucleotide primer are as follows: A,adenosine; C,cytosine; G,guanine; T,thymine; N,adenosine or cytosine or guanine or thymine; R,adenosine or cytosine or thymine; H,adenosine or cytosine or thymine; V,adenosine or cytosine or guanine; D,adenosine or guanine or thymine.

The first seven nucleotides of oligonucleotide #1 (underlined) contain the recognition sequence for the restriction endonuclease BamHI in order to facilitate the manipulation of a specifically amplified DNA sequence encoding the BMP-12 protein and are thus not derived from the consensus amino acid sequence (1) presented above.

A second consensus amino acid sequence is derived from another highly conserved region of BMP/TGF- β /Vg-1 proteins as described below:

His-Ala-Ile-Val/Leu-Gln-Thr (SEQ ID NO:18)

The following oligonucleotide is designed on the basis of the above identified consensus amino acid sequence (2):

#2: TTTCTAGAARNGTYTGNACDATNGCRTG (SEQ ID NO:19)

This oligonucleotide sequence is synthesized on an automated DNA synthesizer. The same nucleotide symbols are used as described above.

The first seven nucleotides of oligonucleotide #1 (underlined) contain the recognition sequence for the restriction endonuclease XbaI in order to facilitate the manipulation of a specifically amplified DNA sequence encoding the BMP-12 protein and are thus not derived from the consensus amino acid sequence (2) presented above.

It is contemplated that the BMP-12 protein of the invention and other BMP/TGF-β/Vg-1 related proteins may contain amino acid sequences similar to the consensus amino acid sequences described above and that the location of those sequences within a BMP-12 protein or other novel related proteins would correspond to the relative locations in the proteins from which they were derived. It is further

contemplated that this positional information derived from the structure of other BMP/TGF- β /Vg-1 proteins and the oligonucleotide sequences #1 and #2 which have been derived from consensus amino acid sequences (1) and (2),respectively, could be utilized to specifically amplify DNA sequences encoding the corresponding amino acids of a BMP-12 protein or other BMP/TGF- β /Vg-1 related proteins.

Based on the knowledge of the gene structures of BMP/TGF- β /Vg-1 proteins it is further contemplated that human genomic DNA can be used as a template to perform specific amplification reactions which would result in the identification of BMP-12 BMP/TGF- β /Vg-1 (BMP-12 related protein) encoding sequences. Such specific amplification reactions of a human genomic DNA template could be initiated with the use of oligonucleotide primers #1 and #2 described earlier. Oligonucleotides #1 and #2 identified above are utilized as primers to allow the specific amplification of a specific nucleotide sequence from human genomic DNA. The amplification reaction is performed as follows:

Human genomic DNA (source: peripheral blood lymphocytes), provided by Ken Jacobs of Genetics Institute, is sheared by repeated passage through a 25 gauge needle, denatured at 100°C for 5 minutes and then chilled on ice before adding to a reaction mixture containing 200 μM each deoxynucleotide triphosphates (dATP, dGTP, dCTP and dTTP), 10 mM Tris-HCl pH 8.3, 50 mM KCl, 1.5 mM MgCl₂, 0.001% gelatin, 1.25 units Taq DNA polymerase, 100 pM oligonucleotide #1 and 100 pM oligonucleotide #2. This reaction mixture is incubated at 94°C for two minutes and then subjected to thermal cycling in the following manner: 1 minute at 94°C, 1 minute at 40°C, 1 minute at 72°C for three cycles; then 1 minute at 94°C, 1 minute at 72°C for thirty-seven cycles, followed by a 10 minute incubation at 72°C.

The DNA which is specifically amplified by this reaction is ethanol precipitated, digested with the restriction endonucleases BamHI and XbaI and subjected to agarose gel electrophoresis. A region of the gel, corresponding to the predicted size of the BMP-12 or other BMP/TGF-β/Vg-1 encoding DNA fragment, is excised and the specifically amplified DNA fragments contained therein are electroeluted and subcloned into the plasmid vector pGEM-3 between the XbaI and BamHI sites of the polylinker. DNA sequence analysis of one of the resulting BMP-

12 related subclones indicates the specifically amplified DNA sequence product contained therein encodes a portion of the BMP-12 protein of the invention.

The DNA sequence (SEQ ID NO:5) and derived amino acid sequence (SEQ ID NO:6) of this specifically amplified DNA fragment of BMP-12 are shown in the SEQUENCE Listings.

Nucleotides #1-#26 of SEQ ID NO:5 comprise a portion of oligonucleotide #1 and nucleotides #103 - #128 comprise a portion of the reverse compliment of oligonucleotide #2 utilized to perform the specific amplification reaction. Due to the function of oligonucleotides #1 and #2 in initiating the amplification reaction, they may not correspond exactly to the actual sequence encoding a BMP-12 protein and are therefore not translated in the corresponding amino acid derivation (SEQ ID NO:6).

DNA sequence analysis of another subclone indicates that the specifically amplified DNA product contained therein encodes a portion of another BMP/TGF- β /Vg-1 (BMP-12 related) protein of the invention named VL-1.

The DNA sequence (SEQ ID NO:7) and derived amino acid sequence (SEQ ID NO:8) of this specifically amplified DNA fragment are shown in the Sequence Listings.

Nucleotides #1 - #26 of SEQ ID NO:7 comprise a portion of oligonucleotide #1 and nucleotides #103 - #128 comprise a portion of the reverse compliment of oligonucleotide #2 utilized to perform the specific amplification reaction. Due to the function of oligonucleotides #1 and #2 in initiating the amplification reaction, they may not correspond exactly to the actual sequence encoding a VL-1 protein of the invention and are therefore not translated in the corresponding amino acid derivation (SEQ ID NO:8).

The following oligonucleotide probe is designed on the basis of the specifically amplified BMP-12 human DNA sequence set forth above (SEQ ID NO:5) and synthesized on an automated DNA synthesizer:

#3: CCACTGCGAGGCCTTTGCGACTTCCCTTTGCGTTCGCAC (SEQ ID NO:20)

This oligonucleotide probe is radioactively labeled with ^{32}P and employed to screen a human genomic library constructed in the vector λFIX (Stratagene catalog

#944201). 500,000 recombinants of the human genomic library are plated at a density of approximately 10,000 recombinants per plate on 50 plates. Duplicate nitrocellulose replicas of the recombinant bacteriophage plaques and hybridized to oligonucleotide probe #3 in standard hybridization buffer (SHB = 5X SSC, 0.1% SDS, 5X Denhardt's, 100 μ g/ml salmon sperm DNA) at 65°C overnight. The following day the radioactively labelled oligonucleotide containing hybridization solution is removed an the filters are washed with 0.2X SSC, 0.1% SDS at 65°C. A single positively hybridizing recombinant is identified and plaque purified. This plaque purified recombinant bacteriophage clone which hybridizes to the BMP-12 oligonucleotide probe #3 is designated \(\lambda\)HuG-48. A bacteriophage plate stock is made and bacteriophage DNA is isolated from the λHuG-48 human genomic clone. The bacteriophage λ HuG-48 has been deposited with the American Type Culture Collection, 12301 Parklawn Drive, Rockville, MD "ATCC" under the accession #75625 on December 7, 1993. This deposit meets the requirements of the Budapest Treaty of the International Recognition of the Deposit of Microorganisms for the Purpose of Patent Procedure and Regulations thereunder. The oligonucleotide hybridizing region of this recombinant, \(\lambda\)HuG-48, is localized to a 3.2 kb BamHI fragment. This fragment is subcloned into a plasmid vector (pGEM-3) and DNA sequence analysis is performed. This plasmid subclone is designated PCR1-1#2 and has been deposited with the American Type Culture Collection, 12301 Parklawn Drive, Rockville, MD "ATCC" under the accession #69517 on December 7, 1993. This deposit meets the requirements of the Budapest Treaty of the International Recognition of the Deposit of Microorganisms for the Purpose of Patent Procedure and Regulations thereunder. The partial DNA sequence (SEQ ID NO:1) and derived amino acid sequence (SEQ ID NO:2) of the 3.2 kb DNA insert of the plasmid subclone PCR1-1#2, derived from clone \(\lambda \text{HuG-48}, \) are shown in the Sequence Listings.

It should be noted that nucleotides #639 - #714 of SEQ ID NO:1 correspond to nucleotides #27 - #102 of the specifically amplified BMP-12 encoding DNA fragment set forth in SEQ ID NO:5 thus confirming that the human genomic bacteriophage clone λHuG-48 and derivative subclone PCR1-1#2 encode at least a portion of the BMP-12 protein of the invention. The nucleotide sequence of a

portion of the 3.2 kb BamHI insert of the plasmid PCR1-1#2 contains an open reading frame of at least 882 base pairs, as defined by nucleotides #1-#882 of SEQ ID NO:1. This open reading frame encodes at least 294 amino acids of the human BMP-12 protein of the invention. The encoded 294 amino acid human BMP-12 protein includes the full mature human BMP-12 protein (amino acids #1-#104 of SEQ ID NO:2), as well as the C-terminal portion of the propeptide region of the primary translation product (amino acid #-190 to #-1 of SEQ ID NO:2).

Additional DNA sequence of the 3.2 kb BamHI insert of the plasmid PCR1-1#2 set forth in SEQ ID NO:33 demonstrates the presence of an 1164 bp open reading frame, as defined by nucleotides #138 through #1301 of SEQ ID NO:33. [NOTE that all the sequence disclosed in SEQ ID NO:1 is contained within SEQ ID NO:33]. As this sequence is derived from a genomic clone it is difficult to determine the boundary between the 5' extent of coding sequence and the 3' limit of intervening sequence (intron/non-coding sequence).

Based on the knowledge of other BMP proteins and other proteins within the TGF- β family, it is predicted that the precursor polypeptide would be cleaved at the multibasic sequence Arg-Arg-Gly-Arg in agreement with a proposed consensus proteolytic processing sequence of Arg-X-X-Arg. Cleavage of the BMP-12 precursor polypeptide is expected to generate a 104 amino acid mature peptide beginning with the amino acid Ser at position #1 of SEQ ID NO:2. The processing of BMP-12 into the mature form is expected to involve dimerization and removal of the N-terminal region in a manner analogous to the processing of the related protein TGF- β [Gentry et al., Molec & Cell. Biol., 8:4162 (1988); Derynck et al. Nature, 316:701 (1985)].

It is contemplated therefore that the mature active species of BMP-12 comprises a homodimer of two polypeptide subunits, each subunit comprising amino acids #1 to #104 of SEQ ID NO:2 with a predicted molecular weight of approximately 12,000 daltons. Further active species are contemplated comprising at least amino acids #3 to #103 of SEQ ID NO:2, thereby including the first and last conserved cysteine residue. As with other members of the TGF- β /BMP family of proteins, the carboxy-terminal portion of the BMP-12 protein exhibits greater sequence conservation than the more amino-terminal portion. The percent amino

acid identity of the human BMP-12 protein in the cysteine-rich C-terminal domain (amino acids #3 - #104) to the corresponding region of human BMP proteins and other proteins within the TGF- β family is as follows: BMP-2, 55%; BMP-3, 43%; BMP-4, 53%; BMP-5, 49%; BMP-6, 49%; BMP-7, 50%; BMP-8, 57%; BMP-9, 48%; BMP-10, 57%; activin WC (BMP-11), 38%; Vg1, 46%; GDF-1, 47%; TGF- β 1, 36%; TGF- β 2, 36%; TGF- β 3, 39%; inhibin β (B), 36%; inhibin β (A), 41%.

The human BMP-12 DNA sequence (SEQ ID NO:1), or a portion thereof, can be used as a probe to identify a human cell line or tissue which synthesizes BMP-12 mRNA. Briefly described, RNA is extracted from a selected cell or tissue source and either electrophoresed on a formaldehyde agarose gel and transferred to nitrocellulose, or reacted with formaldehyde and spotted on nitrocellulose directly. The nitrocellulose is then hybridized to a probe derived from the coding sequence of human BMP-12.

Alternatively, the human BMP-12 sequence is used to design oligonucleotide primers which will specifically amplify a portion of the BMP-12 encoding sequence located in the region between the primers utilized to perform the specific amplification reaction. It is contemplated that these human BMP-12 derived primers would allow one to specifically amplify corresponding BMP-12 encoding sequences from mRNA, cDNA or genomic DNA templates. Once a positive source has been identified by one of the above described methods, mRNA is selected by oligo (dT) cellulose chromatography and cDNA is synthesized and cloned in λ gt10 or other λ bacteriophage vectors known to those skilled in the art, for example, λ ZAP by established techniques (Toole et al., supra). It is also possible to perform the oligonucleotide primer directed amplification reaction, described above, directly on a pre-established human cDNA or genomic library which has been cloned into a λ bacteriophage vector. In such cases, a library which yields a specifically amplified DNA product encoding a portion of the human BMP-12 protein could be screened directly, utilizing the fragment of amplified BMP-12 encoding DNA as a probe.

Oligonucleotide primers designed on the basis of the DNA sequence of the human BMP-12 genomic clone λHuG -48 are predicted to allow the specific amplification of human BMP-12 encoding DNA sequences from pre-established human cDNA libraries which are commercially available (ie. Stratagene, La Jolla,

CA or Clontech Laboratories, Inc., Palo Alto, CA). The following oligonucleotide primer is designed on the basis of nucleotides #571 to #590 of the DNA sequence set forth in SEQ ID NO:1 and synthesized on an automated DNA synthesizer:

#4: TGCGGATCCAGCCGCTGCAGCCGCAAGCC (SEQ ID NO:21)

The first nine nucleotides of primer #4 (underlined) comprise the recognition sequence for the restriction endonuclease BamHI which can be used to facilitate the manipulation of a specifically amplified DNA sequence encoding the human BMP-12 protein of the invention and are thus not derived from the DNA sequence presented in SEQ ID NO:1.

The following oligonucleotide primer is designed on the basis of nucleotides #866 - #885 of the DNA sequence set forth in SEQ ID NO:1 and synthesized on an automated DNA synthesizer:

#5 GACTCTAGACTACCTGCAGCCGCAGGCCT (SEQ ID NO:22)

The first nine nucleotides of primer #5 (underlined) comprise the recognition sequence for the restriction endonuclease XbaI which can be used to facilitate the manipulation of a specifically amplified DNA sequence encoding the human BMP-12 protein of the invention and are thus not derived from the DNA sequence presented in SEQ ID NO:1.

The standard nucleotide symbols in the above identified primers are as follows: A, adenine; C, cytosine; G, guanine; T, thymine.

Primers #4 and #5 identified above are utilized as primers to allow the amplification of a specific BMP-12 encoding nucleotide sequence from preestablished cDNA libraries which may include the following: human fetal brain cDNA/λZAPII (Stratagene catalog #936206), human liver/λUNI-ZAP XR (Stratagene Catalog #937200), human lung/λUNI-ZAP XR (Stratagene catalog #937205).

Approximately 1 x 10^8 pfu (plaque forming units) of λ bacteriophage libraries containing human cDNA inserts such as those detailed above are denatured at 95°C for five minutes prior to addition to a reaction mixture containing 200 μ M each deoxynucleotide triphosphates (dATP, dGTP, dCTP and dTTP) 10 mM Tris-HCl pH 8.3, 50 mM KCl, 1.5 mM MgCl₂, 0.001% gelatin, 1.25 units Taq DNA polymerase, 100 pM oligonucleotide primer #4 and 100 pM oligonucleotide primer

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#5. The reaction mixture is then subjected to thermal cycling in the following manner: 1 minute at 94°C, 1 minute at 50°C, 1 minute at 72°C for thirty-nine cycles followed by 10 minutes at 72°C.

The DNA which is specifically amplified by this reaction would be expected to generate a BMP-12 encoding product of approximately 333 base pairs, the internal 315 bp of which correspond to nucleotides #571 to #885 of SEQ ID NO:1 and also including 9 bp at each end of the BMP-12 specific fragment which correspond to the restriction sites defined by nucleotides #1 - #9 of primers #4 and #5. The resulting 333 bp DNA product is digested with the restriction endonucleases BamHI and XbaI, phenol extracted, chloroform extracted and ethanol precipitated.

Alternatively, to ethanol precipitation, buffer exchange and removal of small fragments of DNA resulting from the BamHI/Xbal restriction digest is accomplished by dilution of the digested DNA product in 10 mM Tris-HCl pH 8.0, 1 mM EDTA followed by centrifugation through a Centricon™ 30 microconcentrator (W.R. Grace & Co., Beverly, MA; Product #4209). The resulting BamHI/Xbal digested amplified DNA product is subcloned into a plasmid vector (ie. pBluescript, pGEM-3 etc.) between the BamHI and Xbal sites of the polylinker region. DNA sequence analysis of the resulting subclones would be required to confirm the integrity of the BMP-12 encoding insert. Once a positive cDNA source has been identified in this manner, the corresponding cDNA library from which a 333 bp BMP-12 specific sequence was amplified could be screened directly with the 333 bp insert or other BMP-12 specific probes in order to identify and isolate cDNA clones encoding the full-length BMP-12 protein of the invention.

Additional methods known to those skilled in the art may be used to isolate other full-length cDNAs encoding human BMP-12 related proteins, or full length cDNA clones encoding BMP-12 related proteins of the invention from species other than humans, particularly other mammalian species.

The following examples demonstrate the use of the human BMP-12 sequence to isolate homologues from BMP-12 related proteins in a murine genomic DNA library.

The DNA sequence which encodes the human BMP-12 protein of the invention is predicted to be significantly homologous to BMP-12 and BMP-12 related

sequences from species other than humans that it could be utilized to specifically amplify DNA sequences from those other species which would encode the corresponding BMP-12 related proteins. Specifically, the following oligonucleotides are designed on the basis of the human BMP-12 sequence (SEQ ID NO:1) and are synthesized on an automated DNA synthesizer:

#6: GCGGATCCAAGGAGCTCGGCTGGGACGA (SEQ ID NO:23)

#7: GGAATTCCCCACCACCATGTCCTCGTAT (SEQ ID NO:24)

The first eight nucleotides of oligonucleotide primers #6 and #7 (underlined) comprise the recognition sequence for the restriction endonucleases BamHI and EcoRI, respectively. These sequences are utilized to facilitate the manipulation of a specifically amplified DNA sequence encoding a BMP-12 or BMP-12 related protein from a species other than human and are thus not derived from the DNA sequence presented in SEQ ID NO:1. Oligonucleotide primer #6 is designed on the basis of nucleotides #607-#626 of SEQ ID NO:1. Oligonucleotide primer #7 is designed on the basis of the reverse compliment of nucleotides #846-#865 of the DNA sequence set forth in SEQ ID NO:1.

Oligonucleotide primers #6 and #7 identified above are utilized as primers to allow the amplification of specific BMP-12 related sequences from genomic DNA derived from species other than humans. The amplification reaction is performed as follows:

Murine genomic DNA (source: strain Balb c) is sheared by repeated passage through a 25 gauge needle, denatured at 100° C for five minutes and then chilled on ice before adding to a reaction mixture containing 200 μM each deoxynucleotide triphosphates (dATP, DGTP, dCTP and dTTP) 10 mM Tris-HCl pH 8.3, 50 mM KCl, 1.5 mM MgCl₂, 0.001% gelatin, 1.25 units Taq DNA polymerase, 100 pM oligonucleotide primer #6 and 100 pM oligonucleotide primer #7. The reaction mixture is then subjected to thermal cycling in the following manner: 1 minute at 95°C, 1 minute at 55°C, 1 minute at 72°C for forty cycles followed by 10 minutes at 72°C.

The DNA which is specifically amplified by this reaction is ethanol precipitated, digested with the restriction endonucleases BamHI and EcoRI and subjected to agarose gel electrophoresis. A region of the gel, corresponding to the

predicted size of the murine BMP-12 or BMP-12 related encoding DNA fragment, is excised and the specifically amplified DNA fragments contained therein are extracted (by electroelution or by other methods known to those skilled in the art) and subcloned in to a plasmid vector, such as pGEM-3 or pBluescript between the BamHI and EcoRI sites of the polylinker. DNA sequence analysis of one of the resulting subclones named mV1, indicates that the specifically amplified DNA sequence contained therein encodes a portion of a protein which appears to be the murine homolog to either the BMP-12 or VL-1 sequence of the invention. The DNA sequence (SEQ ID NO:10) and derived amino acid sequence (SEQ ID NO:11) of this specifically amplified murine DNA fragment are shown in the sequence listings.

Nucleotides #1-#26 of SEQ ID NO:10 comprise a portion of oligonucleotide #6 and nucleotides #246-#272 comprise a portion of the reverse compliment of oligonucleotide #7 utilized to perform the specific amplification reaction. Nucleotide #27 of SEQ ID NO:10 appears to be the last nucleotide of a codon triplet, and nucleotides #244-#245 of SEQ ID NO:10 appear to be the first two nucleotides of a codon triplet. Therefore, nucleotides #28 to #243 of SEQ ID NO:10 correspond to a partial coding sequence of mV1. Due to the function of oligonucleotides #6 and #7 in initiating the amplification reaction, they may not correspond exactly to the actual sequence encoding the murine homolog to the human BMP-12 or VL-1 protein of the invention and are therefore not translated in the corresponding amino acid sequence derivation (SEQ ID NO:11).

Oligonucleotide probes designed on the basis of the specifically amplified murine BMP-12 or VL-1 DNA sequence set forth in SEQ ID NO:10 can be utilized by those skilled in the art to identify full-length murine BMP-12 or VL-1 encoding clones (either cDNA or genomic).

DNA sequence analysis of another of the resulting subclones named mV2, indicates that the specifically amplified DNA sequence contained therein encodes a portion of a murine BMP-12 related sequence of the invention. The DNA sequence (SEQ ID NO:12) and derived amino acid sequence (SEQ ID NO:13) of this specifically amplified murine DNA fragment are shown in the sequence listings.

Nucleotides #1-#26 of SEQ ID NO:12 comprise a portion of oligonucleotide #6 and nucleotides #246-#272 comprise a portion of the reverse compliment of

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oligonucleotide #7 utilized to perform the specific amplification reaction. Nucleotide #27 of SEQ ID NO:12 appears to be the last nucleotide of a codon triplet, and nucleotides #244-#245 of SEQ ID NO:12 appear to be the first two nucleotides of a codon triplet. Therefore, nucleotides #28 to #243 of SEQ ID NO:12 correspond to a partial coding sequence of mV2. Due to the function of oligonucleotides #6 and #7 in initiating the amplification reaction, they may not correspond exactly to the actual sequence encoding the murine BMP-12 related protein of the invention and are therefore not translated in the corresponding amino acid sequence derivation (SEQ ID NO:13).

Oligonucleotide probes designed on the basis of the specifically amplified murine BMP-12 related DNA sequence set forth in SEQ ID NO:12 can be utilized by those skilled in the art to identify full-length murine BMP-12 related encoding clones (either cDNA or genomic).

DNA sequence analysis of another of the resulting subclones named mV9, indicates that the specifically amplified DNA sequence contained therein encodes a portion of a murine BMP-12 related sequence of the invention. This sequence appears to be the murine homolog to the human MP52 DNA sequence described at SEQ ID NO:3. The DNA sequence (SEQ ID NO:14) and derived amino acid sequence (SEQ ID NO:15) of this specifically amplified murine DNA fragment are shown in the sequence listings.

Nucleotides #1-#26 of SEQ ID NO:14 comprise a portion of oligonucleotide #6 and nucleotides #246-#272 comprise a portion of the reverse compliment of oligonucleotide #7 utilized to perform the specific amplification reaction. Nucleotide #27 of SEQ ID NO:14 appears to be the last nucleotide of a codon triplet, and nucleotides #244-#245 of SEQ ID NO:14 appear to be the first two nucleotides of a codon triplet. Therefore, nucleotides #28 to #243 of SEQ ID NO:14 correspond to a partial coding sequence of mV9. Due to the function of oligonucleotides #6 and #7 in initiating the amplification reaction, they may not correspond exactly to the actual sequence encoding the murine BMP-12 related protein of the invention and are therefore not translated in the corresponding amino acid sequence derivation (SEQ ID NO:15).

Oligonucleotide probes designed on the basis of the specifically amplified murine BMP-12 related DNA sequence set forth in SEQ ID NO:14 can be utilized by those skilled in the art to identify full-length murine BMP-12 related encoding clones (either cDNA or genomic).

Alternatively, oligonucleotide primers #6 and #7 identified above are utilized as primers to allow the specific amplification of a 275 base pair DNA probe, the internal 259 bp of which correspond to nucleotides #607 to #865 of SEQ ID NO:1, from the BMP-12 encoding plasmid subclone PCR1-1#2. This 275bp DNA probe was radioactively labelled with ³²P and employed to screen a murine genomic library constructed in the vector λ FIX II (Stratagene catalog #946306). recombinants of the murine genomic library are plated at a density of approximately 20,000 recombinants per plate on 50 plates. Duplicate nitrocellulose replicas of the recombinant bacteriophage plaques are hybridized, under reduced stringency conditions, to the specifically amplified 333 bp probe in standard hybridization buffer (SHB = 5X SSC, 0.1% SDS, 5X Denhardt's, 100 μ g/ml salmon sperm DNA) at 60°C overnight. The following day the radioactively labelled oligonucleotide containing hybridization solution is removed an the filters are washed, under reduced stringency conditions, with 2X SSC, 0.1% SDS at 60°C. Multiple positively hybridizing recombinants are identified and plaque purified. Fragments of the positively hybridizing murine genomic recombinant clones are subcloned into standard plasmid vectors (i.e. pGEM-3) and subjected to DNA sequence analysis.

DNA sequence analysis of one of these subclones named MVR3 indicates that it encodes a portion of the mouse gene corresponding to the PCR product mV1 (murine homolog of the human BMP-12 sequence set forth in SEQ ID NO:1) described above. The partial DNA sequence of this subclone and corresponding amino acid translation are set forth in SEQ ID NO: 29 and SEQ ID NO:30 respectively.

DNA sequence analysis of another one of these subclones named MVR32 indicates that it encodes a portion of the mouse gene corresponding to the PCR product mV2 (murine homolog of the human VL-1 sequence set forth in SEQ ID NO:7) described above. The partial DNA sequence of this subclone and

corresponding amino acid translation are set forth in SEQ ID NO: 31 and SEQ ID NO:32 respectively.

DNA sequence analysis of another of these subclones named MVR23 indicates that it encodes a portion of the mouse gene corresponding to the PCR product mV9 (murine homolog of the MP-52 sequence set forth in SEQ ID NO:3) described above.

In a similar manner to that which is described above for identifying and isolating human genomic clones encoding the BMP-12 protein of the invention, oligonucleotide probe(s) corresponding to the VL-1 encoding sequence set forth in SEQ ID NO:7 can be designed and utilized to identify human genomic or cDNA sequences encoding the VL-1 (BMP-13) protein. These oligonucleotides would be designed to regions specific for VL-1 encoding sequences and would therefore be likely to be derived from regions of the lowest degree of nucleotide sequence identity between the specifically amplified VL-1 encoding sequence (SEQ ID NO:7) and the specifically amplified BMP-12 encoding sequence (SEQ ID NO:5).

Alternatively, oligonucleotide primers #4 and #5 identified above are utilized as primers to allow the specific amplification of a 333 base pair DNA probe, the internal 315 bp of which correspond to nucleotides #571 to #885 of SEQ ID NO:1, from the BMP-12 encoding plasmid subclone PCR1-1#2. This 333 bp DNA probe was radioactively labelled with 32 P and employed to screen a human genomic library constructed in the vector λ DASH II (Stratagene catalog #945203). 1 million recombinants of the human genomic library are plated at a density of approximately 20,000 recombinants per plate on 50 plates. Duplicate nitrocellulose replicas of the recombinant bacteriophage plaques are hybridized, under reduced stringency conditions, to the specifically amplified 333 bp probe in standard hybridization buffer (SHB = 5X SSC, 0.1% SDS, 5X Denhardt's, 100 μ g/ml salmon sperm DNA) at 60°C overnight. The following day the radioactively labelled oligonucleotide containing hybridization solution is removed an the filters are washed, under reduced stringency conditions, with 2X SSC, 0.1% SDS at 60°C. Multiple (approximately 15) positively hybridizing recombinants are identified and plaque purified.

In order to distinguish positively hybridizing recombinants encoding the VL-1 protein of the invention from BMP-12 and other BMP-12-related encoding 34

recombinants which would be predicted to hybridize positively to the 333 bp DNA probe generated from the BMP-12 encoding plasmid PCR1-1#2 utilized in this screening procedure, the following oligonucleotide probe, based on the VL-1 sequence set forth in SEQ ID NO:7, is designed and synthesized on an automated DNA synthesizer:

#8: TGTATGCGACTTCCCGC [SEQUENCE ID NO: 35]

An oligonucleotide corresponding to nucleotides #60 to #76 of SEO ID NO:7 which contains 5 nucleotide differences to the corresponding region of the BMP-12 encoding sequence set forth in SEQ ID NO:1 (nucleotides #672 to #689) One of the recombinant bacteriophage clones which hybridizes to the VL-1 oligonucleotide probe #8 is designated \(\lambda\)JLDc31. This recombinant bacteriophage clone is plaque purified, a bacteriophage plate stock is made and bacteriophage DNA is isolated from the \(\lambda JLDc31\) human genomic clone. The bacteriophage \(\lambda JLDc31\) has been deposited with the American Type Culture Collection, 12301 Parklawn Drive, Rockville, MD "ATCC" under the accession #75922 on October 20, 1994. This deposit meets the requirements of the Budapest Treaty of the International Recognition of the Deposit of Microorganisms for the Purpose of Patent Procedure and Regulations thereunder. The oligonucleotide hybridizing region of this recombinant, \(\lambda \) LDc31, is localized to a 2.5 kb Eco RI fragment. This fragment is subcloned into a plasmid vector (pGEM-3) and DNA sequence analysis is performed. This plasmid subclone is designated pGEMJLDc31/2.5 and has been deposited with the American Type Culture Collection, 12301 Parklawn Drive, Rockville, MD "ATCC" under the accession # 69710 on October 20, 1994. This deposit meets the requirements of the Budapest Treaty of the International Recognition of the Deposit of Microorganisms for the Purpose of Patent Procedure and Regulations thereunder.

The partial DNA sequence (SEQ ID NO:25) and derived amino acid sequence (SEQ ID NO:26) of a portion of the 2.5 kb DNA insert of the plasmid subclone pGEMJLDc31/2.5, derived from clone λJLDc31, are shown in the Sequence Listings

The DNA sequence of a portion of the 2.5 kb EcoRI insert of the plasmid pGEMJLDc31/2.5 is set forth in SEQ ID NO:25.

contains an 912 bp open reading frame, as defined by nucleotides #52 through #963 of SEQ ID NO:25. As this sequence is derived from a genomic clone it is difficult to determine the boundary between the 5' extent of coding sequence and the 3' limit of intervening sequence (intron/non-coding sequence). The entire open reading frame (nucleotides #52 through #963 of SEQ ID NO:25) encodes a portion of the VL-1 protein of the invention of up to 304 amino acids.

Based on the knowledge of other BMP proteins and other proteins within the TGF- β family, it is predicted that the precursor polypeptide would be cleaved at the multibasic sequence Arg-Arg-Arg in agreement with a proposed consensus proteolytic processing sequence of Arg-X-X-Arg. Cleavage of the VL-1 precursor polypeptide is expected to generate a 120 amino acid mature peptide beginning with the amino acid Thr at position #1 of SEQ ID NO:26. The processing of VL-1 into the mature form is expected to involve dimerization and removal of the N-terminal region in a manner analogous to the processing of the related protein TGF- β [Gentry et al., Molec & Cell. Biol., 8:4162 (1988); Derynck et al. Nature, 316:701 (1985)].

It is contemplated therefore that the mature active species of VL-1 comprises a homodimer of two polypeptide subunits, each subunit comprising amino acids #1 to #120 of SEQ ID NO:26 with a predicted molecular weight of approximately 12,000 daltons. Further active species are contemplated comprising at least amino acids #19 to # 119 or #120 of SEQ ID NO:26, thereby including the first and last conserved cysteine residue.

Using such a method, a clone encoding the mature human VL-1 (BMP-13) was obtained. The nucleotide sequence and corresponding amino acid sequence encoded by this clone are listed in the Sequence Listings at SEQ ID NO: 25 and 26, respectively.

EXAMPLE 2

Expression of BMP-12

In order to produce human BMP-12 proteins, the DNA encoding it is transferred into an appropriate expression vector and introduced into mammalian cells or other preferred eukaryotic or prokaryotic hosts by conventional genetic engineering techniques.

In order to produce the human BMP-12 protein in bacterial cells, the following procedure is employed.

Expression of BMP-12 in E. coli

An expression plasmid pALV1-781, for production of BMP-12 in E. coli was constructed which contains the following principal features. Nucleotides 1-2060 contain DNA sequences originating from the plasmid pUC-18 [Norrander et al., Gene 26:101-106 (1983)] including sequences containing the gene for β -lactamase which confers resistance to the antibiotic ampicillin in host E. coli strains, and a colE1-derived origin of replication. Nucleotides 2061-2221 contain DNA sequences for the major leftward promotor (pL) of bacteriophage λ [Sanger et al., J. Mol. Biol. 162:729-773 (1982)], including three operator sequences 0_L1 , 0_L2 and 0_L3 . The operators are the binding sites for \(\lambda \text{I repressor protein, intracellular levels of which } \) control the amount of transcription initiation from pL. Nucleotides 2222-2723 contain a strong ribosome binding sequence included on a sequence derived from nucleotides 35566 to 35472 and 38137 to 38361 from bacteriophage lambda as described in Sanger et al., J. Mol. Biol. 162:729-773 (1982). Nucleotides 2724-3041 contain a DNA sequence encoding mature BMP-12 protein with all 3' untranslated sequence removed. The BMP-12 DNA sequences introduced into the pALV1-781 expression vector were modified at the 5'end to raise the A+T content without altering the coding capacity. These changes were made to increase the efficiency of translation initiated on the BMP-12 mRNA in E. coli. Nucleotides 3042-3058 provide a "Linker" DNA sequence containing restriction endonuclease sites. Nucleotides 3059-3127 provide a transcription termination sequence based on that of the E. coli asp A gene [Takagi et al., Nucl. Acids Res. 13:2063-2074 (1985)]. Nucleotides 3128-3532 are DNA sequences derived from pUC-18.

Plasmid pALV1-781 was transformed into the *E. coli* host strain GI724 (F, lacI^q, lacp^{L8}, ampC::λcI⁺) by the procedure of Dagert and Ehrlich, Gene 6:23 (1979). GI724 (ATCC accession No. 55151) contains a copy of the wild-type λcI repressor gene stably integrated into the chromosome at the ampC locus, where it has been placed under the transcriptional control of *Salmonella typhimurium* trp promotor/operator sequences. In GI724, λCI protein is made only during growth in tryptophan-free media, such as minimal media or a minimal medium supplemented

with casamino acids such as IMC, described above. Addition of tryptophan to a culture of GI724 will repress the <u>trp</u> promoter and turn off synthesis of λcI , gradually causing the induction of transcription from pL promoters if they are present in the cell.

Transformants were selected on 1.5% w/v agar plates containing IMC medium, which is composed of M9 medium [Miller, "Experiments in Molecular Genetics," Cold Spring Harbor Laboratory, New York (1972)] containing 1 mM MgSO₄ and supplemented with 0.5% w/v glucose, 0.2% w/v casamino acids and 100 μ g/ml ampicillin. GI724 transformed with pALV1-781 was grown at 37°C to an A₅₅₀ of 0.5 in IMC medium containing 100 μ g/ml ampicillin. Tryptophan was then added to a final concentration of 100 μ g/ml and the culture incubated for a further 4 hours. During this time BMP-12 protein accumulates within the "inclusion body" fraction.

Preparation of Protein Monomer

18 g of frozen cells were weighed out and resuspended in 60ml of 100 mM Tris, 10 mM EDTA, 1 mM phenylmethylsulfonyl fluoride [PMSF], pH 8.3. Cells were lysed by 3 passes through a MicrofluidizerTM [model #MCF 100 T]. The inclusion body pellet was obtained by centrifugation at 15,000g at 4°C for 20 minutes. The supernatant was decanted, and the pellet was washed with 100 ml of 100 mM Tris, 1.0 M NaCl, 10 mM EDTA, 1 mM PMSF, pH 8.3. The suspension was centrifuged again at 15,000g at 4°C for 10 minutes, and the supernatant decanted. The pellet was then washed with 100 ml of 100 mM Tris, 10 mM EDTA, 1% Triton X-100, 1 mM PMSF, pH 8.3. The suspension was centrifuged again at 15,000g at 4°C for 10 minutes, and the supernatant decanted. The pellet was resuspended with 50 ml of 20 mM Tris, 1 mM EDTA, 1 mM PMSF, pH 8.3, containing 1% DTT in a glass tissue homogenizer. Monomeric BMP-12 was then solubilized by acidification to pH 2.5 with glacial acetic acid. The soluble fraction was isolated by centrifugation at 15,000g for 20 minutes at 4°C.

The supernatant from this centrifugation was collected and chromatographed over a Sephacryl S- 100^{TM} size exclusion column (83 cm x 2.6 cm; \approx 440 ml bed) in 20 ml increments. The Sephacryl S- 100^{TM} column was run with a mobile phase of 1% acetic acid at a flow rate of 1.4 ml/min. Fractions corresponding to BMP-12

monomer were detected by absorbance at 280 nm, and using a computer calculated extinction coefficient of 18200M⁻¹cm⁻¹ and molecular weight (11667 daltons). This size exclusion column pooled material was used as starting material for refolding reactions.

As an alternative to the above, 1.0 g of cells stored at -80°C are measured. Solution (3.4 ml 100 mM TRIS, 10 mM EDTA, pH 8.5) is added. The solution is vortexed until cells are well suspended. 40 µl 100 mM PMSF in isopropanol is added. The cells are lysed at 1000 psi in a French pressure cell. The inclusion bodies are centrifuged at 4°C for 20 minutes in an Eppendorf microfuge to form pellets. The supernatants are decanted. To one pellet (out of 4 total) 1.0 ml degassed 8.0 M guanidine hydrochloride, 0.5 M TRIS, 5 mM EDTA, pH 8.5, containing 250 mM DTT is added. The pellet is dissolved and argon is blown over the liquid for 30 seconds. Next the solution is incubated at 37°C for one hour. Insoluble material is pelleted for 2-3 minutes in an Eppendorf microfuge at 23°C. 0.5-1.0 ml of supernatant is injected onto a Supelco 2 cm guard cartridge (LC-304), and eluted with an acetonitrile gradient in 0.1% TFA from 1-70% over 35 minutes. BMP-12 elutes between 29 and 31 minutes. Fractions are pooled and the protein concentration determined by adsorbance at 280 nanometers versus 0.1% TFA, using the theoretical extinction coefficient based upon the amino acid content.

As a second alternate method to the above, frozen cell pellets obtained from the *E. coli* transformants as described above are thawed in 30 ml of TE8.3(100:10) buffer (100 mM Tris-HCl pH 8.3, 10 mM Na₂EDTA, 1 mM PMSF). Cells are lysed by three passes through a MicrofluidizerTM [model #MCF 100 T]. The initial inclusion body material pellet is dissolved in 8 M guanidine-HCl, TE8.5(100:10) buffer (100 mM Tris-HCl pH 8.5, 10 mM Na₂EDTA which contained 100 mM DTT, and incubated at 37°C for 1 hour. This material is centrifuged at 12,000 x g for 15 minutes at room temperature.

Refolding of BMP-12 protein using CHAPS system

A sufficient volume of the BMP-12 pool is lyophilized to give 10 μ g of protein. 5 μ l of glass distilled water is added to redissolve the residue, then 100 μ l of refold mix (50 mM Tris, 1.0 M NaCl, 2% 3-(3-chlolamido-3?)

propyl)dimethylammonio-1-propane-sulfate (CHAPS), 5 mM EDTA, 2 mM glutathione (reduced) 1 mM glutathione (oxidized); at pH of approximately 8.5). The solution is gently mixed and stored at 23°C for 1-4 days. Dimer formation is assessed by running an aliquot on a Novex 16% tricine gel at 125 volts for 2.5 hours, followed by Coomassie Blue staining and destaining.

BMP-12 dimer was purified using a C4 analytical RP-HPLC (reversed phase-high performance liquid chromatography) column (Vydac 214TP54) which was equilibrated to 1% B buffer (diluted into A buffer) and was run over 35 minutes, during which the protein elutes, using the following gradient (A buffer = 0.1% trifluoroacetic acid, B buffer = 95% acetonitrile, 0.1% trifluoroacetic acid [TFA]), with a flow rate of 1 ml/min:

1-5 minutes 20% B buffer

5-10 minutes 20-30% B buffer

10-30 minutes 30-50% B buffer

30-35 minutes 50-100% B buffer

Protein was monitored by absorbance at 280nm. Peak BMP-12 fractions (eluting between 29 and 31 minutes) were pooled. Purity was assessed by SDS-PAGE. The concentration was determined by absorbance at 280nm, and using the computer calculated extinction coefficient and molecular weight as indicated above.

Expression of BMP-12 in mammalian cells:

Another contemplated preferred expression system for biologically active recombinant human BMP-12 is stably transformed mammalian cells.

One skilled in the art can construct mammalian expression vectors by employing the sequence of SEQ ID NO:1, or other DNA sequences encoding BMP-12 proteins or other modified sequences and known vectors, such as pCD [Okayama et al., Mol. Cell Biol., 2:161-170 (1982)], pJL3, pJL4 [Gough et al., EMBO J., 4:645-653 (1985)] and pMT2 CXM.

The mammalian expression vector pMT2 CXM is a derivative of p91023(b) (Wong et al., Science 228:810-815, 1985) differing from the latter in that it contains the ampicillin resistance gene in place of the tetracycline resistance gene and further contains a XhoI site for insertion of cDNA clones. The functional elements of pMT2 CXM have been described (Kaufman, R.J., 1985, Proc. Natl. Acad. Sci.

USA <u>82</u>:689-693) and include the adenovirus VA genes, the SV40 origin of replication including the 72 bp enhancer, the adenovirus major late promoter including a 5' splice site and the majority of the adenovirus tripartite leader sequence present on adenovirus late mRNAs, a 3' splice acceptor site, a DHFR insert, the SV40 early polyadenylation site (SV40), and pBR322 sequences needed for propagation in <u>E. coli</u>.

Plasmid pMT2 CXM is obtained by EcoRI digestion of pMT2-VWF, which has been deposited with the American Type Culture Collection (ATCC), Rockville, MD (USA) under accession number ATCC 67122. EcoRI digestion excises the cDNA insert present in pMT2-VWF, yielding pMT2 in linear form which can be ligated and used to transform <u>E</u>. <u>coli</u> HB 101 or DH-5 to ampicillin resistance. Plasmid pMT2 DNA can be prepared by conventional methods. pMT2 CXM is then constructed using loopout/in mutagenesis [Morinaga, et al., <u>Biotechnology 84</u>: 636 (1984). This removes bases 1075 to 1145 relative to the Hind III site near the SV40 origin of replication and enhancer sequences of pMT2. In addition it inserts a sequence containing the recognition site for the restriction endonuclease Xho I. A derivative of pMT2CXM, termed pMT23, contains recognition sites for the restriction endonucleases PsI, Eco RI, SalI and XhoI. Plasmid pMT2 CXM and pMT23 DNA may be prepared by conventional methods.

pEMC2β1 derived from pMT21 may also be suitable in practice of the invention. pMT21 is derived from pMT2 which is derived from pMT2-VWF. As described above EcoRI digestion excises the cDNA insert present in pMT-VWF, yielding pMT2 in linear form which can be ligated and used to transform E. Coli HB 101 or DH-5 to ampicillin resistance. Plasmid pMT2 DNA can be prepared by conventional methods.

pMT21 is derived from pMT2 through the following two modifications. First, 76 bp of the 5' untranslated region of the DHFR cDNA including a stretch of 19 G residues from G/C tailing for cDNA cloning is deleted. In this process, a XhoI site is inserted to obtain the following sequence immediately upstream from DHFR. Second, a unique ClaI site is introduced by digestion with EcoRV and XbaI, treatment with Klenow fragment of DNA polymerase I, and ligation to a ClaI linker (CATCGATG). This deletes a 250 bp segment from the adenovirus associated RNA

(VAI) region but does not interfere with VAI RNA gene expression or function. pMT21 is digested with EcoRI and XhoI, and used to derive the vector pEMC2B1.

A portion of the EMCV leader is obtained from pMT2-ECAT1 [S.K. Jung, et al, J. Virol 63:1651-1660 (1989)] by digestion with Eco RI and PstI, resulting in a 2752 bp fragment. This fragment is digested with TaqI yielding an Eco RI-TaqI fragment of 508 bp which is purified by electrophoresis on low melting agarose gel. A 68 bp adapter and its complementary strand are synthesized with a 5' TaqI protruding end and a 3' XhoI protruding end which has a sequence which matches the EMC virus leader sequence from nucleotide 763 to 827. It also changes the ATG at position 10 within the EMC virus leader to an ATT and is followed by a XhoI site. A three way ligation of the pMT21 Eco RI-XhoI fragment, the EMC virus EcoRI-TaqI fragment, and the 68 bp oligonucleotide adapter TaqI-XhoI adapter resulting in the vector pEMC2 β 1.

This vector contains the SV40 origin of replication and enhancer, the adenovirus major late promoter, a cDNA copy of the majority of the adenovirus tripartite leader sequence, a small hybrid intervening sequence, an SV40 polyadenylation signal and the adenovirus VA I gene, DHFR and β -lactamase markers and an EMC sequence, in appropriate relationships to direct the high level expression of the desired cDNA in mammalian cells.

The construction of vectors may involve modification of the BMP-12 DNA sequences. For instance, BMP-12 cDNA can be modified by removing the non-coding nucleotides on the 5' and 3' ends of the coding region. The deleted non-coding nucleotides may or may not be replaced by other sequences known to be beneficial for expression. These vectors are transformed into appropriate host cells for expression of BMP-12 proteins. Additionally, the sequence of SEQ ID NO:1 or other sequences encoding BMP-12 proteins can be manipulated to express BMP-12 protein by isolating the mature coding sequence of nucleotides 571 to 882 of SEQ ID NO:1 and adding at the 5' end sequences encoding the complete propeptides of other BMP proteins.

For example, one skilled in the art can make a fusion protein in which the propertide of BMP-2 is linked in operable fashion to the mature BMP-12 peptide by preparing a DNA vector in which the DNA sequence encoding the BMP-2

propertide is linked in proper reading frame to the DNA sequence encoding the mature BMP-12 peptide. The DNA sequence of such a fusion protein is shown in SEQUENCE ID NO:27.

One skilled in the art can manipulate the sequences of SEQ ID NO:1 by eliminating or replacing the mammalian regulatory sequences flanking the coding sequence with bacterial sequences to create bacterial vectors for intracellular or extracellular expression by bacterial cells, as described above. As another example, the coding sequences could be further manipulated (e.g. ligated to other known linkers or modified by deleting non-coding sequences therefrom or altering nucleotides therein by other known techniques). The modified BMP-12 coding sequence could then be inserted into a known bacterial vector using procedures such as described in T. Taniguchi et al., Proc. Natl Acad. Sci. USA, 77:5230-5233 (1980). This exemplary bacterial vector could then be transformed into bacterial host cells and a BMP-12 protein expressed thereby. For a strategy for producing extracellular expression of BMP-12 proteins in bacterial cells, see, e.g. European patent application EPA 177,343.

Similar manipulations can be performed for the construction of an insect vector [See, e.g. procedures described in published European patent application 155,476] for expression in insect cells. A yeast vector could also be constructed employing yeast regulatory sequences for intracellular or extracellular expression of the factors of the present invention by yeast cells. [See, e.g., procedures described in published PCT application WO86/00639 and European patent application EPA 123,289].

A method for producing high levels of a BMP-12 protein of the invention in mammalian cells may involve the construction of cells containing multiple copies of the heterologous BMP-12 gene. The heterologous gene is linked to an amplifiable marker, e.g. the dihydrofolate reductase (DHFR) gene for which cells containing increased gene copies can be selected for propagation in increasing concentrations of methotrexate (MTX) according to the procedures of Kaufman and Sharp, J. Mol. Biol., 159:601-629 (1982). This approach can be employed with a number of different cell types.

For example, a plasmid containing a DNA sequence for a BMP-12 of the invention in operative association with other plasmid sequences enabling expression thereof and the DHFR expression plasmid pAdA26SV(A)3 [Kaufman and Sharp, Mol. Cell. Biol., 2:1304 (1982)] can be co-introduced into DHFR-deficient CHO cells, DUKX-BII, by various methods including calcium phosphate coprecipitation and transfection, electroporation or protoplast fusion. DHFR expressing transformants are selected for growth in alpha media with dialyzed fetal calf serum, and subsequently selected for amplification by growth in increasing concentrations of MTX (e.g. sequential steps in 0.02, 0.2, 1.0 and 5uM MTX) as described in Kaufman et al., Mol Cell Biol., 5:1750 (1983). Transformants are cloned, and biologically active BMP-12 expression is monitored by the Rosen-modified Sampath-Reddi rat assay described below in Example 5. BMP-12 expression should increase with increasing levels of MTX resistance. BMP-12 polypeptides are characterized using standard techniques known in the art such as pulse labeling with [35S] methionine or cysteine and polyacrylamide gel electrophoresis. Similar procedures can be followed to produce other related BMP-12 proteins.

EXAMPLE 3

Preparation of BMP-2 propeptide/BMP-12 mature peptide fusion

In order to construct a vector encoding the BMP-2 propeptide/BMP-12 mature peptide fusion, the following cloning procedure was used to fuse the two sequences together.

First, a DNA restriction enzyme fragment comprising the propeptide of human BMP-2 protein, comprising nucleotides 1 through 843 of SEQ ID NO:27 is cut from pBMP2 \(^\text{EMC}\). pBMP2 \(^\text{EMC}\) is a plasmid derived from lambda U20S-39 (ATCC #40345) comprising the entire coding sequence for human BMP-2 protein with the non-translated 5' and 3' sequences of BMP-2 deleted from the vector. The 5' restriction enzyme used was Bgl II and it cuts pBMP2 \(^\text{EMC}\) in the vector at nucleotide 979. The 3' restriction enzyme used was Mae II and it cuts pBMP2 \(^\text{EMC}\) in the BMP-2 propeptide at nucleotide 1925, just short of the carboxy terminus. The resulting 954 base pair product was then gel isolated and gene cleaned. Second, a DNA restriction enzyme fragment comprising the 5' portion of the human BMP-12 mature peptide DNA sequence, is cut from pPCR1-1#2 V1-1

(ATCC #69517). The 5' restriction enzyme used was Eae I and it cuts pPCR1-1#2 V1-1 just 3' of N-terminus of the human BMP-12 mature peptide sequence. The resulting 259 base pair product was gel isolated and gene cleaned. Third, two DNA oligos were designed and synthesized, so that when annealed would form a tiny DNA fragment comprising fusion sequence of the extreme 3' end of the human BMP-2 propertide and the 5' end of BMP-12 mature peptide. The DNA fragment has a 5' Mae II complimentary sticky end which anneals to the 3' restriction enzyme fragment comprising the human BMP-2 propeptide. The annealed oligo DNA fragment has a 3' Eae I complimentary sticky end which anneals to the 5' of the restriction enzyme fragment comprising the mature peptide of human BMP-12. The coding strand oligo is named B2/12 and is 13 base pairs long. Next, a DNA fragment encoding the 123 base pairs at the 3' end of the BMP-12 mature peptide fragment was obtained as follows. First, a DNA fragment comprising the propeptide of human BMP-2 protein, comprising nucleotides 1 through 846 is PCR amplified from pBMP2 & EMC. The 5' primer (oligo 655a) anneals just 5' of the polylinker. The 3' primer (BMPpro3) anneals to the BMP-2 propeptide 3' end and introduces a Bgl II restriction enzyme site by silent sequence mutations. The resulting PCR product was cut with Sal I, which cleaves in the polylinker, and Bgl II. The 850 base pair restriction enzyme fragment (ending in amino acid sequence REKR) was gel isolated and gene cleaned. The BMP-12 mature peptide was PCR amplified using a 5' primer (oligo 5-1) encoding the Bgl II restriction enzyme site by silent sequence mutations, and annealing to the 5' end of a possible mature cleavage product, beginning with amino acid sequence SRCS. The 3' primer (V1-1 3) anneals to the BMP-12 mature peptide 3' end and introduces a Xba I restriction enzyme site after the stop codon. The resulting PCR product was cut with Bgl II and Xba I. The 321 base pair restriction enzyme fragment was gel isolated and gene cleaned.

The two restriction fragments were three-way ligated into a previously SalI and XbaI cut vector. The resultant construct was sequenced to check for PCR induced errors and a silent C to T mutation was observed at base pair 185 in the propeptide. This plasmid was designated pREKRSRC. Then pREKRSRC was cut with BglII and NgoMI, and the vector fragment encompassing the last 123 base pairs of the BMP12 mature sequence was thereby isolated. The three restriction fragments

and the annealed oligolinker were four-way ligated to yield pREKR-TAL with the BMP-2 propeptide with the mature cleavage site at the 3' end fused to the (TAL) 5' end of the BMP-12 mature peptide. The coding sequence of the resulting ligated vector is shown in SEQ ID NO:27.

EXAMPLE 4

Biological Activity of Expressed BMP-12

To measure the biological activity of the expressed BMP-12 proteins obtained in Example 2 above, the proteins are recovered from the cell culture and purified by isolating the BMP-12 proteins from other proteinaceous materials with which they are co-produced as well as from other contaminants. The purified protein may be assayed in accordance with the rat assay described below in Example 5.

Purification is carried out using standard techniques known to those skilled in the art.

Protein analysis is conducted using standard techniques such as SDS-PAGE acrylamide [Laemmli, Nature 227:680 (1970)] stained with Coomassie Blue or silver [Oakley, et al. Anal. Biochem. 105:361 (1980)] and by immunoblot [Towbin, et al. Proc. Natl. Acad. Sci. USA 76:4350 (1979)]

Example 5

ROSEN MODIFIED SAMPATH-REDDI ASSAY

A modified version of the rat ectopic implant assay described in Sampath and Reddi, Proc. Natl. Acad. Sci. USA, 80:6591-6595 (1983) is used to evaluate the activity of the BMP-12 proteins. This modified assay is herein called the Rosen-modified Sampath-Reddi assay. The assay has been widely used to evaluate the bone and cartilage-inducing activity of BMPs. The ethanol precipitation step of the Sampath-Reddi procedure is replaced by dialyzing (if the composition is a solution) or diafiltering (if the composition is a suspension) the fraction to be assayed against water. The solution or suspension is then equilibrated to 0.1% TFA. The resulting solution is added to 20 mg of rat matrix. A mock rat matrix sample not treated with the protein serves as a control. This material is frozen and lyophilized and the resulting powder enclosed in #5 gelatin capsules. The capsules are implanted subcutaneously in the abdominal thoracic area of 21-49 day old male Long Evans rats. The implants are removed after 10 days. A section of each implant is fixed

and processed for histological analysis. 1 μ m glycolmethacrylate sections are stained with Von Kossa and acid fuschin to score the amount of induced tendon/ligament-like tissue formation present in each implant.

BMP-12 was implanted in the rats in doses of 1, 5, 25 and 50 μ g per implant for 10 days. BMP-2 at a dose of 5 μ g was included as a positive control. For all doses of BMP-12 tested, no bone or cartilage formation was observed in the implants after ten days. Instead, the implants were filled with tissue resembling embryonic tendon, which is easily recognized by the presence of dense bundles of fibroblasts oriented in the same plane and packed tightly together. [Tendon/ligament-like tissue is described, for example, in Ham and Cormack, Histology (JB Lippincott Co. (1979), pp. 367-369, the disclosure of which is hereby incorporated by reference]. These findings were reproduced in a second set of assays in which tendon/ligament-like tissues was present in all BMP-12 containing implants. In contrast, the BMP-2 implants, as expected, showed cartilage and bone formation, but contained no tendon/ligament-like tissue.

The BMP-12 proteins and related proteins of this invention may be assessed for activity on this assay.

Example 6

Using methods in accordance with the above examples, with minor modifications within the skill of the art, human MP52 protein and the murine homologue of BMP-13 protein were expressed and assayed for tendon/ligament-like tissue inducing activity. All proteins showed comparable results, similar to those described above for human BMP-12.

The foregoing descriptions detail presently preferred embodiments of the present invention. Numerous modifications and variations in practice thereof are expected to occur to those skilled in the art upon consideration of these descriptions. Those modifications and variations are believed to be encompassed within the claims appended hereto. The disclosure of all references discussed herein are hereby incorporated by reference.

SEOUENCE LISTING

- (1) GENERAL INFORMATION:
 - (i) APPLICANT: GENETICS INSTITUTE, INC.
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 - (ii) TITLE OF INVENTION: TENDON-INDUCING COMPOSITIONS
 - (iii) NUMBER OF SEQUENCES: 35
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 - (v) COMPUTER READABLE FORM:
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 - (B) COMPUTER: IBM PC compatible
 - (C) OPERATING SYSTEM: PC-DOS/MS-DOS
 - (D) SOFTWARE: PatentIn Release #1.0, Version #1.25
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 - (C) CLASSIFICATION:
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 - (B) FILING DATE: 07-DEC-1993
 - (C) APPLICATION NUMBER: US 08/217,780
 - (D) FILING DATE: 25-MAR-1994
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- (2) INFORMATION FOR SEQ ID NO:1:
 - (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 926 base pairs
 - (B) TYPE: nucleic acid
 - (C) STRANDEDNESS: single
 - (D) TOPOLOGY: linear
 - (ii) MOLECULE TYPE: DNA (genomic)
 - (vi) ORIGINAL SOURCE:
 - (A) ORGANISM: Homo sapiens
 - (vii) IMMEDIATE SOURCE:
 - (B) CLONE: v1-1
 - (ix) FEATURE:
 - (A) NAME/KEY: mat_peptide
 - (B) LOCATION: 571..882
 - (ix) FEATURE:
 - (A) NAME/KEY: CDS

(B) LOCATION: 1..882

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:1:

	(36-2	, 02	2021				J	JEQ .	10 14	J. 1.						
GCG Ala -19	Arg	AAT Asn	ACG Thr	ACT Thr	CAC His	Tyr	AGG Arg	GCG Ala	AAT Asn	TGG Trp -18	Val	CGG Arg	GGC Gly	CCA Pro	GGC Gly -175	48
AGC Ser	TGG Trp	ACT Thr	TCT Ser	CCG Pro	Pro	TTG Leu	CTG Leu	CTG Leu	CTG Leu -16	Ser	ACG Thr	TGC Cys	CCG Pro	GGC Gly -16	Ala	96
GCC Ala	CGA Arg	GCG Ala	CCA Pro -15	CGC Arg	CTG Leu	CTG Leu	TAC Tyr	TCG Ser -150	Arg	GCA Ala	GCT Ala	GAG Glu	CCC Pro -14	Leu	GTC Val	144
GGT Gly	CAG Gln	CGC Arg -140	Trp	GAG Glu	GCG Ala	TTC Phe	GAC Asp -13	Val	GCG Ala	GAC Asp	GCC Ala	ATG Met -13	Arg	CGC Arg	CAC His	192
CGT Arg	CGT Arg -12	Glu	CCG Pro	CGC Arg	CCC Pro	CCC Pro -120	Arg	GCG Ala	TTC Phe	TGC Cys	CTC Leu -11	Leu	CTG Leu	CGC Arg	GCA Ala	240
GTG Val -11	Ala	GGC Gly	CCG Pro	GTG Val	CCG Pro -105	Ser	CCG Pro	TTG Leu	GCA Ala	CTG Leu -10	Arg	CGA Arg	CTG Leu	GGC Gly	TTC Phe -95	288
GGC Gly	TGG Trp	CCG Pro	GGC Gly	GGA Gly -90	GGG Gly	GGC Gly	TCT Ser	GCG Ala	GCA Ala -85	GAG Glu	GAG Glu	CGC Arg	GCG Ala	GTG Val -80	CTA Leu	336
GTC Val	GTC Val	TCC Ser	TCC Ser -75	CGC Arg	ACG Thr	CAG Gln	AGG Arg	AAA Lys -70	GAG Glu	AGC Ser	TTA Leu	TTC Phe	CGG Arg -65	GAG Glu	ATC Ile	384
CGC Arg	GCC Ala	CAG Gln -60	GCC Ala	CGC Arg	GCG Ala	CTC Leu	GGG Gly -55	GCC Ala	GCT Ala	CTG Leu	GCC Ala	TCA Ser -50	GAG Glu	CCG Pro	CTG Leu	432
CCC Pro	GAC Asp -45	CCA Pro	GGA Gly	ACC Thr	GGC	ACC Thr -40	GCG Ala	TCG Ser	CCA Pro	AGG Arg	GCA Ala -35	GTC Val	ATT Ile	GGC Gly	GGC	480
CGC Arg -30	AGA Arg	CGG Arg	AGG Arg	AGG Arg	ACG Thr -25	GCG Ala	TTG Leu	GCC Ala	GGG Gly	ACG Thr -20	CGG Arg	ACA Thr	GCG Ala	CAG Gln	GGC Gly -15	528
AGC Ser	GGC Gly	GGG Gly	GGC Gly	GCG Ala -10	GGC Gly	CGG Arg	GGC Gly	CAC His	GGG Gly -5	CGC Arg	AGG Arg	GGC Gly	CGG Arg	AGC Ser 1	CGC Arg	576
TGC Cys	AGC Ser	CGC Arg 5	AAG Lys	CCG Pro	TTG Leu	CAC His	GTG Val 10	GAC Asp	TTC Phe	AAG Lys	GAG Glu	CTC Leu 15	GGC Gly	TGG Trp	GAC Asp	624
GAC Asp	TGG Trp 20	ATC Ile	ATC Ile	GCG Ala	CCG Pro	CTG Leu 25	GAC Asp	TAC Tyr	GAG Glu	GCG Ala	TAC Tyr 30	CAC His	TGC Cys	GAG Glu	GGC Gly	672
CTT Leu 35	TGC Cys	GAC Asp	TTC Phe	CCT Pro	TTG Leu 40	CGT Arg	TCG Ser	CAC His	CTC Leu	GAG Glu 45	CCC Pro	ACC Thr	AAC Asn	CAT His	GCC Ala 50	720

ATC Ile	ATT Ile	CAG Gln	ACG Thr	CTG Leu 55	CTC Leu	AAC Asn	TCC Ser	ATG Met	GCA Ala 60	CCA Pro	GAC Asp	GCG Ala	GCG Ala	CCG Pro 65	GCC Ala	768
TCC Ser	TGC Cys	TGT Cys	GTG Val 70	CCA Pro	GCG Ala	CGC Arg	CTC Leu	AGC Ser 75	CCC Pro	ATC Ile	AGC Ser	ATC Ile	CTC Leu 80	TAC Tyr	ATC Ile	816
			AAC Asn													864
GAG GCC TGC GGC TGC AGG TAGCGCGCGG GCCGGGGAGG GGGCAGCCAC Glu Ala Cys Gly Cys Arg 100														912		
GCGGCCGAGG ATCC													926			
(2)	INFO	RMAT	MOI	FOR	SEQ	ID 1	10:2	•								
	((i) S	(B)	LEN TYI	NGTH:	RACTE 294 amino 3Y:]	am:	ino a id		5						
	i)	Li) N	OLEC	CULE	TYPI	E: pı	rote:	Ln								
	()	ci) S	EQUE	ENCE	DES	CRIPT	CION	: SE	O ID	NO:2	2:					
Ala -190	_	Asn	Thr	Thr	His -1	_	Arg	Ala	Asn	_	Val L80	Arg	Gly	Pro	Gly -175	
Ser	Trp	Thr	Ser	Pro -170		Leu	Leu	Leu	Leu -1		Thr	Cys	Pro	_	Ala 160	
Ala	Arg	Ala	Pro -155		Leu	Leu	Tyr	Ser		Ala	Ala	Glu		Leu 145	Val	
Gly	Gln	Arg -140	Trp	Glu	Ala	Phe	Asp -1:		Ala	Asp	Ala		Arg 130	Arg	His	
Arg	Arg -125		Pro	Arg	Pro	Pro	_	Ala	Phe	Сув		Leu L15	Leu	Arg	Ala	
Val -110		Gly	Pro	Val	Pro		Pro	Leu	Ala		Arg 100	Arg	Leu	Gly	Phe -95	
Gly	Trp	Pro	Gly	Gly -90	Gly	Gly	Ser	Ala	Ala -85	Glu	Glu	Arg	Ala	Val -80	Leu	
Val	Val	Ser	Ser -75	Arg	Thr	Gln	Arg	Lys -70	Glu	Ser	Leu	Phe	Arg -65	Glu	Ile	
Arg	Ala	Gln -60	Ala	Arg	Ala	Leu	Gly -55	Ala	Ala	Leu	Ala	Ser -50	Glu	Pro	Leu	
Pro	Asp -45	Pro	Gly	Thr	·Gly	Thr	Ala	Ser	Pro	Arg	Ala -35	Val	Ile	Gly	Gly	
Arg -30	Arg	Arg	Arg	Arg	Thr -25	Ala	Leu	Ala	Gly	Thr	Arg	Thr	Ala	Gln	Gly -15	
Ser	Gly	Gly	Gly	Ala	Gly	Arg	Gly	His	Gly		Arg	Gly	Arg	Ser	Arg	

Cys Ser Arg Lys Pro Leu His Val Asp Phe Lys Glu Leu Gly Trp Asp Asp Trp Ile Ile Ala Pro Leu Asp Tyr Glu Ala Tyr His Cys Glu Gly Leu Cys Asp Phe Pro Leu Arg Ser His Leu Glu Pro Thr Asn His Ala 40 Ile Ile Gln Thr Leu Leu Asn Ser Met Ala Pro Asp Ala Ala Pro Ala Ser Cys Cys Val Pro Ala Arg Leu Ser Pro Ile Ser Ile Leu Tyr Ile Asp Ala Ala Asn Asn Val Val Tyr Lys Gln Tyr Glu Asp Met Val Val Glu Ala Cys Gly Cys Arg 100

- (2) INFORMATION FOR SEQ ID NO:3:
 - (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 1207 base pairs
 - (B) TYPE: nucleic acid
 - (C) STRANDEDNESS: single

 - (D) TOPOLOGY: linear
 - (ii) MOLECULE TYPE: DNA (genomic)
 - (vi) ORIGINAL SOURCE:
 - (A) ORGANISM: Homo sapiens
 - (vii) IMMEDIATE SOURCE:
 - (B) CLONE: MP52
 - (ix) FEATURE:
 - (A) NAME/KEY: CDS
 - (B) LOCATION: 845..1204
 - (xi) SEQUENCE DESCRIPTION: SEQ ID NO:3:

ACCGGGCGC CCTGAACCCA AGCCAGGACA CCCTCCCCAA ACAAGGCAGG CTACAGCCCG 60 120 CCCCAGCTCC TTCCTGCTGA AGAAGGCCAG GGAGCCCGGG CCCCCACGAG AGCCCAAGGA 180 GCCGTTTCGC CCACCCCCA TCACACCCCA CGAGTACATG CTCTCGCTGT ACAGGACGCT 240 GTCCGATGCT GACAGAAAGG GAGGCAACAG CAGCGTGAAG TTGGAGGCTG GCCTGGCCAA 300 CACCATCACC AGCTTTATTG ACAAAGGGCA AGATGACCGA GGTCCCGTGG TCAGGAAGCA 360 GAGGTACGTG TTTGACATTA GTGCCCTGGA GAAGGATGGG CTGCTGGGGG CCGAGCTCCG 420 480 TGCCCAGCTG AAGCTGTCCA GCTGCCCCAG CGGCCGCAG CCGGCCTCCT TGCTGGATGT 540 GCGCTCCGTG CCAGGCCTGG ACGGATCTGG CTGGGAGGTG TTCGACATCT GGAAGCTCTT 600 CCGAAACTIT AAGAACTCGG CCCAGCTGTG CCTGGAGCTG GAGGCCTGGG AACGGGGCAG 660 GGCCGTGGAC CTCCGTGGCC TGGGCTTCGA CCGCGCCGCC CGGCAGGTCC ACGAGAAGGC 720

CCT	STTC	CTG (TGT:	rtgg	CC G	CACC	AAGA	A ACC	EGGA (CCTG	TTC	LATT	ATG 2	AGAT.	raaggc	780
CCG	CTCT	GC (CAGG	ACGAT	A A	BACC	STGT	A TG	AGTA	CCTG	TTC	AGCCI	AGC (GGCG/	AAAACG	840
GCGC					Th						g Pro				C CTT Leu 15	889
				AGT Ser 20												937
				TGG Trp												985
				TGC Cys												1033
				ATC Ile												1081
				TGC Cys												1129
CTC Leu	TTC Phe	ATT Ile	GAC Asp	TCT Ser 100	GCC Ala	AAC Asn	AAC Asn	GTG Val	GTG Val 105	TAT Tyr	AAG Lys	CAG Gln	TAT Tyr	GAG Glu 110	GAC Asp	1177
				TCG Ser					TAG							1207

(2) INFORMATION FOR SEQ ID NO:4:

- (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 120 amino acids
 - (B) TYPE: amino acid(D) TOPOLOGY: linear
- (ii) MOLECULE TYPE: protein
- (xi) SEQUENCE DESCRIPTION: SEQ ID NO:4:

Ala Pro Leu Ala Thr Arg Gln Gly Lys Arg Pro Ser Lys Asn Leu Lys

1 10 15

Ala Arg Cys Ser Arg Lys Ala Leu His Val Asn Phe Lys Asp Met Gly

Trp Asp Asp Trp Ile Ile Ala Pro Leu Glu Tyr Glu Ala Phe His Cys

Glu Gly Leu Cys Glu Phe Pro Leu Arg Ser His Leu Glu Pro Thr Asn

His Ala Val Ile Gln Thr Leu Met Asn Ser Met Asp Pro Glu Ser Thr

Pro Pro Thr Cys Cys Val Pro Thr Arg Leu Ser Pro Ile Ser Ile Leu 85 90 95

Phe Ile Asp Ser Ala Asn Asn Val Val Tyr Lys Gln Tyr Glu Asp Met 105 Val Val Glu Ser Cys Gly Cys Arg (2) INFORMATION FOR SEQ ID NO:5: (i) SEQUENCE CHARACTERISTICS: (A) LENGTH: 128 base pairs (B) TYPE: nucleic acid (C) STRANDEDNESS: single (D) TOPOLOGY: linear (ii) MOLECULE TYPE: DNA (genomic) (vi) ORIGINAL SOURCE: (A) ORGANISM: Homo Sapiens (vii) IMMEDIATE SOURCE: (B) CLONE: V1-1 fragment (ix) FEATURE: (A) NAME/KEY: CDS (B) LOCATION: 28..102 (xi) SEQUENCE DESCRIPTION: SEQ ID NO:5: GGATCCTGGA AGGATTGGAT CATTGCG CCG CTG GAC TAC GAG GCG TAC CAC 51 Pro Leu Asp Tyr Glu Ala Tyr His TGC GAG GGC CTT TGC GAC TTC CCT TTG CGT TCG CAC CTC GAG CCC ACC 99 Cys Glu Gly Leu Cys Asp Phe Pro Leu Arg Ser His Leu Glu Pro Thr AAC CACGCTATAG TCCAAACCTT TCTAGA 128 Asn 25 (2) INFORMATION FOR SEQ ID NO:6: (i) SEQUENCE CHARACTERISTICS: (A) LENGTH: 25 amino acids(B) TYPE: amino acid(D) TOPOLOGY: linear (ii) MOLECULE TYPE: protein (xi) SEQUENCE DESCRIPTION: SEQ ID NO:6: Pro Leu Asp Tyr Glu Ala Tyr His Cys Glu Gly Leu Cys Asp Phe Pro Leu Arg Ser His Leu Glu Pro Thr Asn (2) INFORMATION FOR SEQ ID NO:7: (i) SEQUENCE CHARACTERISTICS: (A) LENGTH: 128 base pairs (B) TYPE: nucleic acid (C) STRANDEDNESS: single

(D) TOPOLOGY: linear

(ii) MOLECULE TYPE: DNA (genomic)	
(vi) ORIGINAL SOURCE: (A) ORGANISM: Homo Sapiens	
(vii) IMMEDIATE SOURCE: (B) CLONE: VL-1	:
(ix) FEATURE: (A) NAME/KEY: CDS (B) LOCATION: 28102	
(xi) SEQUENCE DESCRIPTION: SEQ ID NO:7:	
GGATCCTGGG ATGACTGGAT TATGGCG CCG CTG GAC TAC GAG GCG TAC CAC Pro Leu Asp Tyr Glu Ala Tyr His 1 5	51
TGC GAG GGT GTA TGC GAC TTC CCG CTG CGC TCG CAC CTG GAG CCC ACC Cys Glu Gly Val Cys Asp Phe Pro Leu Arg Ser His Leu Glu Pro Thr 10 15 20	99
AAC CACGCCATGC TACAAACGCT TCTAGA Asn 25	128
(2) INFORMATION FOR SEQ ID NO:8:	
(i) SEQUENCE CHARACTERISTICS:(A) LENGTH: 25 amino acids(B) TYPE: amino acid(D) TOPOLOGY: linear	
(ii) MOLECULE TYPE: protein	
(xi) SEQUENCE DESCRIPTION: SEQ ID NO:8:	
Pro Leu Asp Tyr Glu Ala Tyr His Cys Glu Gly Val Cys Asp Phe Pro 1 5 10 15	
Leu Arg Ser His Leu Glu Pro Thr Asn 20 25	
(2) INFORMATION FOR SEQ ID NO:9:	
(i) SEQUENCE CHARACTERISTICS: (A) LENGTH: 3585 base pairs (B) TYPE: nucleic acid (C) STRANDEDNESS: single (D) TOPOLOGY: linear	
(ii) MOLECULE TYPE: DNA (genomic)	
(vii) IMMEDIATE SOURCE: (B) CLONE: pALV1-781	
(xi) SEQUENCE DESCRIPTION: SEQ ID NO:9:	
CTAACTACCC AACTCAAAAA AAAAAAAAAA AAAAACCCCC TCTAACCCCC ATTGACGAAA	60

GGGCCTCGTG ATACGCCTAT TTTTATAGGT TAATGTCATG ATAATAATGG TTTCTTAGAC

GTCAGGTGGC ACTITTCGGG GAAATGTGCG CGGAACCCCT ATTTGTTTAT TTTTCTAAAT

120

180

ACATTCAAAT	ATGTATCCGC	TCATGAGACA	ATAACCCTGA	TAAATGCTTC	AATAATATTG	240
AAAAAGGAAG	AGTATGAGTA	TTCAACATTT	CCGTGTCGCC	CTTATTCCCT	TTTTTGCGGC	300
ATTTTGCCTT	CCTGTTTTTG	CTCACCCAGA	AACGCTGGTG	AAAGTAAAAG	ATGCTGAAGA	360
TCAGTTGGGT	GCACGAGTGG	GTTACATCGA	ACTGGATCTC	AACAGCGGTA	AGATCCTTGA	420
GAGTTTTCGC	CCCGAAGAAC	GTTTTCCAAT	GATGAGCACT	TTTAAAGTTC	TGCTATGTGG	480
CGCGGTATTA	TCCCGTATTG	ACGCCGGGCA	AGAGCAACTC	GGTCGCCGCA	TAÇACTATTC	540
TCAGAATGAC	TTGGTTGAGT	ACTCACCAGT	CACAGAAAAG	CATCTTACGG	ATGGCATGAC	600
AGTAAGAGAA	TTATGCAGTG	CTGCCATAAC	CATGAGTGAT	AACACTGCGG	CCAACTTACT	660
TCTGACAACG	ATCGGAGGAC	CGAAGGAGCT	AACCGCTTTT	TTGCACAACA	TGGGGGATCA	720
TGTAACTCGC	CTTGATCGTT	GGGAACCGGA	GCTGAATGAA	GCCATACCAA	ACGACGAGCG	780
TGACACCACG	ATGCCTGTAG	CAATGGCAAC	AACGTTGCGC	AAACTATTAA	CTGGCGAACT	840
ACTTACTCTA	GCTTCCCGGC	AACAATTAAT	AGACTGGATG	GAGGCGGATA	AAGTTGCAGG	900
ACCACTTCTG	CGCTCGGCCC	TTCCGGCTGG	CTGGTTTATT	GCTGATAAAT	CTGGAGCCGG	960
TGAGCGTGGG	TCTCGCGGTA	TCATTGCAGC	ACTGGGGCCA	GATGGTAAGC	CCTCCCGTAT	1020
CGTAGTTATC	TACACGACGG	GGAGTCAGGC	AACTATGGAT	GAACGAAATA	GACAGATCGC	1080
TGAGATAGGT	GCCTCACTGA	TTAAGCATTG	GTAACTGTCA	GACCAAGTTT	ACTCATATAT	1140
ACTTTAGATT	GATTTAAAAC	TTCATTTTTA	ATTTAAAAGG	ATCTAGGTGA	AGATCCTTTT	1200
TGATAATCTC	ATGACCAAAA	TCCCTTAACG	TGAGTTTTCG	TTCCACTGAG	CGTCAGACCC	1260
CGTAGAAAAG	ATCAAAGGAT	CTTCTTGAGA	TCCTTTTTTT	CTGCGCGTAA	TCTGCTGCTT	1320
GCAAACAAAA	AAACCACCGC	TACCAGCGGT	GGTTTGTTTG	CCGGATCAAG	AGCTACCAAC	1380
TCTTTTTCCG	AAGGTAACTG	GCTTCAGCAG	AGCGCAGATA	CCAAATACTG	TCCTTCTAGT	1440
GTAGCCGTAG	TTAGGCCACC	ACTTCAAGAA	CTCTGTAGCA	CCGCCTACAT	ACCTCGCTCT	1500
GCTAATCCTG	TTACCAGTGG	CTGCTGCCAG	TGGCGATAAG	TCGTGTCTTA	CCGGGTTGGA	1560
CTCAAGACGA	TAGTTACCGG	ATAAGGCGCA	GCGGTCGGGC	TGAACGGGGG	GTTCGTGCAC	1620
ACAGCCCAGC	TTGGAGCGAA	CGACCTACAC	CGAACTGAGA	TACCTACAGC	GTGAGCATTG	1680
AGAAAGCGCC	ACGCTTCCCG	AAGGGAGAAA	GGCGGACAGG	TATCCGGTAA	GCGGCAGGGT	1740
CGGAACAGGA	GAGCGCACGA	GGGAGCTTCC	AGGGGGAAAC	GCCTGGTATC	TTTATAGTCC	1800
TGTCGGGTTT	CGCCACCTCT	GACTTGAGCG	TCGATTTTTG	TGATGCTCGT	CAGGGGGGCG	1860
GAGCCTATGG	AAAAACGCCA	GCAACGCGGC	CTTTTTACGG	TTCCTGGCCT	TTTGCTGGCC	1920
TTTTGCTCAC	ATGTTCTTTC	CTGCGTTATC	CCCTGATTCT	GTGGATAACC	GTATTACCGC	1980
CTTTGAGTGA	GCTGATACCG	CTCGCCGCAG	CCGAACGACC	GAGCGCAGCG	AGTCAGTGAG	2040
CGAGGAAGCG	GAAGAGCGCC	CAATACGCAA	ACCGCCTCTC	CCCGCGCGTT	GGCCGATTCA	2100
TTAATGCAGA	ATTGATCTCT	CACCTACCAA	ACAATGCCCC	CCTGCAAAAA	ATAAATTCAT	2160
АТААААААСА	TACAGATAAC	CATCTGCGGT	GATAAATTAT	CTCTGGCGGT	GTTGACATAA	2220

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ATACCACTGG	CGGTGATACT	GAGCACATCA	GCAGGACGCA	CTGACCACCA	TGAAGGTGAC	2280
GCTCTTAAAA	ATTAAGCCCT	GAAGAAGGGC	AGCATTCAAA	GCAGAAGGCT	TTGGGGTGTG	2340
TGATACGAAA	CGAAGCATTG	GCCGTAAGTG	CGATTCCGGA	TTAGCTGCCA	ATGTGCCAAT	2400
CGCGGGGGGT	TTTCGTTCAG	GACTACAACT	GCCACACACC	ACCALAGCTA	ACTGACAGGA	2460
GAATCCAGAT	GGATGCACAA	ACACGCCGCC	GCGAACGTCG	CGCAGAGAAA	CAGGCTCAAT	2520
GGAAAGCAGC	AAATCCCCTG	TTGGTTGGGG	TAAGCGCAAA	ACCAGTTCCG	AAAGATTTTT	2580
TTAACTATAA	ACGCTGATGG	AAGCGTTTAT	GCGGAAGAGG	TAAAGCCCTT	CCCGAGTAAC	2640
ааааааасаа	CAGCATAAAT	AACCCCGCTC	TTACACATTC	CAGCCCTGAA	AAAGGGCATC	2700
AAATTAAACC	ACACCTATGG	TGTATGCATT	TATTTGCATA	CATTCAATCA	ATTGTTATCT	2760
AAGGAAATAC	TTACATATGT	CTCGTTGTTC	TCGTAAACCA	CTGCATGTAG	ATTTTAAAGA	2820
GCTCGGCTGG	GACGACTGGA	TCATCGCGCC	GCTGGACTAC	GAGGCGTACC	ACTGCGAGGG	2880
CCTTTGCGAC	TTCCCTTTGC	GTTCGCACCT	CGAGCCCACC	AACCATGCCA	TCATTCAGAC	2940
GCTGCTCAAC	TCCATGGCAC	CAGACGCGGC	GCCGGCCTCC	TGCTGTGTGC	CAGCGCGCCT	3000
CAGCCCCATC	AGCATCCTCT	ACATCGACGC	CGCCAACAAC	GTTGTCTACA	AGCAATACGA	3060
GGACATGGTG	GTGGAGGCCT	GCGGCTGCAG	GTAGTCTAGA	GTCGACCTGC	AGTAATCGTA	3120
CAGGGTAGTA	CAAATAAAAA	AGGCACGTCA	GATGACGTGC	CTTTTTTCTT	GTGAGCAGTA	3180
AGCTTGGCAC	TGGCCGTCGT	TTTACAACGT	CGTGACTGGG	AAAACCCTGG	CGTTACCCAA	3240
CTTAATCGCC	TTGCAGCACA	TCCCCCTTTC	GCCAGCTGGC	GTAATAGCGA	AGAGGCCCGC	3300
ACCGATCGCC	CTTCCCAACA	GTTGCGCAGC	CTGAATGGCG	AATGGCGCCT	GATGCGGTAT	3360
TTTCTCCTTA	CGCATCTGTG	CGGTATTTCA	CACCGCATAT	ATGGTGCACT	CTCAGTACAA	3420
TCTGCTCTGA	TGCCGCATAG	TTAAGCCAGC	CCCGACACCC	GCCAACACCC	GCTGACGCGC	3480
CCTGACGGGC	TTGTCTGCTC	CCGGCATCCG	CTTACAGACA	AGCTGTGACC	GTCTCCGGGA	3540
GCTGCATGTG	TCAGAGGTTT	TCACCGTCAT	CACCGAAACG	CGCGA		3585

(2) INFORMATION FOR SEQ ID NO:10:

- (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 272 base pairs
 (B) TYPE: nucleic acid
 (C) STRANDEDNESS: single
 (D) TOPOLOGY: linear
- (ii) MOLECULE TYPE: DNA (genomic)
- (vi) ORIGINAL SOURCE:
 - (A) ORGANISM: mouse
- (vii) IMMEDIATE SOURCE:
 - (B) CLONE: mV1
- (ix) FEATURE:

 - (A) NAME/KEY: CDS
 (B) LOCATION: 28..243

	(xi)) SE(QUENC	CE DI	ESCR	[PTI	ON: 8	SEQ :	ID NO	0:10	:					
GGA!	rccai	AGG 2	AGCT	CGGC	rg go	BAÇG!		GG AT rp II								51
GAG Glu	GCA Ala 10	TAC Tyr	CAC His	TGC Cys	GAG Glu	GGC Gly 15	GTT Val	TGC Cys	GAC Asp	TTT Phe	CCT Pro 20	CTG Leu	CGC Arg	TCG Ser	CAC His	99
		CCT Pro														147
		GAC Asp														195
		AGC Ser														243
CAA:	racg)	AGG A	ACATO	GTG	ST GO	GGA.	ATTC									272
(2)	INF	ORMAT	CION	FOR	SEQ	ID I	10:13	1:								
		(i) £	(A)	ENCE LEI TYI	NGTH	: 72 amin	amin	no ao id	-							
	(:	ii) N	OLE	CULE	TYP	E: p:	rote:	in								
					220											

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:11:

Trp Ile Ile Ala Pro Leu Asp Tyr Glu Ala Tyr His Cys Glu Gly Val

Cys Asp Phe Pro Leu Arg Ser His Leu Glu Pro Thr Asn His Ala Ile

Ile Gln Thr Leu Leu Asn Ser Met Ala Pro Asp Ala Ala Pro Ala Ser
35 40 45

Cys Cys Val Pro Ala Arg Leu Ser Pro Ile Ser Ile Leu Tyr Ile Asp 50 60

Ala Ala Asn Asn Val Val Tyr Lys

- (2) INFORMATION FOR SEQ ID NO:12:
 - (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 272 base pairs
 - (B) TYPE: nucleic acid
 - (C) STRANDEDNESS: single
 - (D) TOPOLOGY: linear
 - (ii) MOLECULE TYPE: DNA (genomic)
 - (vi) ORIGINAL SOURCE:
 - (A) ORGANISM: mouse
 - (vii) IMMEDIATE SOURCE:
 - (B) CLONE: mV2

	(ix)		1) N2	E: AME/I CATI			. 243										
	(xi)	SEC	QUENC	E DE	SCRI	PTIC	ON: 5	SEQ I	D NO):12:	:					·	
GGAT	rcca;	AGG 1	AGCTO	cgci	rg go	BACG								AG TI lu Ty			5
				TGC Cys													9
				AAC Asn												1	4
				ACC Thr 45												1	9
				CTG Leu												2	4
CAAT	racg <i>i</i>	AGG 1	CATO	GTG	er Go	GGAJ	ATTC									2	7
(2)			SEQUI (A) (B)	FOR ENCE LEI TYI	CHAI	RACTI 72	ERIS' amii	TICS: no ac id									
	(:	ii) P	OLE	TULE	TYPI	2: p	rote:	in									
	(2	ki) 8	SEQUI	ENCE	DESC	RIP	rion	: SE	Q ID	NO:	13:						
Trp 1	Ile	Ile	Ala	Pro 5	Leu	Glu	Tyr	Glu	Ala 10	Tyr	His	Cys	Glu	Gly 15	Val		
Сув	Asp	Phe	Pro 20	Leu	Arg	Ser	His	Leu 25	Glu	Pro	Thr	Asn	His 30	Ala	Ile		
Ile	Gln	Thr 35	Leu	Met	Asn	Ser	Met 40	Asp	Pro	Gly	Ser	Thr 45	Pro	Pro	Ser		
Cys	Cys 50	Val	Pro	Thr	Lys	Leu 55	Thr	Pro	Ile	Ser	Ile 60	Leu	Tyr	Ile	Asp		
Ala 65	Gly	Asn	Asn	Val	Val 70	Tyr	Lys										
(2)	INF	ORMA'	rion	FOR	SEQ	ID 1	NO:1	4:									
	(i)	() ()	A) LI B) T C) S'	CE CI ENGTI YPE: TRANI	nuc DEDN	72 b lei¢ ESS:	ase aci sin	pair d	s								

(vi) ORIGINAL SOURCE:

		(A) O	RGAN	ISM:	mou	se									
	(vii		MEDI B) C													
	(ix	(.	ATURI A) NI B) L	AME/			. 243									
	(xi) SE	QUEN	CE DI	ESCR:	IPTI	ON:	SEQ :	ID N	0:14	:					
GGA	TCCA	AGG /	AGCT	CGGC'	rg g(GACG				TC GO						51
GAG Glu	GCC Ala 10	TTC Phe	CAC His	TGC Cys	GAA Glu	GGA Gly 15	CTG Leu	TGT Cys	GAG Glu	TTC Phe	CCC Pro 20	TTG Leu	CGC Arg	TCC Ser	CAC His	99
TTG Leu 25	GAG Glu	CCC Pro	ACA Thr	AAC Asn	CAC His 30	GCA Ala	GTC Val	ATT Ile	CAG Gln	ACC Thr 35	CTA Leu	ATG Met	AAC Asn	TCT Ser	ATG Met 40	147
GAC Asp	CCT Pro	GAA Glu	TCC Ser	ACA Thr 45	CCA Pro	CCC Pro	ACT Thr	TGT Cys	TGT Cys 50	GTG Val	CCT Pro	ACA Thr	CGG Arg	CTG Leu 55	AGT Ser	195
CCT Pro	ATT Ile	AGC Ser	ATC. Ile 60	CTC Leu	TTC Phe	ATC Ile	GAC Asp	TCT Ser 65	GCC Ala	AAC Asn	AAC Asn	GTG Val	GTG Val 70	TAT Tyr	AAA Lys	243
CAA	TACG	AGG 2	ACATO	GTG	et G	GGA	ATTC									272
(2)	INF	ORMA!	rion	FOR	SEQ	ID 1	VO:15	5:								
		(i) :	(B)	LEN TYI	NGTH:	RACTI 72 mind	amir aci	o ad id								
	(:	li) M	MOLEC	ULE	TYPE	E: pı	otei	in								
	(:	ci) S	SEQUE	NCE	DESC	RIPT	NOI?	SEC) ID	NO:1	.5 :					
Trp 1	Ile	Ile	Ala	Pro 5	Leu	Glu	Tyr	Glu	Ala 10	Phe	His	Сув	Glu	Gly 15	Leu	
Сув	Glu	Phe	Pro 20	Leu	Arg	Ser	His	Leu 25	Glu	Pro	Thr	Asn	His 30	Ala	Val	
Ile	Gln	Thr 35	Leu	Met	Asn	Ser	Met 40	qaA	Pro	Glu	Ser	Thr 45	Pro	Pro	Thr	
Сув	Сув 50	Val	Pro	Thr	Arg	Leu 55	Ser	Pro	Ile	Ser	Ile 60	Leu	Phe	Ile	ĄaĄ	
65			Asn		70											
(2)			NOIT													
	(i)	(2	QUENC A) LE B) TY	NGT	1: 7	amir	o ac	ids	579							

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(C) STRANDEDNESS: single
          (D) TOPOLOGY: linear
    (ii) MOLECULE TYPE: peptide
    (vi) ORIGINAL SOURCE:
          (A) ORGANISM: BMP/TGF-beta consensus sequence
    (xi) SEQUENCE DESCRIPTION: SEQ ID NO:16:
     Trp Xaa Asp Trp Ile Xaa Ala
(2) INFORMATION FOR SEQ ID NO:17:
     (i) SEQUENCE CHARACTERISTICS:
          (A) LENGTH: 27 base pairs
          (B) TYPE: nucleic acid
          (C) STRANDEDNESS: single
          (D) TOPOLOGY: linear
    (ii) MOLECULE TYPE: DNA (genomic)
   (vii) IMMEDIATE SOURCE:
          (B) CLONE: oligonucleotide #1
    (xi) SEQUENCE DESCRIPTION: SEQ ID NO:17:
CGGATCCTGG VANGAYTGGA THRTNGC
                                                                           27
(2) INFORMATION FOR SEO ID NO:18:
     (i) SEQUENCE CHARACTERISTICS:
           (A) LENGTH: 6 amino acids
           (B) TYPE: amino acid
          (C) STRANDEDNESS: single (D) TOPOLOGY: linear
    (ii) MOLECULE TYPE: peptide
   (vii) IMMEDIATE SOURCE:
          (B) CLONE: BMP/TGF-beta consensus sequence
    (xi) SEQUENCE DESCRIPTION: SEQ ID NO:18:
     His Ala Ile Xaa Gln Thr
                      5
     1
(2) INFORMATION FOR SEQ ID NO:19:
     (i) SEQUENCE CHARACTERISTICS:
          (A) LENGTH: 28 base pairs (B) TYPE: nucleic acid
           (C) STRANDEDNESS: single
           (D) TOPOLOGY: linear
```

(vii) IMMEDIATE SOURCE:

(B) CLONE: oligonucleotide #2

(ii) MOLECULE TYPE: DNA (genomic)

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:19:	
TTTCTAGAAR NGTYTGNACD ATNGCRTG	28
(2) INFORMATION FOR SEQ ID NO:20:	
(i) SEQUENCE CHARACTERISTICS: (A) LENGTH: 40 base pairs (B) TYPE: nucleic acid (C) STRANDEDNESS: single (D) TOPOLOGY: linear	
(ii) MOLECULE TYPE: DNA (genomic)	
<pre>(vii) IMMEDIATE SOURCE: (B) CLONE: oligonucleotide #3</pre>	
(xi) SEQUENCE DESCRIPTION: SEQ ID NO:20:	
CCACTGCGAG GGCCTTTGCG ACTTCCCTTT GCGTTCGCAC	40
(2) INFORMATION FOR SEQ ID NO:21:	
(i) SEQUENCE CHARACTERISTICS: (A) LENGTH: 29 base pairs (B) TYPE: nucleic acid (C) STRANDEDNESS: single (D) TOPOLOGY: linear	
(ii) MOLECULE TYPE: DNA (genomic)	
(vii) IMMEDIATE SOURCE: (A) LIBRARY: oligonucleotide #4	
(xi) SEQUENCE DESCRIPTION: SEQ ID NO:21:	
TGCGGATCCA GCCGCTGCAG CCGCAAGCC	29
(2) INFORMATION FOR SEQ ID NO:22:	
(i) SEQUENCE CHARACTERISTICS: (A) LENGTH: 29 base pairs (B) TYPE: nucleic acid (C) STRANDEDNESS: single (D) TOPOLOGY: linear	
(ii) MOLECULE TYPE: DNA (genomic)	
(vii) IMMEDIATE SOURCE: (B) CLONE: oligonucleotide #5	
(xi) SEQUENCE DESCRIPTION: SEQ ID NO:22:	
GACTCTAGAC TACCTGCAGC CGCAGGCCT	29
(2) INFORMATION FOR SEQ ID NO:23:	
(i) SEQUENCE CHARACTERISTICS: (A) LENGTH: 28 base pairs (B) TYPE: nucleic acid (C) STRANDEDNESS: single	

(D) TOPOLOGY: linear (ii) MOLECULE TYPE: DNA (genomic)

GCGGATCCAA GGAGCTCGGC TGGGACGA

(vii) IMMEDIATE SOURCE:
 (A) LIBRARY: oligonucleotide #6

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:23:

GCGGATCCAA GGAGCTCGGC TGGGACGA	28
(2) INFORMATION FOR SEQ ID NO:24:	
(i) SEQUENCE CHARACTERISTICS: (A) LENGTH: 28 base pairs (B) TYPE: nucleic acid (C) STRANDEDNESS: single (D) TOPOLOGY: linear	
(ii) MOLECULE TYPE: DNA (genomic)	
(vii) IMMEDIATE SOURCE: (B) CLONE: oligonucleotide #7	
(xi) SEQUENCE DESCRIPTION: SEQ ID NO:24:	
GGAATTCCCC ACCACCATGT CCTCGTAT	28
(2) INFORMATION FOR SEQ ID NO:25:	
(i) SEQUENCE CHARACTERISTICS: (A) LENGTH: 1171 base pairs (B) TYPE: nucleic acid (C) STRANDEDNESS: single (D) TOPOLOGY: linear	
(ii) MOLECULE TYPE: DNA (genomic)	
(vii) IMMEDIATE SOURCE: (B) CLONE: Human VL-1 protein	
(ix) FEATURE: (A) NAME/KEY: CDS (B) LOCATION: 2964	
<pre>(ix) FEATURE: (A) NAME/KEY: mat_peptide (B) LOCATION: 605964</pre>	
(xi) SEQUENCE DESCRIPTION: SEQ ID NO:25:	
AST TCG GAT CTC TCG CAC ACT CCT CTC CGG AGA CAG AAG TAT TTG ASN Ser Asp Leu Ser His Thr Pro Leu Arg Arg Gln Lys Tyr Leu -201-200 -195 -190	46
Phe Asp Val Ser Met Leu Ser Asp Lys Glu Glu Leu Val Gly Ala Glu -185 -180 -175	94

	Arg	CTC Leu				Ala					Trp					142
GGG	CCG Pro	CTC Leu	CAC His	GTG Val -15	Gln	CTC Leu	TTC Phe	CCT Pro	TGC Cys -14	Leu	TCG Ser	CCC Pro	CTA Leu	CTG Leu -14	Leu	190
GAC Asp	GCG Ala	CGG Arg	ACC Thr	Leu	GAC Asp	CCG Pro	CAG Gln	GGG Gly -13	Ala	CCG Pro	CCG Pro	GCC Ala	GGC Gly -12	Trp	GAA Glu	238
GTC Val	TTC Phe	GAC Asp -12	Val	TGG Trp	CAG Gln	GGC Gly	CTG Leu -11	Arg	CAC His	CAG Gln	CCC Pro	TGG Trp -11	Lys	CAG Gln	CTG Leu	286
TGC Cys	TTG Leu -10	GAG Glu 5	CTG Leu	CGG Arg	GCC Ala	GCA Ala -100	Trp	GGC Gly	GAG Glu	CTG Leu	GAC Asp -95	GCC Ala	GGG Gly	GAG Glu	GCC Ala	334
GAG Glu -90	GCG Ala	CGC Arg	GCG Ala	CGG Arg	GGA Gly -85	CCC Pro	CAG Gln	CAA Gln	CCG Pro	CCG Pro -80	CCC Pro	CCG Pro	GAC Asp	CTG Leu	CGG Arg -75	382
AGT Ser	CTG Leu	GGC Gly	TTC Phe	GGC Gly -70	CGG Arg	AGG Arg	GTG Val	CGG Arg	CCT Pro -65	CCC Pro	CAG Gln	GAG Glu	CGG Arg	GCC Ala -60	CTG Leu	430
CTG Leu	GTG Val	GTA Val	TTC Phe -55	ACC Thr	AGA Arg	TCC Ser	CAG Gln	CGC Arg -50	AAG Lys	AAC Asn	CTG Leu	TTC Phe	GCA Ala -45	GAG Glu	ATG Met	478
CGC Arg	GAG Glu	CAG Gln -40	CTG Leu	GGC Gly	TCG Ser	GCC Ala	GAG Glu -35	GCT Ala	GCG Ala	GGC Gly	CCG Pro	GGC Gly -30	GCG Ala	GGC Gly	GCC Ala	526
GAG Glu	GGG Gly -25	TCG Ser	TGG Trp	CCG Pro	CCG Pro	CCG Pro -20	TCG Ser	GGC Gly	GCC Ala	CCG Pro	GAT Asp -15	GCC Ala	AGG Arg	CCT Pro	TGG Trp	574
CTG Leu -10	CCC Pro	TCG Ser	CCC Pro	GGC Gly	CGC Arg -5	CGG Arg	CGG Arg	CGG Arg	CGC Arg	ACG Thr 1	GCC Ala	TTC Phe	GCC Ala	AGT Ser 5	CGC Arg	622
CAT His	GGC Gly	AAG Lys	CGG Arg 10	CAC His	GGC Gly	AAG Lys	AAG Lys	TCC Ser 15	AGG Arg	CTA Leu	CGC Arg	TGC Cys	AGC Ser 20	AAG Lys	AAG Lys	670
CCC Pro	CTG Leu	CAC His 25	GTG Val	AAC Asn	TTC Phe	AAG Lys	GAG Glu 30	CTG Leu	GGC Gly	TGG Trp	GAC Asp	GAC Asp 35	TGG Trp	ATT Ile	ATC Ile	718
GCG Ala	CCC Pro 40	CTG Leu	GAG Glu	TAC Tyr	GAG Glu	GCC Ala 45	TAT Tyr	CAC His	TGC Cys	GAG Glu	GGT Gly 50	GTA Val	TGC Cys	GAC Asp	TTC Phe	766
CCG Pro 55	CTG Leu	CGC Arg	TCG Ser	CAC His	CTG Leu 60	GAG Glu	CCC Pro	ACC Thr	AAC Asn	CAC His 65	GCC Ala	ATC Ile	ATC Ile	CAG Gln	ACG Thr 70	814
CTG Leu	ATG Met	AAC Asn	TCC Ser	ATG Met 75	GAC Asp	CCC Pro	GGC Gly	TCC Ser	ACC Thr 80	CCG Pro	CCC Pro	AGC Ser	TGC Cys	TGC Cys 85	GTG Val	862
CCC Pro	ACC Thr	AAA Lys	TTG Leu 90	ACT Thr	CCC Pro	ATC Ile	AGC Ser	ATT Ile 95	Leu	TAC Tyr	ATC Ile	GAC Asp	GCG Ala 100	GGC Gly	AAT Asn	910

AAT Asn	GTG Val	GTC Val 105	TAC Tyr	AAG Lys	CAG Gln	TAC Tyr	GAG Glu 110	GAC Asp	ATG Met	GTG Val	Val	GAG Glu 115	rcg Ser	TGC Cys	GGC Gly
TGC Cys		TAGC	GGTG	CC T	TTCC	CGCC	eg co	TTGG	CCCG	GAA	.CCAA	GGT	GGGC	CAAG	GT
CCGC	CTTG	CA G	GGGA	GGCC	T GG	CTGC	AGAG	AGG	CGGA	.GGA	GGAA	.GCTG	GC G	CTGG	GGGAG
GCTG	AGGG	TG A	GGGA	ACAG	C CI	GGAI	GTG	A GAG	CCGG	TGG	GAGA	GAAG	GG A	GCGC	ACCTT
CCCA	.GTAA	CT I	CTAC	CTGC	C AG	CCCA	GAGG	G GAA	TATA	•					
(2)	INFO	RMAI	NOI	FOR	SEQ	ID N	10:26	5 : '							
	((i) S	(A) (B)	ENCE LEN TYP TOP	IGTH: PE: &	321 mino	ami	ino a id		1					
	()	ii) M	OLEC	TULE	TYPE	e: pr	ote:	in							
	()	ci) S	EQUE	ENCE	DESC	RIP	CION	: SE() ID	NO:2	:6:				
	Ser -20	_	Leu	Ser	His		Pro 195	Leu	Arg	Arg		Lys 190	Tyr	Leu	Phe
Asp -185		Ser	Met	Leu	Ser -18	_	Lys	Glu	Glu		Val 175	Gly	Ala	Glu	Leu -170
Arg	Leu	Phe	Arg	Gln -165		Pro	Ser	Ala	Pro -16		Gly	Pro	Pro		Gly 155
Pro	Leu	His	Val -150	Gln)	Leu	Phe	Pro	Cys -1		Ser	Pro	Leu		Leu 140	Asp
Ala	Arg	Thr -135		Asp	Pro	Gln	Gly -1		Pro	Pro	Ala		Trp L25	Glu	Val
Phe	Asp -120		Trp	Gln	Gly	Leu -1		His	Gln	Pro		Lys 110	Gln	Leu	Cys
Leu -105		Leu	Arg	Ala	Ala -10		Gly	Glu	Leu	Asp -9		Gly	Glu	Ala	Glu -90
Ala	Arg	Ala	Arg	Gly -85	Pro	Gln	Gln	Pro	Pro -80	Pro	Pro	Asp	Leu	Arg -75	Ser
Leu	Gly	Phe	Gly -70	Arg	Arg	Val	Arg	Pro -65	Pro	Gln	Glu	Arg	Ala -60	Leu	Leu
Val	Val	Phe -55	Thr	Arg	Ser	Gln	Arg -50	Lys	Asn	Leu	Phe	Ala -45	Glu	Met	Arg
Glu	Gln -40	Leu	Gly	Ser	Ala	Glu -35	Ala	Ala	Gly	Pro	Gly -30	Ala	Gly	Ala	Glu
Gly -25	Ser	Trp	Pro	Pro	Pro	Ser	Gly	Ala	Pro	Asp -15	Ala	Arg	Pro	Trp	Leu -10
Pro	Ser	Pro	Gly	Arg -5	Arg	Arg	Arg	Arg	Thr 1	Ala	Phe	Ala	Ser 5	Arg	His
Gly	Lys	Arg	His	Gly	Lys	Lys	Ser 15		Leu	Arg	Сув	Ser 20	Lys	Lys	Pro

Leu His Val Asn Phe Lys Glu Leu Gly Trp Asp Asp Trp Ile Ile Ala 25 30 35

Pro Leu Glu Tyr Glu Ala Tyr His Cys Glu Gly Val Cys Asp Phe Pro 40 55

Leu Arg Ser His Leu Glu Pro Thr Asn His Ala Ile Ile Gln Thr Leu
60 65 70

Met Asn Ser Met Asp Pro Gly Ser Thr Pro Pro Ser Cys Cys Val Pro
75 80 85

Thr Lys Leu Thr Pro Ile Ser Ile Leu Tyr Ile Asp Ala Gly Asn Asn 90 95 100

Val Val Tyr Lys Gln Tyr Glu Asp Met Val Val Glu Ser Cys Gly Cys 105 115

Arg 120

(2) INFORMATION FOR SEQ ID NO:27:

- (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 1233 base pairs
 - (B) TYPE: nucleic acid
 - (C) STRANDEDNESS: single
 - (D) TOPOLOGY: linear
- (ii) MOLECULE TYPE: DNA (genomic)
- (vii) IMMEDIATE SOURCE:
 - (B) CLONE: DNA encoding BMP2 propeptide/BMP-12 mature peptide
- (ix) FEATURE:
 - (A) NAME/KEY: CDS
 - (B) LOCATION: 1..1233
- (ix) FEATURE:
 - (A) NAME/KEY: mat_peptide
 - (B) LOCATION: 847..1233
- (xi) SEQUENCE DESCRIPTION: SEQ ID NO:27:
- ATG GTG GCC GGG ACC CGC TGT CTT CTA GCG TTG CTG CTT CCC CAG GTC

 Met Val Ala Gly Thr Arg Cys Leu Leu Ala Leu Leu Pro Gln Val
 -282 -280 -275 -270
- CTC CTG GGC GGC GCT GGC CTC GTT CCG GAG CTG GGC CGC AGG AAG 96
 Leu Leu Gly Gly Ala Ala Gly Leu Val Pro Glu Leu Gly Arg Arg Lys
 -265 -260 -255
- TTC GCG GCG GCG TCG TCG GGC CCC TCA TCC CAG CCC TCT GAC GAG

 Phe Ala Ala Ser Ser Gly Arg Pro Ser Ser Gln Pro Ser Asp Glu
 -250 -245 -240 -235
- GTC CTG AGC GAG TTC GAG TTG CGG CTG CTC AGC ATG TTC GGC CTG AAA 192
 Val Leu Ser Glu Phe Glu Leu Arg Leu Leu Ser Met Phe Gly Leu Lys
 -230 -225 -220
- CAG AGA CCC ACC CCC AGC AGG GAC GCC GTG GTG CCC CCC TAC ATG CTA
 Gln Arg Pro Thr Pro Ser Arg Asp Ala Val Val Pro Pro Tyr Met Leu
 -215 -210 -205

	GC AGG CAC TCA G rg Arg His Ser G			
	AG AGG GCA GCC A Diu Arg Ala Ala S -180			
CAC CAT GAA G His His Glu G -170	AA TCT TTG GAA G lu Ser Leu Glu G -165	AA CTA CCA GAA lu Leu Pro Glu -16	Thr Ser Gly I	AAA ACA 384 Lys Thr -155
	TTC TTC TTT AAT 1 The Phe Phe Asn I -150		Pro Thr Glu G	
Ile Thr Ser A	CA GAG CTT CAG G la Glu Leu Gln V 135	TT TTC CGA GAA al Phe Arg Glu -130	CAG ATG CAA G Gln Met Gln A -125	SAT GCT 480 Asp Ala
TTA GGA AAC A Leu Gly Asn A -120	AT AGC AGT TTC C sn Ser Ser Phe F	AT CAC CGA ATT is His Arg Ile 115	AST ATT TAT G AST Ile Tyr G -110	GAA ATC 528 Slu Ile
ATA AAA CCT G Ile Lys Pro A -105	CA ACA GCC AAC 1 la Thr Ala Asn 5 -100	CG AAA TTC CCC er Lys Phe Pro	GTG ACC AGA C Val Thr Arg I -95	CTT TTG 576 Leu Leu
GAC ACC AGG T Asp Thr Arg L -90	TG GTG AAT CAG A eu Val Asn Gln A -85	AT GCA AGC AGG sn Ala Ser Arg -80	Trp Glu Ser F	TTT GAT 624 Phe Asp -75
GTC ACC CCC G Val Thr Pro A	CT GTG ATG CGG T la Val Met Arg T -70	GG ACT GCA CAG rp Thr Ala Gln -65	Gly His Ala A	AAC CAT 672 Asn His 60
Gly Phe Val V	TTG GAA GTG GCC C 'al Glu Val Ala F 55	AC TTG GAG GAG is Leu Glu Glu -50	AAA CAA GGT G Lys Gln Gly V -45	TC TCC 720 Val Ser
AAG AGA CAT G Lys Arg His V -40	TT AGG ATA AGC A al Arg Ile Ser A	GG TCT TTG CAC rg Ser Leu His 35	CAA GAT GAA C Gln Asp Glu H -30	CAC AGC 768 His Ser
TGG TCA CAG A Trp Ser Gln I -25	TA AGG CCA TTG C le Arg Pro Leu I -20	TA GTA ACT TTT eu Val Thr Phe	GGC CAT GAT G Gly His Asp G -15	GA AAA 816 Hy Lys
GGG CAT CCT C Gly His Pro L -10	TC CAC AAA AGA G eu His Lys Arg G -5	AA AAA CGT ACG lu Lys Arg Thr 1	GCG TTG GCC G Ala Leu Ala G	GGG ACG 864 Sly Thr 5
Arg Thr Ala G	AG GGC AGC GGC G In Gly Ser Gly G 10	GG GGC GCG GGC ly Gly Ala Gly 15	CGG GGC CAC G Arg Gly His G	GG CGC 912 Ely Arg
AGG GGC CGG A Arg Gly Arg S 25	GC CGC TGC AGC C er Arg Cys Ser A	GC AAG CCG TTG rg Lys Pro Leu 30	CAC GTG GAC THIS Val Asp F	TC AAG 960 Phe Lys
	GG GAC GAC TGG A TP Asp Asp Trp 1 45			
	AG GGC CTT TGC G lu Gly Leu Cys A 60		Arg Ser His L	

CCC Pro	ACC Thr	AAC Asn	CAT His	GCC Ala 75	ATC Ile	ATT Ile	CAG Gln	ACG Thr	CTG Leu 80	CTC Leu	AAC Asn	TCC Ser	ATG Met	GCA Ala 85	CCA Pro	1104
													AGC Ser 100			1152
													AAG Lys			1200
		ATG Met														1233

(2) INFORMATION FOR SEQ ID NO:28:

- (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 411 amino acids
 - (B) TYPE: amino acid
 - (D) TOPOLOGY: linear
- (ii) MOLECULE TYPE: protein
- (xi) SEQUENCE DESCRIPTION: SEQ ID NO:28:

Met Val Ala Gly Thr Arg Cys Leu Leu Ala Leu Leu Leu Pro Gln Val -282 -280 -275 -270

Leu Leu Gly Gly Ala Ala Gly Leu Val Pro Glu Leu Gly Arg Arg Lys
-265 -260 -255

Phe Ala Ala Ser Ser Gly Arg Pro Ser Ser Gln Pro Ser Asp Glu
-250 -245 -240 -235

Val Leu Ser Glu Phe Glu Leu Arg Leu Leu Ser Met Phe Gly Leu Lys
-230 -225 -220

Gln Arg Pro Thr Pro Ser Arg Asp Ala Val Val Pro Pro Tyr Met Leu
-215 -210 -205

Asp Leu Tyr Arg Arg His Ser Gly Gln Pro Gly Ser Pro Ala Pro Asp
-200 -195 -190

His Arg Leu Glu Arg Ala Ala Ser Arg Ala Asn Thr Val Arg Ser Phe
-185 -180 -175

His His Glu Glu Ser Leu Glu Glu Leu Pro Glu Thr Ser Gly Lys Thr
-170 -165 -160 -15

Thr Arg Arg Phe Phe Phe Asn Leu Ser Ser Ile Pro Thr Glu Glu Phe
-150 -145 -140

Ile Thr Ser Ala Glu Leu Gln Val Phe Arg Glu Gln Met Gln Asp Ala
-135
-130
-125

Leu Gly Asn Asn Ser Ser Phe His His Arg Ile Asn Ile Tyr Glu Ile -120 -115 -110

Ile Lys Pro Ala Thr Ala Asn Ser Lys Phe Pro Val Thr Arg Leu Leu
-105 -95

Asp Thr Arg Leu Val Asn Gln Asn Ala Ser Arg Trp Glu Ser Phe Asp
-90 -85 -80 -75

Val Thr Pro Ala Val Met Arg Trp Thr Ala Gln Gly His Ala Asn His

Gly Phe Val Val Glu Val Ala His Leu Glu Glu Lys Gln Gly Val Ser

Lys Arg His Val Arg Ile Ser Arg Ser Leu His Gln Asp Glu His Ser

Trp Ser Gln Ile Arg Pro Leu Leu Val Thr Phe Gly His Asp Gly Lys

Gly His Pro Leu His Lys Arg Glu Lys Arg Thr Ala Leu Ala Gly Thr

Arg Thr Ala Gln Gly Ser Gly Gly Gly Ala Gly Arg Gly His Gly Arg

Arg Gly Arg Ser Arg Cys Ser Arg Lys Pro Leu His Val Asp Phe Lys

Glu Leu Gly Trp Asp Asp Trp Ile Ile Ala Pro Leu Asp Tyr Glu Ala

Tyr His Cys Glu Gly Leu Cys Asp Phe Pro Leu Arg Ser His Leu Glu

Pro Thr Asn His Ala Ile Ile Gln Thr Leu Leu Asn Ser Met Ala Pro

Asp Ala Ala Pro Ala Ser Cys Cys Val Pro Ala Arg Leu Ser Pro Ile

Ser Ile Leu Tyr Ile Asp Ala Ala Asn Asn Val Val Tyr Lys Gln Tyr

Glu Asp Met Val Val Glu Ala Cys Gly Cys Arg

- (2) INFORMATION FOR SEQ ID NO:29:
 - (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 1203 base pairs
 - (B) TYPE: nucleic acid
 - (C) STRANDEDNESS: single
 - (D) TOPOLOGY: linear
 - (ii) MOLECULE TYPE: DNA (genomic)
 - (vii) IMMEDIATE SOURCE:
 - (B) CLONE: murine MV1
 - (ix) FEATURE:
 - (A) NAME/KEY: CDS
 - (B) LOCATION: 2..721
 - (xi) SEQUENCE DESCRIPTION: SEQ ID NO:29:
- A AAG TTC TGC CTG GTG CTG GNG NCG GTG ACG GCC TCG GAG AGC AGN Lys Phe Cys Leu Val Leu X01 X02 Val Thr Ala Ser Glu Ser X03
- CNG CTG GCC CTG AGA CGA CTG GGC TTC GGC TGN CCG GGC GGT GGC GAC X04 Leu Ala Leu Arg Arg Leu Gly Phe Gly X05 Pro Gly Gly Asp 25 68 20

94

46

				GCG Ala												142
CGT Arg	ACG Thr	CAA Gln 50	AGG Arg	AAA Lys	GAG Glu	AGT Ser	CTG Leu 55	TTC Phe	CGG Arg	GAG Glu	ATC Ile	CGA Arg 60	GCC Ala	CAG Gln	GCC Ala	190
				GCC Ala												238
				AAA Lys												286
				ACT Thr 100												334
				GGC Gly												382
				GGG Gly												430
				TTT Phe												478
				GAG Glu												526
				CTG Leu 180												574
				GCG Ala												622
				CCC Pro									_			670
				CAG Gln												718
AGG Arg 240	TAG	CATG	CGG '	rctg	3GGA	GG G'	rctg(GCCG	c cc	AGGA	CCCT	AGC	TCAA	GAG		771
CAG	GTGT(CAT (CAGG	ccca	AG G	GACG	GCGG:	A CT	ATGG	CCTC	TGC	CAGC	ACA (GAGG	AGAGCA	831
CAC	AGTT	AAC 2	ACTC	ACAT	TT A	CACA	CTCC'	T TC	ACTC.	ACGC	ACA	TGTT	TAC	CGTG	GACGGC	891
AGG	CGCT	AAA	AGCC	TTGC	TT A	TTTG	CTAC	C AT	TGAT.	ACAA	ACC	TCTG	TCC	TTTT	CGGGAG	951
AGG	GAAG	GGC :	ATCT	GTGT	TT A	TGTT	GCAG	AA T	TTGG	CACT	AAA	TCCA	AGT .	AGAA	ATGGGT	1011
TAG	CATT	GGA '	TTCT	CCTT	TT A	GTTG	GAGG	C GG	TGTG	GCTG	GAT	TCCT	GAC	GTTG	GATATG	107
GAG	TGCA	CTG	CAGG	GCTG	GG A	TACC	CAGA	T TC	ŢĊŢĠ	GAGT	GGG	CATT	GGG	AACC	TTCAAA	1131

PCT/US94/14030 WO 95/16035

AGTAAGGAGC CACTGGGGCT TGGGAGGGAG CACCCGGTTC CTAAACAAGT CTGATGTGTA 1191 1203 CTGCTCAGTT TG

(2) INFORMATION FOR SEQ ID NO:30:

- (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 240 amino acids
 - (B) TYPE: amino acid
 - (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: protein

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:30:

Lys Phe Cys Leu Val Leu X01 X02 Val Thr Ala Ser Glu Ser X03 X04

Leu Ala Leu Arg Arg Leu Gly Phe Gly X05 Pro Gly Gly Asp Gly

Gly Gly Thr Ala X06 Glu Glu Arg Ala Leu Leu Val Ile Ser Ser Arg

Thr Gln Arg Lys Glu Ser Leu Phe Arg Glu Ile Arg Ala Gln Ala Arg

Ala Leu Arg Ala Ala Ala Glu Pro Pro Pro Asp Pro Gly Pro Gly Ala

Gly Ser Arg Lys Ala Asn Leu Gly Gly Arg Arg Arg Gln Arg Thr Ala

Leu Ala Gly Thr Arg Gly X07 X08 Gly Ser Gly Gly Gly Gly Gly Gly

Arg Gly His Gly Arg Arg Gly Arg Ser Arg Cys Gly Arg Lys Ser Leu

His Val Asp Phe Lys Glu Leu Gly Trp Asp Asp Trp Ile Ile Ala Pro 145

Leu Asp Tyr Glu Ala Tyr His Cys Glu Gly Val Cys Asp Phe Pro Leu

Arg Ser His Leu Glu Pro Thr Asn His Ala Ile Ile Gln Thr Leu Leu

Asn Ser Met Ala Pro Asp Ala Ala Pro Ala Ser Cys Cys Val Pro Ala 200

Arg Leu Ser Pro Ile Ser Ile Leu Tyr Ile Asp Ala Ala Asn Asn Val

Val Tyr Lys Gln Tyr Glu Asp Met Val Val Glu Ala Cys Gly Cys Arg

(2) INFORMATION FOR SEQ ID NO:31:

- (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 1046 base pairs
 - (B) TYPE: nucleic acid

(C)	STRANDEDNESS:	single
	TODOLOGY. 1400	

- (ii) MOLECULE TYPE: DNA (genomic)
- (iii) HYPOTHETICAL: NO
- (iv) ANTI-SENSE: NO
- (vii) IMMEDIATE SOURCE:
 - (B) CLONE: MURINE MV2
- (ix) FEATURE:
 - (A) NAME/KEY: CDS (B) LOCATION: 2..790
- (xi) SEQUENCE DESCRIPTION: SEQ ID NO:31:

A AGA AAA CAA GC Arg Lys Gln Al	T TGC ATT CCT G a Cys Ile Pro A 5	CA GGT CCG AC la Gly Pro Th	CT CTA AGA GGA TC or Leu Arg Gly Se 1	r
TCA GGG ACC CAA Ser Gly Thr Gln	CCC AGG CCG GCT Pro Arg Pro Ala 20	GGG AAG TCT Gly Lys Ser 25	TTC GAC GTG TGG Phe Asp Val Trp	CAG 94 Gln
GGC CTG CGC CCT Gly Leu Arg Pro	CAG CCT TGG AAG Gln Pro Trp Lys	CAG CTG TGC Gln Leu Cys 40	CTG GAG TTG CGG Leu Glu Leu Arg	GCA 142 Ala
GCC TGG GGT GAG Ala Trp Gly Glu	CTG GAC RCC GGG Leu Asp X01 Gly 55	Asp Thr Gly	GCG CGC GCG AGG Ala Arg Ala Arg	GGT 190 Gly
CCC CAG CAG CCA Pro Gln Gln Pro	CCG CCT CTG GAC Pro Pro Leu Asp 70	CTG CGG AGT Leu Arg Ser	CTG GGC TTC GGT (Leu Gly Phe Gly 75	CGG 238 Arg
AGG GTG AGA CCG Arg Val Arg Pro : 80	CCC CAG GAG CGC Pro Gln Glu Arg 85	GCC CTG CTT Ala Leu Leu 90	GTA GTG TTC ACC AVAI Val Phe Thr	AGA 286 Arg 95
TCG CAG CGC AAG AS Ser Gln Arg Lys	AAC CTG TTC ACT Asn Leu Phe Thr 100	GAG ATG CAT Glu Met His 105	GAG CAG CTG GGC S Glu Gln Leu Gly S 110	TCT 334 Ser
GCA GAG GCT GCG (Ala Glu Ala Ala (115	GGA GCC GAG GGG Gly Ala Glu Gly	TCA TGT CCA Ser Cys Pro 120	GCG CCG TCG GGC : Ala Pro Ser Gly : 125	TCC 382 Ser
Pro Asp Thr Gly 9	TCT TGG CTG CCC Ser Trp Leu Pro 135	TCG CCC GGC Ser Pro Gly	CGC CGG CGG CGA (Arg Arg Arg Arg 1	CGC 430 Arg
ACC GCC TTC GCC ATT Ala Phe Ala 8	AGC CGT CAC GGC Ser Arg His Gly 150	AAG CGA CAT Lys Arg His	GGC AAG AAG TCC 7 Gly Lys Lys Ser 7 155	AGG 478 Arg
CTG CGC TGC AGC A Leu Arg Cys Ser A 160	AGA AAG CCT CTG Arg Lys Pro Leu 165	CAC GTG AAT His Val Asn 170	Phe Lys Glu Leu (GGC 526 Gly 175
TGG GAC GAC TGG A	ATT ATC GCG CCC Ile Ile Ala Pro 180	CTA GAG TAC Leu Glu Tyr 185 7/	GAG GCC TAT CAC ! Glu Ala Tyr His (TGC 574 Cys

GAG Glu	GGC Gly	GTG Val	TGC Cys 195	GAC Asp	TTT Phe	CCG Pro	CTG Leu	CGC Arg 200	TCG Ser	CAC His	CTT Leu	GAG Glu	CCC Pro 205	ACT Thr	AAC Asn		622
CAT His	GCC Ala	ATC Ile 210	ATT Ile	CAG Gln	ACG Thr	CTG Leu	ATG Met 215	AAC Asn	TCC Ser	ATG Met	GAC Asp	CCG Pro 220	GGC Gly	TCC Ser	ACC Thr		670
CCG Pro	CCT Pro 225	AGC Ser	TGC Cys	TGC Cys	GTT Val	CCC Pro 230	ACC Thr	AAA Lys	CTG Leu	ACT Thr	CCC Pro 235	ATT Ile	AGC Ser	ATC Ile	CTG Leu		718
TAC Tyr 240	ATC Ile	GAC Asp	GCG Ala	GGC Gly	AAT Asn 245	AAT Asn	GTN X02	GTC Val	TAC Tyr	AAG Lys 250	CAG Gln	TAT Tyr	GAG Glu	GAC Asp	ATG Met 255		766
			TCC Ser						CGGT	GCT (GTCC(CGCC	AC C'	TGGG	CCAGG		820
GAC	CATG	GAG (GGAG	CCT	GA C	TGCC(GAGA	A AG	GAGC	AGGA	GCT	GCC'	TTG (GAAG	AGGCCA		880
CAG	STGG	GGG :	ACAG	CCTG	AA AA	GTAG(GAGC	A CA	GTAA(GAAG	CAG	CCCA	GCC '	TTCC	CAGAAC	:	940
CTT	CCAA'	rcc ·	CCCA	ACCC	AG A	AGCA	GCTA	A GG	GTT'	TCA C	AAC'	TTTT(GGC	CTTG	CCAGCC	: :	1000
TGG	AAAG	ACT .	AGAC:	AAGA	GG G	ATTC'	TTCT	C TT	TTTA'	TAT	GGC'	TTG				:	1046

(2) INFORMATION FOR SEQ ID NO:32:

- (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 263 amino acids(B) TYPE: amino acid(D) TOPOLOGY: linear
- (ii) MOLECULE TYPE: protein
- (xi) SEQUENCE DESCRIPTION: SEQ ID NO:32:

Arg Lys Gln Ala Cys Ile Pro Ala Gly Pro Thr Leu Arg Gly Ser Ser

Gly Thr Gln Pro Arg Pro Ala Gly Lys Ser Phe Asp Val Trp Gln Gly 20 25 30

Leu Arg Pro Gln Pro Trp Lys Gln Leu Cys Leu Glu Leu Arg Ala Ala

Trp Gly Glu Leu Asp X01 Gly Asp Thr Gly Ala Arg Ala Arg Gly Pro 50 60

Gln Gln Pro Pro Pro Leu Asp Leu Arg Ser Leu Gly Phe Gly Arg Arg

Val Arg Pro Pro Gln Glu Arg Ala Leu Leu Val Val Phe Thr Arg Ser

Gln Arg Lys Asn Leu Phe Thr Glu Met His Glu Gln Leu Gly Ser Ala

Glu Ala Ala Gly Ala Glu Gly Ser Cys Pro Ala Pro Ser Gly Ser Pro

Asp Thr Gly Ser Trp Leu Pro Ser Pro Gly Arg Arg Arg Arg Thr

Ala 145	Phe	Ala	Ser	Arg	His 150	Gly	Lys	Arg	His	Gly 155	Lys	Lys	Ser	Arg	Leu 160	
Arg	Cys	Ser	Arg	Lys 165	Pro	Leu	His	Val	Asn 170	Phe	Lys	Glu	Leu	Gly 175	Trp	
qaA	Asp	Trp	Ile 180	Ile	Ala	Pro	Leu	Glu 185	Tyr	Glu	Ala	Tyr	His 190	Cys	Glu	
Gly	Val	Cys 195	Asp	Phe	Pro	Leu	Arg 200	Ser	His	Leu	Glu	Pro 205	Thr	Asn	His	
Ala	Ile 210	Ile	Gln	Thr	Leu	Met 215	Asn	Ser	Met	Asp	Pro 220	Gly	Ser	Thr	Pro	
Pro 225	Ser	Cys	Сув	Val	Pro 230	Thr	Lys	Leu	Thr	Pro 235	Ile	Ser	Ile	Leu	Tyr 240	
Ile	Asp	Ala	Gly	Asn 245	Asn	X02	Val	Tyr	Lys 250	Gln	Tyr	Glu	Asp	Met 255	Val	
Val	Glu	Ser	Cys 260	Gly	Cys	Arg										
(2)	INFO	RMA	rion	FOR	SEQ	ID h	10:33	3:								
	(ii) (iii) (iv) (vii) (ix)	MOI HYI ANT IMM (I	QUENCAL LECUI POTHE POTHE POTHE PATURE ATURE ATURE ATURE ATURE	ENGTH (PE: (PANI) (POLO) ETICH ETICH ENSE: (ATE SE) (ATE S	M: 13 nucl DEDNI DEDNI DETNI D	MAS ALEIC ESS: line DNA NO CE: VAN V	oase acid sing ear (ger	pain i yle nomic								
	4	(E	3) LC	CAT	ON:	990.	.130)1								
			QUENC													
AACI	CATAC	SCA (CTGC	AGT	C CI	rggt	CTTGG	GTC	TAG	GGT	GCGC	CTCC	rgg 1	rccc	SCGGCT	60
CAGO	GATA	ATG (AGTO	BACCA	LA TO	GGT	GTT	G GCC	CTGA	rggg	ACT	TTG	CT 1	rgct?	AACCA	120
AAGO	TCGC	TT (GGAT	1				Arg 1			CTG (Leu I		rp 7			170
			TTC Phe -270	Arg					Arg					Trp		218

CAA Gln	CAG Gln	GCC Ala -255	Trp	CTC Leu	CCA Pro	CAT His	CGA Arg -250	AGA Arg	CAG Gln	CTG Leu	GGC Gly	CAT His -245	Leu	CTG Leu	TTA Leu	266
GGA Gly	GGC Gly -240	Pro	GCG Ala	CTG Leu	ACA Thr	GTG Val -235	Cys	AGG Arg	ATT Ile	TGC Cys	TCT Ser -230	Tyr	ACA Thr	GCT Ala	CTT Leu	314
TCT Ser -225	Leu	TGT Cys	CCC Pro	TGC Cys	CGG Arg -220	Ser	CCC Pro	GCA Ala	GAC Asp	GAA Glu -215	Ser	GCA Ala	GCC Ala	GAA Glu	ACA Thr -210	362
					Phe			TCC Ser		Leu					Glu	410
				Glu				CTG Leu -18	Arg					Glu		458
			Ser					CCG Pro					Ser			506
		Ala					Arg	CTG Leu				Arg				554
	Leu					Trp		GCG Ala			Val					602
					Glu			CCC Pro		Arg					Leu	650
				Ala				CCG Pro -10	Ser					Arg		698
								GGG Gly								746
								ACG Thr								794
								GCG Ala								842
								GGC Gly								890
								ACG Thr -25							ACA Thr	938
								GGC Gly								986
								TTG Leu		Val						1034

				TGG Trp 20											1082
				TGC Cys											[.] 1130
				ATT Ile									-	-	 1178
_				TGC Cys											 1226
				GCC Ala											1274
ATG Met	GTG Val	GTG Val	GAG Glu	GCC Ala 100	TGC Cys	GGC Gly	TGC Cys	AGG Arg	TAGO	CGCGC	GG (CCGC	GGA	S G	1321
GGGC	AGCC	CAC O	CGGC	CCGAC	G AT	rcc									1345

(2) INFORMATION FOR SEQ ID NO:34:

- (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 388 amino acids
 (B) TYPE: amino acid

 - (D) TOPOLOGY: linear
- (ii) MOLECULE TYPE: protein
- (xi) SEQUENCE DESCRIPTION: SEQ ID NO:34:

Pro Gly Arg Arg Pro Leu Leu Trp Ala Arg Leu Ala Ala Phe Arg -275

Leu Gly Gln Arg Arg Gly Val Gly Arg Trp Leu Gln Gln Ala Trp Leu -260

Pro His Arg Arg Gln Leu Gly His Leu Leu Gly Gly Pro Ala Leu

Thr Val Cys Arg Ile Cys Ser Tyr Thr Ala Leu Ser Leu Cys Pro Cys

Arg Ser Pro Ala Asp Glu Ser Ala Ala Glu Thr Gly Gln Ser Phe Leu -220 -215

Phe Asp Val Ser Ser Leu Asn Asp Ala Asp Glu Val Val Gly Ala Glu

Leu Arg Val Leu Arg Arg Gly Ser Pro Glu Ser Gly Pro Gly Ser Trp -180

Thr Ser Pro Pro Leu Leu Leu Ser Thr Cys Pro Gly Ala Ala Arg

Ala Pro Arg Leu Leu Tyr Ser Arg Ala Ala Glu Pro Leu Val Gly Gln

Arg Trp Glu Ala Phe Asp Val Ala Asp Ala Met Arg Arg His Arg Arg -135 -130 75-

Glu Pro Arg Pro Pro Arg Ala Phe Cys Leu Leu Leu Arg Ala Val Ala

Gly Pro Val Pro Ser Pro Leu Ala Leu Arg Arg Leu Gly Phe Gly Trp

Pro Gly Gly Gly Ser Ala Ala Glu Glu Arg Ala Val Leu Val Val

Ser Ser Arg Thr Gln Arg Lys Glu Ser Leu Phe Arg Glu Ile Arg Ala

Gln Ala Arg Ala Leu Gly Ala Ala Leu Ala Ser Glu Pro Leu Pro Asp

Pro Gly Thr Gly Thr Ala Ser Pro Arg Ala Val Ile Gly Gly Arg Arg

Arg Arg Arg Thr Ala Leu Ala Gly Thr Arg Thr Ala Gln Gly Ser Gly

Gly Gly Ala Gly Arg Gly His Gly Arg Gly Arg Ser Arg Cys Ser
-10 -5 1

Arg Lys Pro Leu His Val Asp Phe Lys Glu Leu Gly Trp Asp Asp Trp

Ile Ile Ala Pro Leu Asp Tyr Glu Ala Tyr His Cys Glu Gly Leu Cys

Asp Phe Pro Leu Arg Ser His Leu Glu Pro Thr Asn His Ala Ile Ile

Gln Thr Leu Leu Asn Ser Met Ala Pro Asp Ala Ala Pro Ala Ser Cys

Cys Val Pro Ala Arg Leu Ser Pro Ile Ser Ile Leu Tyr Ile Asp Ala

Ala Asn Asn Val Val Tyr Lys Gln Tyr Glu Asp Met Val Val Glu Ala

Cys Gly Cys Arg

(2) INFORMATION FOR SEQ ID NO:35:

- (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 17 base pairs

 - (B) TYPE: nucleic acid (C) STRANDEDNESS: single (D) TOPOLOGY: linear
- (ii) MOLECULE TYPE: DNA (genomic)
- (iii) HYPOTHETICAL: NO
- (iv) ANTI-SENSE: NO
- (vi) ORIGINAL SOURCE: (C) INDIVIDUAL ISOLATE: primer number 8
- (xi) SEQUENCE DESCRIPTION: SEQ ID NO:35:

TGTATGCGAC TTCCCGC

What is claimed is:

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1.

1. A DNA molecule comprising an isolated DNA sequence encoding a BMP-12 related protein.

- 5 2. A DNA molecule according to claim 1, wherein said DNA sequence is selected from the group consisting of:
 - (a) nucleotides #496, #571 or #577 to #882 of SEQ ID NO:1;
 - (b) nucleotides #605 or #659 to #964 of SEQ ID NO:25; and
- (c) sequences which hybridize to (a) or (b) under stringent hybridization conditions and encode a BMP-12 related protein which exhibits the ability to form tendon/ligament-like tissue.
 - 3. A DNA molecule comprising the DNA sequence of claim 1 wherein said DNA sequence is selected from the group consisting of:
- (a) nucleotides encoding for amino acids #-25, #1 or #3 to #104 of SEQ ID NO:2;
 - (b) in a 5' to 3' direction, nucleotides encoding a propeptide selected from the group consisting of native BMP-12 propeptide and a BMP protein propeptide; and nucleotides encoding for amino acids #-25, #1 or #3 to #104 of SEQ ID NO:2; and
- 20 (c) nucleotides encoding for amino acids #1 or #19 to #120 of SEQ ID NO:26;
 - (d) in a 5' to 3' direction, nucleotides encoding a propeptide selected from the group consisting of native BMP-12 propeptide and a BMP protein propeptide; and nucleotides encoding for amino acids #1 or #19 to #120 of SEQ ID NO:26:
 - (e) sequences which hybridize to any of (a) through (d) under stringent hybridization conditions and encode a BMP-12 related protein which exhibits the ability to form cartilage and/or bone.
 - 4. A host cell transformed with a DNA molecule according to claim
 - 5. A host cell transformed with the DNA molecule of claim 2.
 - 6. A host cell transformed with the DNA molecule of claim 3.

7. An isolated DNA molecule having a sequence encoding a BMP-12 protein which is characterized by the ability to induce the formation of tendon/ligament-like tissue, said DNA molecule comprising a DNA sequence selected from the group consisting of:

- 5
- (a) nucleotide #496, #571 or #577 to #882 of SEQ ID NO:1;
- (b) nucleotide #605 or #659 to #964 of SEQ ID NO:25; and
- (c) naturally occurring allelic sequences and equivalent degenerative codon sequences of (a) or (b).
 - 8. A host cell transformed with the DNA molecule of claim 7.
- 10
- 9. A vector comprising a DNA molecule of claim 7 in operative association with an expression control sequence therefor.
 - 10. A host cell transformed with the vector of claim 9.
- 11. A method for producing a purified BMP-12 protein, said method comprising the steps of:
- (a) culturing a host cell transformed with a DNA molecule according to claim 2, comprising a nucleotide sequence encoding a BMP-12 related protein; and
 - (b) recovering and purifying said BMP-12 related protein from the culture medium.
- 20 12. A method for producing a purified BMP-12 related protein said method comprising the steps of:
 - (a) culturing a host cell transformed with a DNA molecule according to claim 3, comprising a nucleotide sequence encoding a BMP-12 related protein; and
- 25 (b) recovering and purifying said BMP-12 related protein from the culture medium.
 - 13. A method for producing a purified BMP-12 related protein said method comprising the steps of:
- (a) culturing a host cell transformed with a DNA molecule according to
 claim 7, comprising a nucleotide sequence encoding a BMP-12 related protein;
 and

(b) recovering and purifying said BMP-12 related protein from the culture medium.

- 14. A purified polypeptide comprising an amino acid sequence selected from the following group:
- 5 (a) from amino acid #-25 to amino acid #104 as set forth in SEQ ID NO:2;
 - (b) from amino acid #1 to amino acid #104 as set forth in SEQ ID NO:2.
 - (c) from amino acid #3 to amino acid #104 as set forth in SEQ ID NO:2.
- (d) from amino acid #1 to amino acid #120 as set forth in SEQ ID NO:26; and
 - (d) from amino acid #19 to amino acid #120 as set forth in SEQ ID NO:26.
 - 15. A purified polypeptide wherein said polypeptide is in the form of a dimer comprised of two subunits, each with the amino acid sequence of claim 14.
 - 16. A purified protein produced by the steps of

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- (a) culturing a cell transformed with a DNA molecule comprising the nucleotide sequence from nucleotide #496, #571 or #577 to #882 as shown in SEQ ID NO:1; and
- 20 (b) recovering and purifying from said culture medium a protein comprising the amino acid sequence from amino acid #-25, amino acid #1 or amino acid #3 to amino acid #104 as shown in SEQ ID NO:2.
 - 17. A purified BMP-12 related protein characterized by the ability to induce the formation of tendon/ligament-like tissue.
- 25 18. A pharmaceutical composition comprising an effective amount of the BMP-12 related protein of claim 17 in admixture with a pharmaceutically acceptable vehicle.
 - 19. A method for inducing tendon/ligament-like tissue formation in a patient in need of same comprising administering to said patient an effective amount of the composition of claim 18.

20. A pharmaceutical composition for tendon/ligament-like tissue healing and tissue repair said composition comprising an effective amount of the protein of a BMP-12 related protein in a pharmaceutically acceptable vehicle.

- 21. A method for treating tendinitis, or other tendon or ligament defect in a patient in need of same, said method comprising administering to said patient an effective amount of the composition of claim 20.
- 22. A chimeric DNA molecule comprising a DNA sequence encoding a propertide from a member of the TGF- β superfamily of proteins linked in correct reading frame to a DNA sequence encoding a BMP-12 related polypeptide.
- 23. A chimeric DNA molecule according to claim 22, wherein the propeptide is the propeptide from BMP-2.
- 24. A heterodimeric protein molecule comprising one monomer having the amino acid sequence of the polypeptide of claim 14, and one monomer having the amino acid sequence of a protein of the TGF- β superfamily.
- 25. A method for inducing tendon/ligament-like tissue formation in a patient in need of same comprising administering to said patient an effective amount of a composition comprising a protein encoded by a DNA sequence selected from the group consisting of:

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- (a) nucleotides #496, #571 or #577 to #882 of SEQ ID NO:1;
- (b) nucleotides #845 or #899 to #1204 of SEQ ID NO:3;
- (c) nucleotides #605 or #659 to #964 of SEQ ID NO:25; and
- (d) sequences which hybridize to (a), (b) or (c) under stringent hybridization conditions and encode a protein which exhibits the ability to form tendon/ligament-like tissue.

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26. A method for inducing tendon/ligament-like tissue formation in a patient in need of same comprising administering to said patient an effective amount of the composition comprising a tendon/ligament-like tissue inducing protein having an amino acid sequence selected from the group consisting of:

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- (a) amino acids #-25, #1 or #3 to #104 of SEQ ID NO:2:
- (b) amino acids #1 or #19 to #120 of SEQ ID NO:4;
- (c) amino acids #1 or #19 to #120 of SEQ ID NO:26; and

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(d) mutants and/or variants of (a), (b) or (c) which exhibit the ability to form tendon and/or ligament.

- 27. A pharmaceutical composition for tendon/ligament-like tissue repair, said composition comprising an effective amount of a BMP-12 related protein in a pharmaceutically acceptable vehicle.
- 28. A method for treating tendinitis, or other tendon or ligament defect in a patient in need of same, said method comprising administering to said patient an effective amount of the composition of claim 27.

FIG 1/1
COMPARISON OF HUMAN V1-1 VS. HUMAN MP-52

```
V1-1 Ser Arg Cys Ser Arg Lys Pro Leu His Val Asp Phe Lys Glu Leu
1 AGC CGC TGC AGC CGC AAG CCG TTG CAC GTG GAC TTC AAG GAG CTC
MP52 GCT CGC TGC AGT CGG AAG GCA CTG CAT GTC AAC TTC AAG GAC ATG
 1 Ala Arg Cys Ser Arg Lys Ala Leu His Val Asn Phe Lys Asp Met
 16 Gly Trp Asp Asp Trp Ile Ile Ala Pro Leu Asp Tyr Glu Ala Tyr
     GGC TGG GAC GAC TGG ATC ATC GCG CCG CTG GAC TAC GAG GCG TAC
     GGC TGG GAC GAC TGG ATC ATC GCA CCC CTT GAG TAC GAG GCT TTC
   Gly Trp Asp Asp Trp Ile Ile Ala Pro Leu Glu Tyr Glu Ala Phe
     His Cys Glu Gly Leu Cys Asp Phe Pro Leu Arg Ser His Leu Glu CAC TGC GAG GGC CTT TGC GAC TTC CCT TTG CGT TCG CAC CTC GAG
     CAC TGC GAG GGG CTG TGC GAG TTC CCA TTG CGC TCC CAC CTG GAG
 31 His Cys Glu Gly Leu Cys Glu Phe Pro Leu Arg Ser His Leu Glu
 46
     Pro Thr Asn His Ala Ile Ile Gln Thr Leu Leu Asn Ser Met Ala
     Pro Thr Asn His Ala Val Ile Gln Thr Leu Met Asn Ser Met Asp
     Pro Asp Ala Ala Pro Ala Ser Cys Cys Val Pro Ala Arg Leu Ser
     CCA GAC GCG GCG CCG GCC TCC TGC TGT GTG CCA GCG CGC CTC AGC
     CCC GAG TCC ACA CCC ACC TGC TGT GTG CCC ACG CGG CTG AGT
61 Pro Glu Ser Thr Pro Pro Thr Cys Cys Val Pro Thr Arg Leu Ser
     Pro Ile Ser Ile Leu Tyr Ile Asp Ala Ala Asn Asn Val Val Tyr CCC ATC AGC ATC CTC TAC ATC GAC GCC GCC AAC AAC GTT GTC TAC
226
     CCC ATC AGC ATC CTC TTC ATT GAC TCT GCC AAC AAC GTG GTG TAT
226
     Pro Ile Ser Ile Leu Phe Ile Asp Ser Ala Asn Asn Val Val Tyr
91 Lys Gln Tyr Glu Asp Met Val Val Glu Ser Cys Gly Cys Arq
```

Homology at the nucleotide level: 249/312 = 79.8\$ Homology at the amino acid level: 84/104 = 80.8\$

INTERNATIONAL SEARCH REPORT

Inter/ nal Application No PCT/US 94/14030

		PC	1/03 34/14030
A. CLASS IPC 6	IFICATION OF SUBJECT MATTER C12N15/12 C12N15/70 C12N15 A61K38/17	/62 C12N1/21	C07K14/51
According t	to International Patent Classification (IPC) or to both national cla	assification and IPC	
B. FIELD	S SEARCHED		
Minimum d IPC 6	documentation searched (classification system followed by classifi C12N C07K A61K	cation symbols)	
	tion searched other than minimum documentation to the extent th		
Electronic o	lata base consulted during the international search (name of data	base and, where practical, search	terms used)
C. DOCUM	IENTS CONSIDERED TO BE RELEVANT		
Category *	Citation of document, with indication, where appropriate, of the	: relevant passages	Relevant to claim No.
X	WO,A,93 16099 (BIOPHARM GESSELLS BIOTECHNOLOGISCHEN ENTWICKLUNG S PHARMAKA) 19 August 1993 cited in the application see page 4, paragraph 3 see page 7, paragraph 3 see page 9, paragraph 2	SCHAFT ZUR VON	25,26
A	WO,A,91 18047 (GENENTECH, INC.) November 1991 cited in the application see page 4, line 4 - line 22	28	22,23
A	WO,A,93 00432 (GENETICS INSTITUT January 1993 cited in the application see page 4, line 21 - line 33 see page 8, line 15 - page 10, 1		1-28
Furth	er documents are listed in the continuation of box C.	X Patent family member	rs are listed in annex.
'A' documer which is citation other m'P' documer later the	nt which may throw doubts on priority claim(s) or s cited to establish the publication date of another or other special reason (as specified) nt referring to an oral disclosure, use, exhibition or	or priority date and not in cited to understand the prinvention "X" document of particular rel cannot be considered now involve an inventive step "Y" document of particular rel cannot be considered to in document is combined wi ments, such combination in the art. "&" document member of the interest of mailing of the interest.	mational search report
19	May 1995	24.	(ô. C.
Name and m	ailing address of the ISA European Patent Office, P.B. 5818 Patentiaan 2 NL - 2280 HV Rijswijk Tel. (+ 31-70) 340-2040, Tx. 31 651 epo nl, Fax (+ 31-70) 340-3016	Authorized officer Montero Lop	pez. B

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ernational application No.

INTERNATIONAL SEARCH REPORT

PCT/US 94/ 14030

Box 1	Observations where certain claims were found unsearchable (Continuation of item 1 of first sheet)
This inte	ernational search report has not been established in respect of certain claims under Article 17(2)(a) for the following reasons:
1. X	Claims Nos.: because they relate to subject matter not required to be searched by this Authority, namely: Remark: Although claims 19,21,25,26 and 28 are directed to a mehtod of treatment of the human/animal body the search has been carried out and based on the alleged effects of the composition.
2.	Claims Nos.: because they relate to parts of the international application that do not comply with the prescribed requirements to such an extent that no meaningful international search can be carried out, specifically:
3.	Claims Nos.: because they are dependent claims and are not drafted in accordance with the second and third sentences of Rule 6.4(a).
Box II	Observations where unity of invention is lacking (Continuation of item 2 of first sheet)
This Int	ernational Searching Authority found multiple inventions in this international application, as follows:
1.	As all required additional search fees were timely paid by the applicant, this international search report covers all searchable claims.
2.	As all searchable claims could be searches without effort justifying an additional fee, this Authority did not invite payment of any additional fee.
3.	As only some of the required additional search fees were timely paid by the applicant, this international search report covers only those claims for which fees were paid, specifically claims Nos.:
4.	No required additional search fees were timely paid by the applicant. Consequently, this international search report is restricted to the invention first mentioned in the claims; it is covered by claims Nos.:
Remark	The additional search fees were accompanied by the applicant's protest. No protest accompanied the payment of additional search fees.

INTERNATIONAL SEARCH REPORT

information on patent family members

Inter 2al Application No
PCT/US 94/14030

Patent document cited in search report	Publication date	Patent fi membe		Publication date
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Form PCT/ISA/210 (patent family annex) (July 1992)